MORPHOLOGICAL STUDIES AND CHARACTERSTIC OF ALKALOID CONTENTS OF THE SUDANESE PLANT KNOWN AS (WINCA)

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ABSTRACT on Anida to make this gallering and harmon.

A plant grown in Sudan locally named (winca) was studied for its morphological features and its characteristic alkhaloids that determine its classification. It was found that this plant morphologically belongs to the genus Catharanthus. Its chemical constituents indicate that it contains the dimmeric indole alkaloid called vinblastine which is not found in the other plant species, confirming that it belongs to the species roseus, hence this plant which is locally grown is Catharanthus roseus.

الملخص

أجريت دراسة لمعرفة الجنس والنوع الذي ينتمي إليه نبات الونكا الذي ينسو في السودان، وذلك عن طريق دراسة الوصف النباتي والتركيب الكيميائي له؛ جمعت عينات من النباتات ذات الزهرة الحمراء والبيضاء من منطقة شهبات (الخرطوم)، أثبتت نتائج الدراسة الوصفية أن نبات الونكا الذي ينمو في المودان ينتمي لجنس الكاتار انسن (Catharanthus)، كما أثبتت نتائج التحاليل الكيميائية أن هذا النبات يحتوي على قلويد الفيلاستين (Vinblastine) والذي لايوجهد (لا في عينة (Catharanthus roseus)، مما يثبت أن نبات الونكا الذي ينمو في المسودان هو (Catharanthus roseus L.)

INTRODUCTION:

Catharanthus roesus (L.) G.Don; formerly known by various names such as Vinca rosea L., V.speciosa (L.) Salisb, Provinca rosea (L.)

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Moench, Ammocallis rosea (L.) Small and Lochnera rosea (L.) Reichenbach, belongs to the family Apocynaceas (1). It contains more than 100 alkaloids among them are the powerful anti-tumor drugs (Vincristine and Vinblastine), and the monomeric indole alkaloid Ajmalicine which is used to improve cerebral circulation (2,3).

The genus Catharanthus G. Don; a genus in the subtribe Catharanthine of the Apocynaceae, is widely spread as ornamental and comprising eight species; seven of which are endemic to Madagascar. The best-known species of those of Madagascar is C.roseus, hence the name "Madagascar periwinkle". The genus is closely allied to the genera Amsonia and Vinca and that is why the species is wrongly designated in the past as Vinca rosea. Most of the differences are found in the floral struture. Differences in characters between these genera and their similarities are cited by Van Bergen et al 1996 (1).

Vinca series as V.minor, V.major, and V. herbacea are found in temperate zones, whilst Catharanthus genus such as (C. corianceus, C. Lanceus, C. trichophyllus, C. longifolius C. ovalis, C. scitulus and C. corianceus,)) are indigenous to Madagascar. And C. pusillus is indigenous to India, they occur in warm dry areas, mainly in tropical and subtropical countries (1). Moreover, Catharanthus roseus L.; contains the toxic antineoplastic dimmeric indole alkaloids; Vinblastine and Vincristine which are not present in Vinca species and other Catharanthus species (4). Vinblastine content varies with locality, part and age of the plant, it was reported that it ranges between 0.0045-0.0007% in the leaf(5,6,7), while Hirata et al 1989 (8) in Japan found that it is about 1.4% of the dry weight of leaves. Ajmalicine content in roots ranges between 0.63-1.2% in roots of one year old(6).

The objectives of this study were to investigate the morphological features and the characteristic alkaloids of the plant grown in Sudan, locally named as (winca), to clear up the confusion of whether it belongs to genus Catharanthus or Vinca.

EXPERIMENTS:

PLANT MATERIAL: Samples of 9 months old plants of red flowered, white flowered and white with red eye flowered of winca plant were uprooted from the field of the experimental farm at Shambat (Khartoum North). The age of leaves was three months after the last cut, observations of the vegetative and floral parts of the plant were noted. The various vegetative parts were examined with (X10 magnification handlens, whereas the structure of floral parts were carefully examined using a binocular dissecting microscope (X40) magnification.

CHEMICAL ANALYSIS: The dried samples of leaves and roots of the white and red flowered plant were analyzed by (Gorlaeus Laboratories) Division of Pharmacognosy-Center of Drug Research-Leiden.

