



بسم الله الرحمن الرحيم

Sudan University of Science & Technology

College of Graduate Studies



## Assessment of Prolactin level, Cholesterol and Triglyceride among Women with Polycystic Ovary Syndrome.

تقييم مستويات هرمون البرولاكتين, الكوليسترول وثلاثي الغليسريدات لدى النساء المصابات بتكيس المبايض

A Dissertation Submitted in Partial Fulfillment for the Requirements of M.Sc. Degree in Medical Laboratory Science  
-Clinical chemistry

**By:**

**Hind Ismail Abdalla Ahmad**

**B.Sc. in clinical-chemistry-medical laboratory**

**University of AL ImamAL Mahdi(2013)**

**Supervisor:**

Dr.Abdelgadir ElmugadamPh.D. Clinicalchemistry-Medical Lab Science

**Feb.2020**

## الآية

قال تعالى :

وَقُلِ اعْمَلُوا فَسَيَرَى اللَّهُ عَمَلَكُمْ وَرَسُولُهُ وَالْمُؤْمِنُونَ ۖ وَسَتُرَدُّونَ إِلَىٰ  
عَالِمِ الْغَيْبِ وَالشَّهَادَةِ فَيُنَبِّئُكُمْ بِمَا كُنْتُمْ تَعْمَلُونَ ﴿105﴾

صدق الله العظيم

سورة التوبة (الآية 105)

## *Dedication*

*With love and gratification I dedicate this work*

*To:*

*That person who hold my hand to teach me how*

*I step up on my life*

*My mother*

*Person who protect and help me all the timetolerate the*

*Sun burn and difficulties for us*

*My father*

*Person who stand with me and continuing to support me*

*My husband*

*My daughters I love you so much you are the reason of my happiness*

*keep twinkling on my sky ,godbless you*

*My sister,brothers,friends and relatives for their support*

*To all un visible hand work.*

## *Acknowledgments*

First great thanks referring to Allah for his mercies and guidance to live and achieve my goals.

Secondary it gives me a pleasure and most honored to become supervised by such nice person **Dr.Abd elgadirElmugadam.**

Also special thanks to all members of Sudan university for science and technology (SUST) specially to staff members of clinical chemistry.

Great thankfulness to volunteers for their nice dealing with the research demands without any growl; as same as Dr. AL sir abo Alhassan infertility center staff.

## Abstract

### Background:

Polycystic ovary syndrome (PCOS) is a heterogeneous endocrinopathy characterized by both reproductive and metabolic abnormalities. This study was carried out to evaluate serum prolactin, cholesterol and triglyceride in polycystic ovary syndrome women in Khartoum state.

### Methodology:

The study was carried on one hundred volunteers in reproductive age, including 50 women as cases and 50 as controls at DR AL SIR Abo- Alhassan infertility center from May to November (2019). Venous blood samples were collected, processed, and analyzed. Prolactin level was estimated by Enzyme linked immune sorbent assay technique and cholesterol and triglyceride levels were estimated by spectrophotometer biosystem -310. Experimental case-control methodology was used. The obtained results were analyzed statistically by using SPSS software program.

### Result:

The level of prolactin was significantly increased with ( $p$ -value=0.000), also cholesterol significantly increased ( $p$ -value= 0.009) and triglyceride ( $p$ -value=0.001).

In polycystic ovary syndrome patients compared to control group according to infertility, duration of disease, menstrual regulation, family history, cosmetic use, and medication, the study showed that prolactin increase was insignificant with infertility, therapy, cosmetic use, and history with ( $p$ .value=0.124) ( $p$ .value=0.761) ( $p$ .value =0.611) ( $p$ .value= 0.986) respectively. But cholesterol increase was significant compared with infertility with ( $p$ .value=0.031) and insignificant with

therapy (p.value=0.908), cosmetic (p.value=0.786) and menstrual cycle (p.value=0.180).

Triglyceride increased insignificantly with infertility (p.value=0.430), therapy (p.value=0.084) cosmetic (p.value=0.137) and menstrual cycle (p.value =0.721).

## المستخلص

### خلفية:

متلازمة المبيض المتعدد الكيسات (متلازمة تكيس المبايض) هي اعتلال الغدد الصماء الغير متجانس يتميز بكل من التشوهات التناسليه واعتلال في التمثيل الغذائي, أجريت هذه الدراسة لتقييم مصل البرولاكتين, الكوليسترول وثلاثي الغليسريدات لدى النساء المصابات بمتلازمة المبيض المتعدد الكيسات في ولاية الخرطوم.

