بسم الله الرحمن الرحيم

Sudan University of Science and Technology College of Graduate Studies

Immunotoxicity of Convolvulus arvensis
(Binweed)
in
Sheep and Rats
(دراسة السمية المناعية لنبات الكونفلفيولس أرفينسس (البنويد

By: Mohammed Eissa Ahmed Al-Bowait BVSc (1990), MVSc (2000)

A thesis submitted in accordance with the requirements for PhD degree in Toxicology

Supervisors:

Intrnal: Dr.Seif Eldawla Mustafa Barakat External: Professor Abdel Gadir Musa Homeida

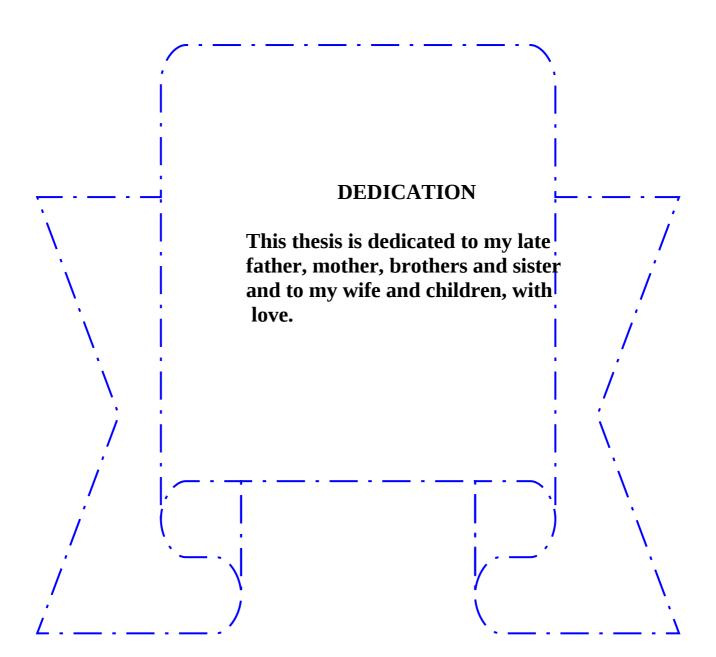
July 2007

TABLE OF CONTENTS

Table of contents Dedication Acknowledgements V Arabic Summary Finglish Summary (Abstract) List of figures List of figures List of tables XIII INTRODUCTION LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2 Collection of blood samples 2.3 Chemicals and drugs 2.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.5 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32 3. RESULTS	ITEM	PAGE No.
Acknowledgements V Arabic Summary VII English Summary (Abstract) List of figures XI List of figures XII List of tables XIIII INTRODUCTION 1 LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 8 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2 Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 2.5 Toxicity of binweed to sheep 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	Table of contents	II
Arabic Summary VII English Summary (Abstract) List of figures XI List of tables XIII INTRODUCTION 1 LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2 Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 2.5 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	Dedication	IV
English Summary (Abstract) List of figures XI List of tables XIIII INTRODUCTION 1 LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 8 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2 Collection of blood samples 2.3 Chemicals and drugs 1.9 2.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.5 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	Acknowledgements	V
List of figures XI List of tables XIIII INTRODUCTION 1 LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 8 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 15 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2 Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 25 2.7 Effects of binweed extracts on phagocytic function of macrophages 27 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 29 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 32 2.11 Statistical analysis 33	Arabic Summary	VII
List of tables XIII INTRODUCTION 1 LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 8 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 15 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2 Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 25 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 29 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 32	English Summary (Abstract)	IX
INTRODUCTION LITERATURE REVIEW 3 1.1 Primary toxins 6 1.2 Secondary toxins 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2 Collection of blood samples 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 3	<u> </u>	XI
LITERATURE REVIEW 1.1 Primary toxins 6 1.2 Secondary toxins 8 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2Collection of blood samples 1.9 2.3 Chemicals and drugs 1.9 2.4 Collection of plant material 2.0 2.5 Toxicity of binweed to sheep 2.2 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32		XIII
1.1 Primary toxins 1.2 Secondary toxins 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2 Collection of blood samples 1.9 2.3 Chemicals and drugs 1.9 2.4 Collection of plant material 2.0 2.5 Toxicity of binweed to sheep 2.2 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	INTRODUCTION	1
1.2 Secondary toxins 1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2Collection of blood samples 1.9 2.3 Chemicals and drugs 1.9 2.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	LITERATURE REVIEW	
