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Bovine Babesiosis in Elfasher Locality - North Darfur State

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ABSTRACT

This study was conducted in September 2017 to March 2018 in Elfasher Locality - North Darfur State to identify Babesia species and ticks infected cattle and to determine some hematological and biochemical parameters associated with these species. A total of 227 whole blood samples were collected from cattle randomly in Elfasher Locality for identification of Babesia parasites using Gemisa stain. Packed cell volume was determined in 100 samples by using microhematocrit technique. Biochemical parameters [Total protein, albumin, bilirubin, calcium, sodium and potassium] were determined in 100 serum samples. One hundred and fifty five tick samples were collected from 23 cattle for identification. The result revealed that 133 smear samples were positive to Babesia species with overall prevalence rate 58.5%. A significant decrease of packed all volume [p<0.01]. All biochemical parameters showed no statistical significant difference, but serum calcium showed significant difference [p<0.01]. There were 2 genera of ticks were identified and these were Ripicephalus and Hyalomma, included 7 species and these were Ripicephalus evensi eversi [37.5%], Hyalomma anatolicum [21.9%], H.impletatum [16.2%], *H.dromedarii* [12.9%], *H. rufipes* [1.2%], *H. truncatumim* [9.0%] and *H.predixtatus* [1.2%]. In conclusion, bovine babesiosis is the most important tick – borne disease of cattle in Elfasher locality leading to death of the animals due to anemia.

Introduction:

Bovine babesiosis considered as important arthropod-borne disease of cattle in wide regions in the world causing a significant morbidity and mortality rates. The major economic impact of the disease in cattle is cattle industry (Bock *et al.*, 2004; Enbiyale *et al.*, 2018). Also costs resulting from babesiosis are not only from morability, ill-thirft, abortions, loss of milk and meat production, draft power, and control measures such as acricide treatment, purchase vaccines and therapeutics, but also through its impact in international cattle trade (Bock *et al.*, 2004). Now a day babesiosis affecting many species of mammals with a major on cattle and man [Schorn *et al.*,

2011; Hamsho et al., 2015]. This disease is tick intra-erythrocytic apicomplexan parasites distributed in cattle in tropical and subtropical regions and the major species are Babesia bovis, B. bigemina, B. divergens, B. major, B. ovate, B. occultans and B. Jakiovi [Sharma et al., 2000; Criado Fornelio et al., 2003; Bock et al., 2004; Spickler et al., 2013]. Implications of babesiosis as zoonotic disease is reported and B. divergens infected cattle population and Ioxides ricinus infested areas. This species causing human babesiosis in Europe and North America resulting in In United States human babesiosis caused by Babesia microti and this parasite transmitted to the human by contact deer ticks (Radostits et al., 2007). During penetration and development of Babesia spp. pharmacological role that resembles a hemolytic anemia [B. bigemina] and kallikren [Kinin] produces in the plasma which increase vascular permability and vasodilation occurs leading to circulatory stasis and shock. Also due to destruction of erythrocytes packed cell volume is decreased. This substance causing changes of coagulation parameters and excessive loss red blood cells by absorption of circulating antigen-antibody complexes to the surface of red blood cells and removed by phagositosis. Severe anemia can be developed due to hypocomplementemia and gromular deposits [gromulonephritis] of 1g and C3 [Soulsby, 1982].