### منهجية:

أجريت الدراسة على مائة متطوعة في سن الأنجاب ومنهم 50 امرأة (حالة) و50 (مريض) في مركز الدكتور أبو الحسن للأنجاب وامراض الخصوبه في الفتره من مايو إلى نوفمبر (2019) وتم جمع عينات من الدم الوريدي ومعالجتها وتحليلها. قدرت بواسطة استخدام تقنية فحص الانزيمات المناعي المرتبط لمصل البرولاكتين و قدرت مستوى الكوليسترول والدهون الثلاثية بالنظام الحيوي الطيف \_310. وقد تم تحليل النتيجة التي تم الحصول عليها إحصائيا باستخدام برنامج الحزمة الإحصائية للعلوم الاجتماع

### النتيجة:

كان مستوى البرولاكتين في زيادة كبيرة بقيمة ( $p=0.000$ ) وكذلك زيادة كبيرة في مستوى الكوليسترول بقيمة ( $p=0.009$ ) ومستوى ثلاثي الغليسريدات بقيمة ( $p=0.001$ ) في مرضى متلازمة تكيس المبايض مقارنة مع النساء التي لم تنجب, وفترة الحيض, وتاريخ الأسرة المرضي واستخدام مستحضرات التجميل والأدوية المستخدمة. أظهرت الدراسة ان هرمون البرولاكتين يزيد عند النساء اللاتي لم ينجبن ويستخدمن بعض العلاجات ومستحضرات التجميل ومن لديهن تاريخ مرضي بقيمة ( $p.value=0.986$ )

( $p.value=0.611$ ) ( $p.value=0.761$ ) ( $p.value=0.124$ ) على التوالي . لكن الكوليسترول يزداد طرديا مقارنة مع اللاتي لم ينجبن ب ( $p.value=0.031$ ) وليس لديه علاقة واضحة مع اللتي يستخدمن العلاج ( $p.value=0.908$ ) ومستحضرات التجميل ( $p.value=0.786$ ) ودورة الحيض ( $p.value=0.180$ ) ويزداد ثلاثي الغليسريدات بشكل ضئيل عند النساء اللاتي لم ينجبن ( $0.430$ ) واللتي يستخدمن العلاج ( $p.value=0.048$ ) ومستحضرات التجميل ( $p.value=0.137$ ) وفترة الحيض

( $p.value=0.721$ ).

### الخلاصة:

وجد أن هناك زيادة في مستوى البرولاكتين , وثلاثي الغليسريدات والكوليسترول في المرضى الذين يعانون من متلازمة تكيس المبايض.

## List of Contents

Content	Page Number
الآية	II
Dedication	III
Acknowledgement	IV
Abstract	V
List of Contents	VI
List of Tables	VII
List of Figures	VIII
Abbreviation	XIV
<b>Chapter One: Introduction</b>	
1.1 Introduction	2
1.2. Rationale	4
1.3 Objectives/	4
<b>Chapter Two: Literature Review</b>	
2. Literature review	7
2.1 Definition	8
2.2 Symptoms	8
2.3 Cause	8
2.3.1 Excess insulin.	8
2.3.2 Low-grade inflammation	9
2.3.3 Heredity	9
2.3.4 Excess androgen.	9



2.3.5 Complications	9
2.3.6 Diagnosis of poly cystic ovary syndrome	12
2.4 Prolactin	12
2.4.1 Production of prolactin	13
2.4.2 clinical manifestation	13
2.4.3 Synthesis of prolactin	14
2.4.4 Function	15
2.4.5 Reference Range	16
2.5.5 Interpretation	16
2.5 Dyslipidemia and polycystic ovary syndrome	17
2.5.1 cholesterol	17
2.5.1.1 Sources of cholesterol	17
2.5.1.2 Functions of cholesterol	17
2.5.1.3 Causes of high cholesterol	17
2.5.1.4 Diagnosis of cholesterol	19
2.6 triglyceride:	19
2.6.2 Normal level of triglyceride	20
<b>Chapter Three: Material and Methodology</b>	
3. Material and Methodology	23
3.1 study design	23
3.2 study area	23
3.3 study population and sample size	23
3.4 inclusion and exclusion criteria	23
3.5 collection sample	23