1.3 Activation of the immune response and immune effector functions 1.4 Field bindweed 15 2. MATERIALS AND METHODS 19 2.1 Animals and housing 19 2.2Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	1.1 Primary toxins	6
effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.9 2.1 Animals and housing 1.9 2.2 Collection of blood samples 1.9 2.3 Chemicals and drugs 1.9 2.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	1.2 Secondary toxins	8
effector functions 1.4 Field bindweed 1.5 2. MATERIALS AND METHODS 1.1 Animals and housing 2.2 Collection of blood samples 1.2 Collection of blood samples 1.3 Chemicals and drugs 1.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	1.3 Activation of the immune response and immune	10
2. MATERIALS AND METHODS192.1 Animals and housing192.2Collection of blood samples192.3 Chemicals and drugs192.4 Collection of plant material202.5 Toxicity of binweed to sheep222.6 Toxicity of binweed to rats252.7 Effects of binweed extracts on phagocytic function of macrophages272.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations282.9 Antidiarrheal effects of binweed extracts292.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats312.11 Statistical analysis32	effector functions	13
2.1 Animals and housing 2.2 Collection of blood samples 19 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 25 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	1.4 Field bindweed	15
2.2 Collection of blood samples 2.3 Chemicals and drugs 19 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 25 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2. MATERIALS AND METHODS	19
2.3 Chemicals and drugs 2.4 Collection of plant material 20 2.5 Toxicity of binweed to sheep 22 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.1 Animals and housing	19
2.4 Collection of plant material 2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.2Collection of blood samples	19
2.5 Toxicity of binweed to sheep 2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.3 Chemicals and drugs	19
2.6 Toxicity of binweed to rats 2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 25 27 28 31 31 32	2.4 Collection of plant material	20
2.7 Effects of binweed extracts on phagocytic function of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.5 Toxicity of binweed to sheep	22
of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 27 28 29 31 31 32	2.6 Toxicity of binweed to rats	25
of macrophages 2.8 Effects of binweed extracts on serum lysozome activity, neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.7 Effects of binweed extracts on phagocytic function	27
neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	of macrophages	2/
neopterin, leptin and immunoglobulins concentrations 2.9 Antidiarrheal effects of binweed extracts 2.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats 2.11 Statistical analysis 32	2.8 Effects of binweed extracts on serum lysozome activity,	20
2.9 Antidiarrheal effects of binweed extracts292.10 Effects of binweed on the activity of drug metabolizing enzymes in liver of rats312.11 Statistical analysis32	neopterin, leptin and immunoglobulins concentrations	28
enzymes in liver of rats 2.11 Statistical analysis 31 32		29
enzymes in liver of rats 2.11 Statistical analysis 31 32	2.10 Effects of binweed on the activity of drug metabolizing	
2.11 Statistical analysis 32		31
		32
	3. RESULTS	34

ITEM	PAGE No.
3.1 Clinical findings	34
3.2 Post – mortem findings	36
3.3 Histological findings	39
3.4 Changes in serum constituents	40
3.5 Hematological changes	53
3.6 Extraction yield	56
3.7 LD ₅₀ of extracts	56
3.8 Effects of Extracts on phagocytic function	59
3.9 Effects of extracts on other immune parameters	63
3.10 Antispasmodic Effect of extracts	65

4. DISCUSSION	70
5. REFERENCES	81



ACKNOWLEGEMENTS

This work has been carried out jointly by Sudan University for Science and Technology, Khartoum and King Faisal University (KFU), Al-ahsa, Saudi Arabia.