The main vectors of Babesia species in cattle are Boophilus microphus, B. decoloratus, B. annulatus, B. geigyi, Rhipicephalus evertsi, Ixodes ricinus; I. persulcatus, Haemaphysalis punctuate and H. longicoronis [Bock et al., 2004]. The infection of this hemotropic disease is diagnosed by high fever [40-41°c], weight loss, ruminal atony, dyspnea, hymoglobin urea (red water urine), lethargy, diarrhea, hemolytic anemia [The mucous membranes are first hypremic but later they become ictric and pallor], and tachycardia. In chronic infection there is no Acute B. bovis develops fatal cerebral babesiosis with hyperaesthesia, hemoglobinuria. convulsion and paralysis due to aggregation of red blood cells in the cerebral capillaries and extravascular following endothelial damage [Figueroa et al., 2010; Moghyazy et al., 2014; Mohamed Saied 2017]. For laboratory examination hematological methods are used [RBC, PVC, Hb, Smears 1 to showed alteration of blood [Callow et al., 1993; Brose et al., 1995]. Serological tests are used for diagnosis of the disease [Jonston et al., 1977; Brose e. al., 1995; DeVos et al., 2000]. PCR assays also used to detect the relative sensitivity DNA probes. For detection of Babesia in both tick vector and vertebrate host simple or multiplex PCRamplification of 18S rRNA gene fragment can be applied [Brose et al., 1995; Alhassan et al., 2005]. The alteration of enzyme activity in babesiosis in cattle resulting from liver damage. Also the changes in bilirubin due to congestion of hepatic vessels and liver destruction [Akinboade et al., 1984; Mohamed Saied, 2017]. At necropsy there is hypotensive shock syndrome with intravascular stasis and sequestration of parasitized red boold cells in the peripheral circulation. The characteristic lesion that cherry pink discoloration of cerebral cotex due to acute B.bovis infection. Also the spleen is enlarged soft and pulpy, sowollen liver, congested dark-colored kidneys. Other organs are congested or there are petechial hemorrahages and pulmonary edema [Bock et al., 2004; Sharma et al., 2013].

Successful treatment after early diagnosis of the parasites. Antiprotozoal agents diminazene acceturate and imidocarb dipropionate have a high affectivityagainst *Babesia* parasites [DeWaal and Combrink, 2006; Mosqueda *et al.*, 2012]. For prevention and control of this disease there are immunization, chemoprophlaxis and vector control. Vaccination can be used with caution in adult animals due to virulent of contamination with other disease agents and leading to hypersensitivity reactions in young animals [Demessia and Derso, 2015].

The Sudan has a huge size of cattle and the population are estimated of about 41.43 million head [Anon, 2008] and is ranking as the second biggest country in Africa and sixth in the world. The types of breeds found indigenous ecotype of zebu cattle, and this included Foga, Baggara, Kenana, Gaash and Butana or cross-breed of this ecotype with exogenous breed and these Frisian, Ayrshire and Holstein [Fadlelmoula *et al.*, 2007]. Boshara (2013) stated the prevalence rate of *Babesia* species in 300 cattle in East Darfur State and the overall prevalence rate reached 18%. While Awad *et al.* (2011) recorded the overall prevalence rate in 692 healthy cattle by using polymerase chain reaction to *Babesia bigemina*, *Babesia bovis* and *Anaplasma marginal* 4.0%, 1.9% and 6.1% respectively in northern region of Sudan. The aim of this paper was to identify *Babesia* spp. and tick vectors in cattle and to determine some hematological and biochemical parameters associated with the parasites in Elfasher locality.

Materials and Methods:

Study area:

This study was carried out from September, 2017 to March 2018 in Elfasher Locality – North Darfur State.

Collection of samples:

Atotal of 227 whole blood samples were taken from 2-4 years old cattle [5ml] by jugular vainpuncture randomly for smearing by using Giemsa staining method [McCosker, 1975[and estimation of packed cells volume [100 samples] by using microhemtocit method [Coles, 1986]. Also 100 blood samples were collected simultaneously for serum speparation for determination some biochemical parameters [Total proten, albumin, bilirubin, calcium, sodium and potassium] using test kits [Bio-system, S.A. Spain]. One hundred and fifty five ticks samples were collected from 23 animals in 70% ethanol for identification using Estrada standard [Estrada -Pena *et al.*, 2004].

Statistical Analysis:

The data statistically analyzed by SPSS version 13. Descriptive statistical and T-test were used for analysis.

Results:

The overall prevalence of bovine babesiosis

Examination of 227 smear samples and there were 133 samples positive to *Babesia* spp and 94 were negative. The overall prevalence of the disease was 58.5% [Table 1]

Table (1)The Overall prevalence of babesiosis in cattle [n=227] in Elfasher Locality – North Darfur

	<u> </u>	<u> </u>
	Number of animals	Frequency %
Positive	133	58.50
Negative	94	41.50
Total	227	100.00

Biochemical analysis:

As shown in table 2, there was significant decreased in packed cell volume in the infected animals, compared with non-infected animals. While all biochemical parameters showed no statistical significant difference to disease. But serum calcium levels showed significant difference [p<0.01].