3.6 Ethical approval	23
3.7 Methodology	24
3.7.1 prolactin:(immune enzymatic assay)	24
3.7.1.1 Principle	24
3.7.1.2 Procedure of Measurement (appendix I)	24
3.7.2 Cholesterol	24
3.7.2.1 Principle of cholesterol oxidase method	25
3.7.2.2 Procedure of measurement(appendix II)	25
3.7.3 Triglyceride	25
3.7.3.1 Principle	25
3.7.3.2 Procedure of measurement (appendix III)	25
3.8 quality control	25
3.9 statistical analysis	25
<b>Chapter Four: Results</b>	
4. Results	27
<b>Chapter Five: Discussion, Conclusion &amp; Recommendation</b>	
5.1 Discussion	41
5.2 Conclusion	43
5.3 Recommendation	43
References	44
Appendices	51

## List of Tables

<b>Table</b>	<b>Page Number</b>
Table (4-1): show the demographic data of the patient of polycystic ovary syndrome.	24
Table(4-2):Mean concentration of patient age and duration of diseases.	25
Table (4-3): show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in patient with control group.	25
Table (4-4): show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in Infertile patient and non-infertile.	26
Table(4-5): show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in patient use therapy and non-use.	26
Table (4-6) show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in patient use Cosmetic and not use.	27
Table (4-7): show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in patients have history of poly cystic ovary syndrome and not.	27
Table (4-8): show the Mean $\pm$ SD of prolactin, cholesterol and triglyceride in regular menstrual cycle and irregular menstrual cycle patient.	28

## List of Figures

<b>Figure</b>	<b>Page Number</b>
	29
Figure (4-1): Scatter plot of pearson correlation between prolactein and age of PCOS woman	30
Figure (4-2): Scatter plot of pearson correlation between cholesterol and age of PCOS woman	32
Figure (4-3): Scatter plot of pearson correlation between triglyceride and age of PCOS woman	33
Figure (4-4): Scatter plot of pearson correlation between prolactein and BMI of PCOS woman	34
Figure (4-5): Scatter plot of pearson correlation between cholesterol and BMI of PCOS woman	35
Figure (4-6): Scatter plot of pearson correlation between triglyceride and BMI of PCOS woman	36
	37

## Abbreviations

<b>ABBREVIATION</b>	<b>MEANING</b>
BMI	Body Mass Index
FFAs	Free Fatty Acid
GnRH	Gonadotropin releasing hormone
HDL-C	High Density Lipo Protein
HL	Hepatic lipase
IR	Insulin Resistance
JACC	Journal of The American College of Cardiology
LDL-C	Low Density Lipo Protein
MS	Metabolic syndrome
PCOS	Polycystic Ovary Syndrome
T2DM	Type Two Diabetes Mellitus
TRH	Thyroid Releasing Hormone
TSH	Thyroid Stimulating Hormone
VLDL	Very Low Density Lipo Protein



# *Chapter One*

## *Introduction*

## **1.1 Introduction:**

Polycystic ovary syndrome (PCOS), characterized by hormonal imbalance and ovarian dysfunction, often starts during adolescence. Inconsistent diagnostic criteria, variable provider knowledge, and lack of consensus pose specific challenges for the care of women with PCOS. These factors encourage inaccurate diagnosis with both under and overdiagnosis. This unfavorable diagnostic experience exasperates affected women and limits timely opportunities for intervention to minimize associated comorbidities, especially during the transition from pediatric to adult care. Recognition of these issues in the care of adolescents and women with PCOS inspired the development of the International Evidence-Based PCOS Guidelines, which emphasize the prevention, screening, and treatment of PCOS across the reproductive lifespan. The Guidelines and accompanying meta-analyses focus on three major categories of associated comorbidities: reproductive; metabolic and psychological. With the exception of infertility, this article considers common manifestations and comorbidities associated with PCOS throughout the lifecycle. The world wide prevalence of PCOS is estimated to be 5-10%. (kovanci, 2015).