The plant material was extracted with methanol by grinding three times for 5 minutes using an Ultra Turrax. The extracts were concentrated under reduced pressure and dissolved in a mixture of Iml of methanol, Iml of internal standard solution (50 g/ml dihydroquinone) and 8ml of 0.05M sodium phosphate buffer solution (pH 7.0). After filtration the filterate was applied to an RP-8 preconcentration column, previously prepared by washing with 3ml of methanol and 3ml of 0.05 M sodium phosphate buffer solution (pH 7.0). After application, the column was washed with 1.2ml of sodium phosphate buffer solution (pH9.5)-isopropanol (70:30) and 1ml of 0.05 M sodium phosphate buffer solution (pH9.5)-isopropanol (95:5). The alkaloids were diluted with 1.5ml of 0.05 M sodium phosphate buffer solution (pH 2.0)-isopropanol (60:40). The dilute was evaporated to dryness, the residue was dissolved in 0.5 ml of HPLC mobile phase and 25-100ml of this solution were injected in the HPLC system. A suspension culture medium was adjusted to (pH 7.0) by adding 2ml of 0.05 M sodium phosphate buffer solution (pH 7.0). After adding 1ml of internal standard solution the medium was filtered and applied to an RP-8 pre-concentration column. The washing and dilution steps were the same as described above.

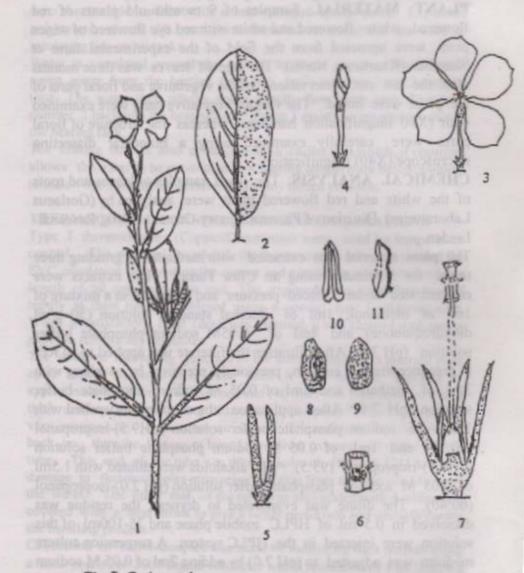


Fig. L Catharanthus roscus.

1. habit; 2, leaf; 3, flower; 4,bud; 5, fruit; 6, stem; 7, pistil; 8 &9, seed; 10&11, anther.

HPLC:

The extracts were analyzed by HPLC for alkaloids. The apparatus consists of an LKB 2150 pump (LKB, Bromma, Sweden) combined with a WISP 710B automated injection system (Waters Assoc., Milford, MA, U.S.A.), two LKB2158 Uvicord fixed-wavelength detectors operating at 275 and 313 nm, a Shimadzu Chromatopac C-R3A computing integrator (Shimadzu, Kyoto, Japan) and two-channel recorder (Kipp and Sons, Delf, The Netherlands). The column was a Waters Assoc. μ Bondapak-Phenyl (300X 3.9mm 1.D) and eluent was of 0.05 M sodium dihydrogen phosphate solution (6.8 g/L Na H2PO4 H2O, pH adjusted to 3.9 2-methoxyethanol (80:15:5) at a flow-rate of 2m1/min. Peak areas were used for calculation of the ejected amounts of alkaloids. The identities of the analytes were established by photodiod-array detection of their UV spectra (9).

RESULTS AND DISCUSSION GENERAL MORPHOLOGY:

The plant is a perennial herb, 50-120 cm high, erect or decubment, with white latex and a somewhat unpleasant smell. Trunk up to 1 cm in diameter, pale gray. Stems terete, narrowly winged, green, pink or pale red. Leaves glossy, decussate, petiole green or pink; blade dark green with paler main veins on both sides, oblong or obovate, acute or rounded or slightly retuse at the apex, cuneate at the base, apiculate, margin entire. Flowers axillary, solitary or paired. Sepals green. Corolla lobes pink with a darken eyes, white with yellowish eyes or white with red-purple eyes, paler or whitish outside; tube pale green, throat pale green, apex very pale greenwhite, bud white. Stamens with apex below the mouth of corolla tube, anthers subsessile. Pistil pale green surrounded by ring of hairs. Ovary ovoid. Style filiform. Fruit green fellicles forming a V shaped. Seeds small black (Fig 1.)

These morphological features indicate that the plant under study belongs to genus Catharanthus.