There were 2 genera of ticks were identified and these were *Ripicephalus* and *Hyalomma*, included 7 species and these were *Ripicephalus evensi eversi* [37.5%], *Hyalomma anatolicum* [21.9%], *H.impletatum* [16.2%], *H.dromedarii* [12.9%], *H. rufipes* [1.2%], *H. truncatumim* [9.0%] and *H.predixtatus* [1.2%].

Table (2) The mean ± Standard error for some biochemical parameters associated with babesiosis in cattle [n=227] in Elfasher Locality – North Darfur State.

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Parameters	Infected	Non-infected	Significant
PCV	24.02±10.26	34.08 ± 10.57	**
Total protein	6.02 ± 071	6.10 ± 1.28	NS
Albumin	2.55 ± 2.36	3.06 ± 1.52	NS
Bilirubin	1.42 ± 0.51	1.60 ± 0.98	NS
Calcium	11.10±5.78	14.52 ± 5.93	**
Sodium	134.82 ± 5.60	136.66 ± 14.17	NS
Potassium	3.80 ± 4.41	3.97 ± 0.85	NS

^{** =} Significant at p<0.01, NS= No significant differences

Discussion:

Tick-borne diseases have affected the health of livestock leading to economical loss of productivity [ElAshker et al., 2015; Omar et al., 2015]. In the present study, the overall prevalence rate was 58.5% by microscopic examination of Babesia species [Table 1]. This prevalence is highest than prevalence in Danapur [35.5%] and Fatuna [26.21%] after retrospective data collected from Pantna using Giemsa stained thin blood smear method [Kumari and Kithin, 2018]. The result of Matovu et al. [2014] using microsopical prevalence for identification of Babesia species is lower. Also Shuaib et al. [2015] found that after examination of 803 thin blood smears the prevalence rate is 5.20% and this is lower than this result. This results are higher than that reported by Salih et al. (2008), in equatorial region (51.50%). Also Kivzari et al. (2012) recorded the prevalence rate 52.80% in South region. But the overall prevalence in this study is lower [Table 1] than recorded by Radostits et al. [1994] and Kumari and Jithin (2018) during rainy season [58.55%) with increase humidity which favor the development of ticks as vector of the disease. Immunofluorescent antibody test is more sensitive test and giving of high prevalence rate [68.7%] of bovine babesiosis [Shuaib et al., 2015].

Packed cell volume was significantly decreased [Table 2]. This finding in agreement with findings of Pandy and Misra [1987] and Mohamed Said [2017] who recorded that when the parasites are entered the blood circulation and invaded red blood cell membrane leading to intravascular hemolysis. Erythrocyte count, packed cell volume and hemoglobin level can be continued to decline steady after patency even the animals are recovered, but after the hematopoiesis is increased [Lewis *et al.*, 1981; Soulsby, 1982].

The study showed there was decreased in serum calcium level, this result in accordance with data recorded in cattle babesiosis [O'Neill, 1983; Zintl *et al.*, 2003; Mohamed Saied, 2017]. Generally, changes of calcium level due to mineral metabolism disturbances resulting from clinical or bone disease [Coles, 1986].

In these results, there were no changes in serum level of sodium, potassium, total protein and albumin. These results in contrast with Mohamed Saied (2017) who found that the increase in potassium level can be decreased, calcium, sodium and protein due to the harmful effect of toxic metabolites of *Babesia* spp. Also in these results serum bilirubin level was not altered, because no congestion of hepatic vessels due to the action of the parasites [Allen and Frerichs, 1975. Akinboade *et al.*, 1984]. From this study there were 2 genera of ticks were identified and these were *Ripicephalus* and *Hyalomma*, for transmission of bovine babesiosis. Friedcoff (1988) and Uilenberg (2016) found that *Ripicephalus* species have been implicated in the transmission of *Babesia* species to cattle and human. Also Altay *et al.* (2008) recorded that *R. bursi* was the main vector of cattle babesiosis in eastern Turkey. There are several studies carried out in Mediterranean region confirming that *R. bursi* transmitted *Babesia bovis and B. bigemena* (Bouattour and Darghough, 1996; Ravindran *et al.*, 2006; Ghirbi *et al.*, 2010).