Healthy lifestyle interventions with prevention of excess weight gain comprise the primary intervention for all comorbidities. Hence, early identification of girls “at risk” for PCOS and those with PCOS is a priority. Extensive guidelines for provider and patient education aim to decrease the medical, psychosocial, and economic burdens attributable to PCOS and its associated comorbidities. **(Selma et al., 2019).**

PCOS is not the only condition that can cause these types of menstrual irregularities or infertility but it is the most prevalent along with high prolactin



levels. High prolactin levels have many of the same symptoms as PCOS and needs to be ruled out using a blood test to be certain of a PCOS diagnosis. Prolactin is a hormone whose primary function is to initiate lactation. It is released by the pituitary gland, a small organ located at the base of the brain that influences the entire body. The pituitary gland produces prolactin and a number of other key hormones including growth hormone, luteinizing hormone, thyroid stimulating hormone, and adrenocorticotropin hormone. Excessive prolactin can cause a decrease in sex hormones like estrogen. When you have excessive amounts of prolactin in your blood it is called hyperprolactinemia. This is sometimes a marker of a condition known as prolactinoma, which is a tumor of the pituitary gland (usually non-cancerous). Prolactinomas produce higher than normal levels of prolactin, which can wreak havoc on the body and produce many PCOS-like symptoms (Robert et al., 2012) several factors, such as genetics, are variables in the development of polycystic ovarian syndrome. If your sister or mother has PCOS, your risk of also having it increases. Hormone imbalance is definitely a major influencing element in PCOS, together with a condition known as Insulin Resistance. (leo et al., 2016.)

The diagnosis of PCOS is based on the Rotterdam criteria for the presence of any two of the following conditions: (i) chronic anovulation, (ii) clinical/biochemical parameters for hyperandrogenism, and (iii) polycystic ovaries on ultrasonography. Insulin resistance, hyperandrogenism, and dyslipidemia are presumed to be the major risk factors for CVD in women with PCOS. (wang, 2017).

Insulin resistance and dyslipidemia allegedly play a key role on the risk of cardiovascular pathology in women with PCOS. The extent to which dyslipidemia leads to this risk is still not well understood. Dyslipidemia is the

most common abnormality in PCOS, with elevated levels of total cholesterol, triglycerides, and low-density lipoprotein cholesterol (LDL-C) and with low levels of high-density lipoprotein cholesterol (HDL-C). Talbott et al. reported increased level of LDL-C in patients with PCOS, and Conway et al. reported that the most characteristic lipid alteration is decreased levels of HDL-C. There are few studies done to know the alteration in serum lipid profile in PCOS patients; thus, this study was done to know the lipid profile variation in women with PCOS. (Gobal et al., 2016)

## **1.2.Rationale:**

Polycystic ovary syndrome(PCOS) is the most series problem in woman worldwide and has public health importance as it is very common, affecting up to one in five woman of reproductive age and there is the most reason of infertility and abortion for many woman in last years which to be a target for research to find out the reason of thisdisease .Therefor,my objective is to examine the prolactin level ,cholesterol and triglyceride in polycystic ovary syndrome woman.

## **1.3Objectives:**

General objective:

To evaluate serum prolactin level, cholesterol and triglyceridein polycystic ovary syndrome woman in Khartoum state.

Specific objective:

1. To measure serum prolactin level, cholesterol and triglyceride in patient with PCOS compared with control group.
2. To correlate between prolactin level, cholesterol and triglyceride.
3. To calculate body mass index(BMI).

4. To correlate the biochemical parameter of the study with the variable age, BMI, history of disease, use of cosmetic and regulation of menstrual cycle.

# *Chapter Two*

## *Literature Review*

## **2.Literature review**

### **2.1 Definition:**

Polycystic ovary syndrome (PCOS) is a heterogeneous endocrinopathy characterized by both reproductive and metabolic abnormalities. In 2003, the European Society for Human Reproduction and Embryology and the American Society for Reproductive Medicine redefined PCOS as the presence of two or more of the following features: oligo- or anovulation, signs of clinical or biochemical hyper androgen and ultrasonographic evidence of polycystic ovaries once other related endocrine and gynecological disorders are excluded. (R. Azziz et al., 2016)

Recent studies have shown an early onset of abnormal cardiovascular risk profile in women with PCOS. Many of these women develop abnormal glucose and lipid metabolism, hypertension, obesity, insulin resistance and other features suggestive of systemic inflammatory response. Because of the higher rates of prevalence of these risk factors, the symptoms of majority of the women with PCOS also fit into metabolic syndrome, which is a known risk factor for cardiovascular disorders.