I am very grateful to the internal supervisor professor SeifEldawla Mustafa Barakat for his support, valuable suggestions and guidance.

I am indebted to my external supervisor professor Abdelgadir Musa Homeida for his close supervision, constructive criticism, encouragement and the confidence he bestowed upon me throughout the years to complete this thesis and for being a great friend as well as excellent advisor.

Special thanks to professor Mohammed Al-shiek Barri for fruitful discussion of experimental toxicity, professor Al-Mahi Bilal Abdelsalam for his valuable help with histopathology.

I wish to express my gratitude to previous dean of College of Veterinary Medicine, Al-Ahsa, Dr. Abdullah Al-Dughym, the current dean Dr. Abderhman Al-Ankari and the deputy vice dean for postgraduate studies Dr. Mohammed Al-Nazawi for their continual support.

My gratitude is also for Professor Yahia Hussian, Dr. Khalid Al-Busadah and Dr. Fahad Al-Hazab for their valuable suggestions, Dr. Mohammed Al-Fredan, College of Science (KFU) for identification of the plant.

Thanks are due to Ahmed Al-Subaie, Abdullah Al-botayan,
Abdalaziz Al-Amir, Kefah Al-Khars, Yousif Al-Hamdan and
Mohammed Al-Zair for excellent technical assistance and support and
for Ahmed Al-Ibrahim, Mohammed Al-Sinahir and Saleh Al-Ashgir for
typing the thesis.

I am very grateful to the Dean ship of Scientific Research (KFU) for financial support of this work.

I wish to give my family the biggest acknowledgment. My wife, Om Eisa, always supported me. I had to leave her alone with kids. Thanks to her and to my sons Eisa and Abdullah and daughters Nada and Nouf for love and support they gave me.

الملخص بالعربي

يعتبر نبات البنويد (الفضاخ) من النباتات الزاحفة التي تنتشر في الشرق الأوسط. وربما احتاجت الحيوانات للأكل منه في أوقات الجفاف. لذا كان من المفيد اجراء دراسة عن سمية النبات في الأغنام والفئران.

في البداية تم تقسيم الأغنام والفتران إلى ثلاثة مجموعات. تم استخدام مجموعة كشاهد وتم استخدام المجموعتين الأخريين في التجربة حيث تم اطعام إحداهما بنبات الفضاخ وحده وتم اطعام المجموعة الأخرى بنبات الفضاخ مخلوط بنسبة 50 % مع نبات الرودس. لقد تم ملاحظة ان الحيوانات نفقت او تسممت بعد مضي 7 أيام من إطعام الفضاخ. واشتملت الأعراض الاكلينيكية على جحوظ العين وبهتان الغشاء المخاطي وزيادة في سرعة التنفس وترنح وتشنجات واسهال. ولقد اشتملت التغيرات الظاهرية والمجهرية على تراكم الغازات في الأمعاء ،احتقان و ودمة ونزف لأعضاء داخلية كثيرة.