In conclusion, bovine babesiosis is the most important tick-bone disease of cattle in Elfasher locality leading to death of the animals from anemia when the disease occurred in acute stage and may lowering the productive performance, specially the breed exports for improving the productivity of local cattle.

References:

Akinboacle, O. A; Diepolu, O.O.; Adtunji, A. [1984]. Enzyme and bilirubin reactions in bovine babesiosis. *Annals Researches Veterinaries*, 15(3), pp. 3133-3318.

Alhassan, A.; W. Pumidoming; M. Okamura; H. Hirata; R. Battestseg; K. Fujisaki; N. Hokoyama and I. igarashi (2005). Development of a Single – round and multiplex PCR method for the simultaneous detection of *Babesia cabatti* and *Babesia equi* in horse blood. *Veterinary Parasitology*, 129:4-49.

Allen, P.C. and Frewich, H. (1975). Experimental acute *Babesia caballi* infection: 1. Red blood cells dynamics-*Exp. Parasitol.* 37, 67-77.

Altay, K.; Fatih, Aydin, M.; Dumanli, N.; Aktas, M. (2008). Moleular detection of *Theileria* and *Babesia* infection in cattle. *Vet. Parasitol.*, 158: 295-301.

Anonymous (2008). Development of Statistic and Information. Annual Report. Ministry of Animal Resources and Fisheries, Khartoum, Sudan.

Awad, H. Antunes, S.; Galinda, RC; doRosario, VE; dela Fuente, J. Domingos; A.l Elhussein, A.M. (2011). Prevalence and genetic diversity of *Babesia* and *Anaplasma* species in cattle in Sudan. *Vet. Parsitol*, 181 (2-4): 146-52

Bock, R.'; ackson, L.; DE Vos, A.; Jorgensen, W. (2004). Babesiosis of Cattle. *Parasitology*, 129, 5247-5269.

Boshara, Sh. (2013). *Babesia of Cattle in East Darfur*. M. V. SC. A thesis, Sudan University of Science and Technology.

Bouattour, A. and Darghouth, M. A. (1996). First report of *Babesia divergens* in Tunisia. *Vet. Parasitol.*, 63: 161-165.

Brose, R.; Jorgensen, W.K.; Dalgliesh, R.J.; Friedhoff, K. T. and DE Vos, A.J. (1995). Current State and Future Trends in the Diagnosis of Babeiosis. *Veterinary Parasitology*. 57, 61-74.

Callow, L. L.; Rogers, R. J. and DE Vos, A.J. (1993). Tick – borne diseases: Cattle-Pathology and Serology. In *Australian Standards Diagnostic Techniques* for Animal Diseases (ed. Corners, L. A. and Bagust, T. J.), PP. 1-16. East Melborune, CSIRO Information Services.

Coles, M. H. (1986). *Veterinary Clinical Pathology*. Fourth Edition. B. Saunders Company, Philadelphia, London, Toronto, Montreal, Sydney, Tokyo. PP. 234.

Criado-Fornelio, A.; Marinez-Marcos, A.; Buling-Sarana, A. and Barba-Carretero, J.C. (2003). Molecular Studies on *Babesia, Theileria* and *Hepatozoon* in Southern Europe. Part 11. Phylogenetic analysis and evolutionary history. *Veterinary Parasitology*. 14, 173-994.

De Waal, D. T. and Comink, M. P. (2006). Live vaccines against bovine babesiosis. *Vet. Parasitol.* 138:88-96

Demessie, Y. and S. Derso (2015). Tick borne Hemoparasitic Diseases of ruminants: A review – *Advance* in *Biological Research*, 9(4): 210-224.