CHEMICAL CONSTITUENTS:

The results of the chemical analysis of the plant samples as shown in (table 1) indicate that: in the leaf material, vinblastine and Ajmalicine was about (0.7 & 0.66mg/g dry weight), respectively, in the Red flowered plant, and about (0.57 & 0.64mg/g dry weight), respectively, in the White flowered plant. The detection of Vinblastine in the leaves confirms that this plant is Catharanthus roseus L. (4,10) who declared that C. roseus is the only plant that contains Vinblastine. The presence of 3, 4- Anhydro-vinblastine which is the key intermediate in the coupling of Catharanthine and Vindoline to give Vinbastine - is a preliminary indicator for Vinblastine (11,12).

On the other hand, the roots were devoid of vinblastine and contain about (4.94&6.03 mg/g dry weight) of Ajmalicine, in the Red and White flowered plants respectively. These results were in conformity with the previous result(6).

CONCLUSION:

The results of this study indicate that the (Red and White flowered) plants grown in Sudan and locally Known as (winca) do not belong to the genus Vinca but to the genus Catharanthus, species roseus. Synonym Vinca rosea L.

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Table 1. Alkaloid content of the Red and White flowered Catharanthus roseus plants grown in Sudan (mg/g DW)

Strains	Alkaloids (mg/g DW)			
5-5-0	atification 3	Ajmalicine	Vinblastine	AVLB
Red	Leaf	0.66±0.11	0.73±0.03	7.67±1.43
flowered	Root	4.96±0.96	nd	nd
White	Leaf	0.64±0.11	0.57±0.02	9.34±1.02
flowered	Root	6.03±0.96	nd	nd

Notes:

nd: not detected

AVBL= 3,4-anhydro-vinblastine

DW=Dry weight

REFERENCES:

- Van Bergen, M. and Snoeijer, W. Catharanthus G. Don. (1996).,
 The Madagascar periwinkle and related species. Agri. Uni. Prosea Project, Wageninge, The Netherlands,pp,10-58
- Van der Heijden, R., Snoeijer, W. and Verpoorte R.C. roseus (L.)
 G.Don(1992): enhoudstoffen en gebruik. Pharmaceutish
 Weekblad, 127: 1283-1290.
- Van der Heijden, R., Verpoorte, R. and Ten Hoopen, J.J.G (1989).
 Cell, Tissue and Organ Culture, 18:231-280.
- 4.Khoury.R.(1993),.History Botany, Geography, Phytochemistry Pharmacology, dosage and toxicity of 6 species of C. roseus and 3 species of Vinca. Australian J. of medicinal Herbalism, 5,3:66-68.
- Paradasani, K.M.(1979), Singh, S. and Sarin, J.P.S.
 Chromatographic estimation f Vincaleukoblastine in C. roseus L.
 Indian J. of Pharmaceutical Science, 41,5:207-208.
- 6.Atal, C.K. and kapur, B.M (1982). In: Cultivition and Utilization of Medicinal Plants. Regional Reaserch Laboratory. Council of Scientific & Industrial Research, Jammu-Tawi, India,pp.279-287.
- 7.Atta-Ur-Rahman, Bashir, M., Hafeez, M., Perveen, N., Fatima, J. and Mistry, A.N (1982). A rapid proedure for isolation of C. roseus L alkaloids. Planta medica, 47:246-247.
- 8.Hirata, K., Kobayshi, M. Miyamoto, K., Hoshi, T. Okazaki, M. and Muira, Y (1989). Quantitiative determination of Vinblastine in tissue culture of C. roseus (L.) by Radio-immunoassay, Planta medica, 55:262-264.
- 9. Van der Heijden, R.(1987), Lambing, P.J., Out, P.P. and Verpoorte, R.high performance liquid chromotographic determination of indole Alkaloids in suspension culture of Tabernaemontana divariata. J. Chromatogr. 396:287-295.

- 10.Madati, P.J., Pazi, H.A.M (1979), and Asha Ernest. Phytochemical investigation of C.roseus (L.) growing wildly in East Africa J. of African Medicinal Plants, 2:1-15.
- 11. Endo, T.(1987), Goodbody, A., Vukovic, J. and Misawa, M. Enzymes from C. roseus L. cell suspension culture, that couple Vindoline and Catharanthine to form 3,4-Anhydro-vinblastine. Phytochemistry:2147-2149.
- 12.Endo,T(1987)., Goodbody, A., Vukovic, J. and Misaa, M. Biotransformation of Anhydro-vinblastine to Vinblastine by a cell - free Extract of C. roseus L. suspension cultures::32333-3234

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