إن انخفاض تركيز الهيموجلوبين وبنسبة خلايا الدم المتراصة وعدد كريات الدم الحمراء دل على حدوث أنيميا في الأغنام. لم تلاحظ أي أعراض على الأغنام التي تم إطعامها بالفضاخ مخلوط مع الرودس. وفي تجربة أخرى تم ملاحظة إن إطعام الفتران بالفضاخ في جرعة مقدارها 0.5 جم ولمدة 10 أيام تسبب في نقصان في تركيز البروتين ونشاط إنزيمات بعض أيض الأدوية والسموم في الكبد. لقد تم الحصول على مستخلص كحولي وآخر مائي لنبات الفضاخ حيث تبين أن الجرعة السمية الوسطية للمستخلص المكحولي في الفئران هي 100 ± 0.5 ملجرام للكيلو جرام وإن الجرعة السمية للمستخلص المائي هي 100 ± 0.5 ملجرام للكيلو جرام مما يقود إلى الاستنتاج بأن سمية الفضاخ في الأغنام والفئران ربما نتجت من سمية المستخلص الكحولي . لقد تم المستخلص الكحولي للفضاخ بجرعة مقدارها 1.0 من المستخلص الكحولي للفضاخ بجرعة مقدارها 1.0 من المستخلص الكحولي الفضاغ بجرعة مقدارها 1.0 من الجرعة السمية الوسطية مثلما الجرعة السمية الوسطية في الغشاء التنوري للفئران قد تسبب في زيادة كريات الدم الحمراء ونسبة الخلايا الليمفاوية وتنشيط وظيفة البلعمة من خلايا الشبكة الاندثولية وإيقاف التثبيط على المناعة بواسطة الديكساميثازون . لقد تسبب حقن المستخلص المستخلص المتخلص المواقية البلعمة من خلايا الشبكة الاندثولية وإيقاف التثبيط على المناعة بواسطة الديكساميثازون . لقد تسبب حقن المستخلص وإيقاف التثبيط على المناعة بواسطة الديكساميثازون . لقد تسبب حقن المستخلص

المائي على زيادة بعض وسائط المناعة مثل تركيز هرمون اللبتين ونيوبترين والأمينوجلوبولين ونشاط انزيم اليسوزوم.

تشير هذه النتائج إلى أن أوراق نبات الفضاخ تحتوي على مستخلص كحولي شبيه بالأتروبين وهو سام للحيوانات ومستخلص مائي يزيد من وظيفة المناعة. وإن النبات يعتبر سام للحيوانات لذا يجب منع الحيوانات من أكله.

ABSTRACT

Convolvulus arvensis (Binweed) is a creeping weed widely distributed in the Middle East. During drought animals may be forced to consume the plant. Therefore, this study was carried out to investigate the toxicity of the plant in sheep and rats.

Initially sheep and rats were divided into 3 groups. One group served as controls and fed Rhodes in case of sheep and rat diet in case of rats. The other groups were fed either exclusively on binweed leaves or on 50 % binweed combined with Rhodes or rat diet. Sheep or rats either died or intoxicated by the plant 7 days post dosing with clinical signs included dilated pupil, pale mucous membrane, respiratory distress, ataxia, convulsion and diarrhea. Macroscopic and microscopic lesions included gaseous distended intestine, congestion, oedema and hemorrhage of many organs. Decreased Hb, PCV and RBC count were suggestive of anemia. Elevation of enzymes indicating liver and kidney dysfunction also occurred.

Animals fed 50 % binweed did not develop any signs of toxicity. Oral administration at a dose of 0.5 g rat of binweed for 10 days caused decreased protein in liver homogenate and inhibited the activity of phase-1 drug metabolizing enzymes. Successful extraction of the plant yielded alcoholic and aqueous fractions with LD50 of 160 $\pm~5$ mg / kg and 410 $\pm~6$ mg / kg , respectively, suggesting that the binweed toxicity occurring in sheep and rats may possibly be due to alcoholic fraction. Oral administration of 1/10 LD50 of alcoholic extract blocked diarrhea, enteropooling and intestinal transits induced by castor oil, comparable to that of atropine. Intraperitoneal injection of 1/10 LD50 of aqueous extract to rats significantly increased total leukocytes and percentage lymphocyte, enhanced the phagocytic function of reticular endothelial system and blocked immunosuppressive effect produced by

dexamethasone. Furthermore, the aqueous extract significantly increased the concentration of some immunomodulators such as leptin, neopterin, immunoglobulins and lysosmal enzyme activity.

These results show that binweed leaves contain alcoholic soluble atropine-like fraction that was toxic to animals and water soluble fraction that was immunostimulant. The whole plant is toxic to animals, therefore care should be taken that animals are not allowed to eat the plant.