DeVos, A. J.; Jorgensen, W. K. and Molloy, J. B. (2000). Bovine babesiosis in *Manual of Standards for Diagnostic Tests and Vaccines*. Fourth edition, PP. 412-422. Paris, Office International des Epizootic.

- El-Ashker, M.; Hotzel, H.; Gwida, M.; El-Baskawy, M.; Silaghi, C. and Tomaso, H. (2015). Moleculary Biological Identification of *Babesia, Theileria* and *Anaplasma* species in Cattle in Egypt using PCR assays, gene sequence analysis and DNA microarray. *Veterinary Parasitology*, 207(3), 329-334.
- Enbiyale, G.; Debalke, D.; Aman, E.; Eodmim, B and Abebe, S. (2018). Review on bovine babesiosis. *Acta Parasitologica Globalis*, 9 (1): 15-26.
- Estrada-Pena, A.; Bouattour, A.; Camicas, J. L.; Walker, A.R. (2004). Ticks of domestic animals in Mediterranean region, a guide to identification of species. University of Zaragoza, Spain.
- Friedhoff, K. T. (1988). Transmission of Babesia. In: Ristic, M. editor. *Babesiosis of Domestic Animal and Man.* CRC press, Boca Raton, Florida, pp. 23-52.
- **Fadelmoula, A. A.; Yousief, I. A. and AbuNikhaila, AM (2007)**. Location curve and persistency of crossbred dairy cows in the Sudan. *Journal Applied Sciences Research*, 3(10), 1127-1133.
- Figueroa, V.; J.M. Hostis and E. Camus. (2010). Bovine babesiosis. In: Lefevre, P.C., Blancou, J.; Chermette, R.; Uilerbery, G., editrs. *Infectious and Parasitic Diseases of Livestock: bacterial disease, fungal diseases*, parasitic diseases. Paris, France: Lavoisioer, PP: 189: 1819-38.
- Ghirbi, M.; Hurtado, A. and Bouattour, A. (2010). *Theileria* and *Babesia* parasites in ticks Tunisia. *J. Emerg. Med.*, 57: 49-51.
- Hamsho, A.; G. Tesfamarym; Megersa and M. Megersa (2015). A cross-sectional study of bovine babesiosis in Teltrele District, Borena Zone, Southern Ethiopia. *Journal of Veterinary Science and Technology*, 6:230.
- Jonston, L. A.; Trueman, K. F. and Pearson, R. D. (1977). Bovine babesiosis, comparison of fluorescent antibody and giemsa staining in Post-mortem diagnosis of infection. *Australian Veterinary Journal* 53, 222-226.
- Kivari, F. M. Kapage, A. M.; Mbassa, G. K.; Mtut, and Wani, R.J. (2012). Epidemiological Perspectives of Ticjs and tick-borne disease in South Sudan: Cross-sectional survey results. *Onderspoort J.Vet. Res.*, 79(1): E1-E10.
- **Kumari, I. D. B. M. and Jithin, M. V. (2018)**. Prevalence of Babesiosis in Cattle in Pantna Region. *Int. J. Curr-Microbiol App. Sci.* Special issue 7: 5167-5174.
- Lewis, D.; Purnell, R. E.; Francis, LM.; Young, ER. (1981). The Effect of Treatment with Imidocrabdipropionate on the Course of *Babesia divergens* infections in splenectomised calves and their subsequent immunity to homologous challenge. *J. Comp. Pathol.*, 91(2): 285-292
- Matovu, E.; Tayebwa, D.S.; Natuhwera, J.; Mugezi, I. and Mahero, M. (2014). Epidemiology of increasing hemo- Parasite burden in Ugandan cattle. *Open Journal of Veterinary Medicine*, 4, pp.220-231.
- Moghazy, E. L.; M.H. Ebied; G. Mohamed; A. Abdelwahab Amr and E. S. Aziz. (2014). Epidemiological studies on bovine babesiosis and theileriosis in Qulubia Governorate. *Benha Veterinary Medical Journal*, 27 (1): 36-48.
- **Mohamed Saied, MM. (2017).** Studies of naturally infected babesiosis and its effect on some hematological and biochemical parameters in cattle in Qena, Egypt *Journal Dairy and Veterinary Sciences*. Vol. 4(3): 2573-2196.
- Mosqueda, A.; Olvera-Ramirez; G. Aguilar-Trapacamu and G. J. Canto (2012).