A close correlation was observed between adiposity and the severity of symptoms in women with PCOS. The android type of body fat distribution, which is more commonly associated with metabolic disturbances, was found to be more common in women with PCOS. (Ghaffar et al., 2014)

In fact, studies that emphasized on anthropometric parameters in women with PCOS have revealed higher body mass index (BMI) and increased waist circumference in women with PCOS. (Patel et al., 2018) (Broughton et al., 2017).

## **2.2 Symptoms:**

Signs and symptoms of PCOS often develop around the time of the first menstrual period during puberty. Sometimes PCOS develops later, for example, in response to substantial weight gain. A diagnosis of PCOS is made when you experience at least two of these signs: Irregular periods. Infrequent, irregular or prolonged menstrual cycles are the most common sign of PCOS. For example, you might have fewer than nine periods a year, more than 35 days between periods and abnormally heavy periods, Excess androgen, Elevated levels of male hormone may result in physical signs, such as excess facial and body hair (hirsutism), and occasionally severe acne and male-pattern baldness. Polycystic ovaries Your ovaries might be enlarged and contain follicles that surround the eggs. As a result, the ovaries might fail to function regularly. PCOS signs and symptoms are typically more severe if you're obese. (Fahimeh et al., 2015).

## **2.3 Causes:**

The exact cause of PCOS isn't known. Factors that might play a role include:

### **2.3.1 Excess insulin.**

Insulin is the hormone produced in the pancreas that allows cells to use sugar, your body's primary energy supply. If your cells become resistant to the action of insulin, then your blood sugar levels can rise and your body might produce more insulin. Excess insulin might increase androgen production, causing difficulty with ovulation. (John et al., 2013)

### **2.3.2 Low-grade inflammation.**

This term is used to describe white blood cells' production of substances to fight infection. Research has shown that women with PCOS have a type of low-

gradeinflammation that stimulates polycystic ovaries to produce androgens, which can lead to heart and blood vessel problems. (Milica et al.,2019)

### **2.3.3 Heredity.**

Research suggests that certain genes might be linked to PCOS.(Jones et al., 2016)

### **2.3.4 Excess androgen.**

The ovaries produce abnormally high levels of androgen, resulting in hirsutism and acne.(Jameson et al., 2017).

### **2.3.5Complications**

Complications of PCOScan include:

Infertility

Gestational diabetes or pregnancy-induced high blood pressure Miscarriage or premature birthNonalcoholic steatohepatitis —a severe liver inflammation caused by fat accumulation in the liverMetabolic syndrome —a cluster of conditions including high blood pressure, high blood sugar, and abnormal cholesterol or triglyceride levels that significantly increase your risk ofcardiovascular disease

Type 2 diabetes or prediabetes ,Sleep apnea, Depression, anxiety and eating disorders ,Abnormal uterine bleedingCancer of the uterine lining (endometrial cancer).Obesity is associated with PCOSand can worsen complications of the disorder.

**(Lobo et al., 2017)**

Studies have revealed that there are different degrees of obesity, dyslipidemia, insulin resistance (IR), oxidative stress and other metabolicabnormalities(Tehrani etal., 2014), and among these abnormalities,

dyslipidemia is one of the most common phenomena observed in women with(PCOS). (Macut et al., 2013).

Lipid abnormalities are found in women affected by PCOS. A recent study showed that mild hypercholesterolemia is frequently encountered in women with PCOS (Pergialiotis et al., 2018) .

Different lipid patterns are present in PCOS, including low levels of high-density lipoprotein cholesterol (HDL-C), high triglyceride (TG), total cholesterol (TC) and low-density lipoprotein cholesterol(LDL-C) and significantly higher lipoprotein concentrations. A recent animal study found that the implementation of a high-fat diet in prepubertal rats induced metabolic and ovarian alterations that were frequently present in PCOS, thus suggesting a potential impact of hyperlipidemia on the hormonal profile (Patel, 2018).

Obesity exerts a clear effect on oocyte quality and early embryo growth that is triggered by lipo toxicity induced endoplasmic reticulum stress, mitochondrial dysfunction and apoptosis (Broughton, 2017).

Observed phenotype-specific differences in lipid profiles based on androgen levels suggesting that androgens play an important role in hyperlipidemia. (Spalkowska et al., 2018).