LIST OF FIGURES

ITEM	DESCRIPTION	PAGE No.
Fig (1)	Convolvulus arvensis (Binweed)	17
Fig (2)	General weakness, dullness, and diarrhea in sheep of group 3, fed <i>C. arvensis</i>	35
Fig (3)	Lung: Diffuse pulmonary oedema and congestion. The alveoli are flooded with oedema fluid which appears as homogeneous pinkish material. Note remarkable distension of alveolar capillaries with large numbers of erythrocytes, group 3 sheep (H & E x 120)	41
Fig (4)	Liver: Acute hepatic congestion. The central veins, portal veins and hepatic sinusoids are fully distended with erythrocytes. Note mild vacuolation of hepatocytes, group 3 sheep (H & E x 120)	42
Fig 5	Spleen: Severe congestion of the splenic parenchyma, group 3 sheep (H & E x 120)	43
Fig(6)	Brain: Meningial haemorrhage. Also note perivascular oedema, group 3 sheep (H & E x 120)	44
	Kidney: Massive haemorrhage, congestion and	
Fig(7)	degeneration of renal tubular epithelium, group 3	45
Fig(8)	sheep (H & E x 120) Kidney: Dilatation of renal tubules, group 3 sheep (H & E x 120)	46
Fig(9)	Small intestine: Catarrhal enteritis. Note massive infiltration of inflammatory cells (mainly lymphocytes and macrophages) on the intestinal mucosa with remarkable desquamation of the lining epithelium, group 3 sheep (H & E x 120)	47
Fig(10)	Heart: Capillary congestion and mild degeneration of cardiac muscles, group 3 sheep (H & E x 120)	48
Fig(11)	Lung: Focal haemorrhages inside alveoli. Note the presence of emphysema and lymphoid hyperplasia in group III rats (H & E X 150).	49
ITEM	DESCRIPTION	PAGE No.
Fig(12)	Liver:-Acute hepatic congestion. Note blood vessels and sinusoids are filled with large numbers of red blood cells in group III rats (H & E X 150).	50
Fig(13)	Focal haemorrhage and congestion of glomerular tuft with degeneration and necrosis of renal tubular	51

	epithelium in group III rats (H & E X 150).	
	Spleen: Acute congestion with mild deposition of	
Fig(14)	haemosiderin pigmen in group III rats (H & E X	52
	150)	
Fig (15)	Determination of LD ₅₀ of binweed extracts in rats	58
Eig (16)	Effect of binweed extracts on carbon clearance by	60
Fig (16)	reticular endothelial system in rats	60

LIST OF TABLES

ITEM	DESCRIPTION	PAGE No.
Table 1	Several plant species that contain toxins	3
Table 2	Details of the Awasi sheep fed Convolvulus	22
	arvensis	23
Table 3	Details of the rats fed with Convolvulus arvensis	26
Table 4	Summary of necropsy findings in sheep fed	37
Table 4	C.arvensis	3/
m 11 =	Summary of postmortem findings in rats given <i>C</i> .	20
Table 5	arvensis	38
_ 11 6	Biochemical changes in the serum of sheep fed	
Table 6	Convolvulus arvensis	54
	Haematological values in sheep fed with	
Table 7	Convolvulus arvensis for one week	55
Table 8	Determination of LD ₅₀ of binweed extracts in rats	57
Tubic 0	Effect of binweed extracts on phagocytic function	37
Table 9	and spleen weight in dexamethasone treated rats	62
	Effect of binweed extracts on concentration of	
T 11 40		
Table10	leptin, neopterin, immunoglobulins and activity of	64
	lysosomal enzymes	
Table 11	Effect of binweed extracts on castor oil induced	66
	diarrhea in rats	00
Table 12	Effect of binweed extracts on castor oil induced	0.0
	enteropooling in rats	66
	Effect of binweed extracts on castor oil induced	
Table 13	small intestine transits in rats	67
	Mean ± SD concentration of protein and values of	
m 11 44	•	60
Table 14	activity of drug metabolizing enzymes in	68
	microsomal protein homogenate of liver of rats	