- Current advances in detection and treatment of babesiosis. *Current Medical Chemistry*, pp. 1504-1518.
- O'Neill, AR. (1983). Blood Transfusions in the Treatment of Redwater. Ir. Vet. News.
- Omar, A.O.J.; Sadat, M.; Emam, F.A. (2015). Determination of Oxidative Stress Markers Association with *Babesia bigemina*. *Agri. J. Biotechol. Res.* 3 (6): 150-5.
- Pandy, N.; Misra, S. (1987). Hematological and biochemical response to haemolytic anaemia of chemical babesiosis in cattle and therapy. *Ind. Vet. J.* 64: 882-886
- **Radostits, O. M. C.; Blood, C. C. and Hinchcliff, K. W. (1994).** *Veterinary Medicine*, 9th. Ed., WB. Saunders, London.
- Radostits, O. M.; G. C. Gay; K. w. Hinchliff and P. O. Constable (2007). A Text book of the disease of cattle, sheep, goat, pigs and horses, 10th, ed. London, Saunders Elsevier, pp.: 1110-1489, 1527-1530.
- Ravindran, R.; Rao, J. R.; Mishra, A.K. (2006). Detection of *Babesia bigemina* DNA in ticks by DNA hybridization using a nonradioactive probe generated by arbitrary PCR. *Vet. Parasitol.*, 141: 181-185.
- Salih, D.A.; Hassan, S.M.; Julla, I.I.; Kyule, M.N.; Zessin, K.H. and ElHussien, A.M. (2008). Distribution and application f ELISA for the seroprevalence of tick-borne disease in Central Equatorial State, Sudan. *Transbound. Emerg. Dis.*, 55 (5-6): 257-262.
- Schorn, S.; K. Plister; H. Reulen; M. Mathling and C. Silagh (2011). Occurrence of *Babesia* spp., *Richettsia* spp and *Bartonella* spp in *Ixodes ricinus* in Bavarian Public Parks, Germany. *Parasit. Vectors*, 4: 135.
- Sharma, A. R., R. C. Katoch; K. B. Nagal; R. S. Kishtwarta and S. K. Sharma (2000). Bovine babesiosis in Palam Valley of Himachal Paradesh. *Ind. Vet. J.* 77: 731-732.
- Sharma, A.; L. D. Singla; A. Tuli; P. Kuar, B. K. Bath, M. Javed and P. d. Juyal (2013). Molecular Prevalence of *Trypanosoma evansi* in dairy animals from Punjab, India, by duplex PCR: a step forward to the detection and management of concurrent latent infections. *Biomed. Research International*.
- Shuaib, Y. A.; Halima, M. O.; Hussien, M. O.; Bakhet, M. A.; Rihab, A. O.; Al-Nahas, A.; Siham, E. E.; Abdella, M. A.; Ismail, A. A. (2015). Sero prevalence of *Babesia bigamin* antibodies in cattle in North Kordofan State, the Sudan *ARC Journal of Animal and Veterinary Sciences*, Vol. 1, issue 2, pp. 1-11.
- **Soulsby, E. J. L. (1982)**. *Helminths, Arthropods and Protozoa of Domesticated Animals*. Seventh edition. Bailliere Tindall, London.
- **Spicker, A. R. J. r. Rothard G. Dvorak (2010)**. Emerging and exotic diseases of animals. 4th. ed. CFSPH Lowa State University, Lowa USA, p:102.
- **Uilenberg, G. (2006).** Babesia A historical Overview. Vet. Parasitol., 138: 3-10.
- Zintl, A.; Mulcahy, G. Skerrett, H. E.; Taylor, SM.; Gray, J. S. (2003). *Babesia adivergens*, a bovine blood parasite of veterinary and zoonotic importance. *Clin. Microbiol. Rev.* 16(4): 622-636.