However, some studies have shown that hypomethylated genes related to the synthesis of lipids and steroids may promote the synthesis of steroid hormones, including androgen, which could partially explain the mechanisms of hyperandrogenism in PCOS (Pan et al., 2018). These studies suggest that hyperandrogenism is an important cause of lipid abnormalities; in contrast, changes in genes related to lipids promote the development of hyperandrogenism. Women with mild hypercholesterolemia have a 11 higher body mass index (BMI)



and higher fasting insulin and IR levels than women with PCOS and normal cholesterol levels (Pergialiotis et al., 2018)

Experimental model rats exhibited ovarian changes, such as an increase in the number of cystic follicles and an increase in follicular wall thickness, in a high-fat diet-induced model of PCOS (Patel et al., 2018) suggesting that infertility is associated with dyslipidemia in PCOS. In conclusion, abnormal lipid metabolism can promote the pathophysiology of hyperandrogenism, IR, oxidative stress, and infertility in PCOS. The waist-to-hip ratio, which is higher in PCOS women, is more sensitive to dyslipidemia because centrally located adipocytes appear to exert a detrimental effect on blood lipids; centrally distributed adipose tissue can also secrete a range of adipokines into circulation to increase inflammation (Spritzer et al., 2015) suggesting that adipokines are associated with inflammation in PCOS. (Mannera et al., 2011)

In addition, a high level of circulating free fatty acids (FFAs), which contribute to the development of PCOS, is most common among obese PCOS patients (Mlinar et al., 2011).

A study found low levels of HDL and high levels of TGs in women with PCOS (Zhan et al., 2018) and HDLs are predictive for the occurrence of metabolic syndrome (MS) in PCOS (Shaman et al., 2017).

A clear increase in LDL has been observed in PCOS, and LDL levels decrease after treatment with statins. Therefore, an abnormal lipid profile affects the pathological development of PCOS. FFAs, HDL and LDL related to lipid abnormalities play an important role in PCOS. Obesity, IR and hyperandrogenemia exert independent and interrelated effects on circulating lipid profiles, but the effect of lipid profiles on IR, hyperandrogenemia, oxidative stress and anovulation remain under exploration. (Seyam et al., 2017).

### **2.3.6 Diagnosis of poly cystic ovary syndrome:**

Diagnosis can generally be accomplished with a careful history, physical examination, and basic laboratory testing, without the need for ultrasonography or other imaging. Hyperandrogenism can be diagnosed clinically by the presence of excessive acne, androgenic alopecia, or hirsutism (terminal hair in a male-pattern distribution) or chemically, by elevated serum levels of total, bioavailable, or free testosterone or dehydroepiandrosterone sulfate measurement of androgen levels is helpful in the rare occasion that an androgen-secreting tumor is suspected (e.g., when a patient has marked virilization or rapid onset of symptoms associated with PCOS)(Gibson et al., 2016).

Ovulatory dysfunction refers to oligomenorrhea (cycles more than 35 days apart but less than six months apart) or amenorrhea (absence of menstruation for six to 12 months after a cyclic pattern has been established (Dewailly et al., 2014).

### **2.4 Prolactin:**

Prolactin is a hormone that affects many different hormones in the body. Present in both men and women, it rarely causes problems, but those who are serious about their health should understand what it is and how it impacts the body's overall health and well-being .Prolactin, as its name implies, is a hormone that promotes lactation (breast milk production) in mammals and is responsible for a number of other functions and systems. Prolactin is created in the front portion of the pituitary gland in your brain, as well as in the uterus, brain, breasts, prostate, adipose tissue, skin, and immune cells. (Bernard V et al., 2019).

Prolactin is released when a newborn baby suckles at his/her mother's breast, causing the production of milk. However, this is just the primary and most well-known purpose of prolactin. (Jinet al., 2019)

### **2.4.1 Production of prolactin:**

Production of prolactin is controlled by two main hormones: dopamine and estrogen. These hormones send a message to the pituitary gland primarily indicating whether to begin or cease the production of prolactin. Dopamine restrains the production of prolactin, while estrogen increases it. (Li et al., 2019)

### **2.4.2 clinical manifestation:**

For most people, prolactin does its job without a problem, and few are aware of the impact it has on their health. Yet some people can struggle with prolactin levels, which can cause a variety of problems. (Bernard et al., 2019) Too much prolactin in the blood causes hyperprolactinemia, a condition that can lead to menstrual disturbances, estrogen deficiency and testosterone deficiency. High prolactin levels also can cause unwanted lactation. This often occurs during pregnancy or when the thyroid is not functioning properly. Pituitary tumors, known as prolactinomas, and medications that reduce dopamine can also lead to increased prolactin levels. High levels of prolactin are linked to sexual problems. Some of these conditions can be treated with medications that mimic the action of dopamine. It's also possible to have too little prolactin, a condition known as hypoprolactinaemia. This is extremely rare, but it can occur if people have under-active pituitary glands. This is commonly noticed in women after pregnancy who are not able to produce sufficient milk. No other proven health effects of low prolactin levels have been noted. Research is underway to determine if those with low prolactin levels suffer from a reduction in immune system responses.

(Vilaret al., 2019).

Prolactin has a major role in the physiology of the breast, especially in females. Both a lack of prolactin secretion and excessive prolactin secretion result in

clinical presentations. The level of prolactin hormone is detrimental to the female's ability to lactate. Thus, imbalances in the prolactin level can compromise this ability. Furthermore, disruption in the prolactin balance can have significant effects on the menstrual cycle. In females, too much prolactin leads to amenorrhea (absence of menstruation). The physiological reason for this is related to the prolactin role in the hypothalamus-pituitary reproductive axis which will be discussed in detail later. In males, however, prolactin level imbalances have different clinical manifestations. Too much prolactin in males' results in headaches and decreased libido. The decreased libido in males is associated with decreased spermatogenesis as a result of elevated prolactin affecting hypothalamus pituitary reproductive axis.(Matalliotakis et al., 2019)

(Auriemma et al., 2019)

### **2.4.3 Syntheses of prolactin:**

Prolactin is synthesized by lactotrophs in the anterior pituitary gland. The number of lactotrophs will increase during pregnancy in response to the physiological need to develop breast tissues and to prepare for milk production. Prolactin production is regulated at the gene transcription level. Factors that stimulate prolactin secretion to upregulate prolactin gene transcription while factors that inhibit prolactin secretion downregulate prolactin gene transcription.(Auriemma et al., 2019).

The most important factors that regulate prolactin secretion are: thyrotropin-releasing hormone (TRH) and dopamine both secreted by the hypothalamus. TRH has a stimulatory effect on thyroid-stimulating hormone (TSH) as well as prolactin; whereas, dopamine has an inhibitory effect on prolactin. In the absence of pregnancy (i.e., high estrogen) or lactation, prolactin is tonic ally inhibited by dopamine and the effect of dopamine trumpets the effect of TRH. Prolactin has a

negative feedback on its own production by stimulating the release of dopamine in the hypothalamus. Medications that antagonize dopamine production, for example, an antipsychotic block, the tonic inhibition of dopamine result in symptoms of excessive prolactin. Conversely, medications that are dopamine agonists such as bromocriptine or cabergoline inhibit prolactin secretion. Thus, these medications are used in the treatment of prolactinoma. Estrogen in high levels, as the case with pregnancy, stimulate prolactin release directly from the anterior pituitary. Interestingly, suckling stimulates sensory nerves in the nipple that carries the signal via the spinal cord to arcuate nucleus which inhibits dopamine release by removing the inhibitory action of dopamine on prolactin. At the same time, the afferent signal from the nipple activates supraoptic and paraventricular nuclei to increase the production of oxytocin which allows for milk ejection. Prolactin also has an inhibitory effect on the release of gonadotropin-releasing hormone (GnRH) produced by the hypothalamus-inhibiting FSH and LH release from the anterior pituitary. This leads to inhibition of the ovulatory cycle in females which explains the lactational amenorrhea. This mechanism serves as a natural contraceptive and may play a role in spacing out pregnancies. Similarly, prolactin in males inhibits GnRH release resulting in decreased spermatogenesis and infertility.(Raut et al., 2019)(Kumar, 2019) (Štelcletal., 2018).

#### **2.4.4 Function:**

Prolactin function is still being studied, but research seems to show a variety of purposes for this hormone. For instance, it also regulates behavior, the immune system, metabolism, reproductive systems, and many different bodily fluids.

This makes it a crucial hormone for overall health and well-being, for both men and women(Kumar, 2019).

The main two functions of prolactin are to stimulate milk production and to develop breast tissues. Prolactin plays a role in breast development with estrogen and progesterone by stimulating further breast growth and enlargement of the alveoli in preparation for lactation. In addition to breast tissues development, prolactin is an essential player in milk production. Prolactin stimulates milk production by inducing the enzyme that synthesizes the constituents of milk, such as lactose (the carbohydrate of milk), casein (the protein of milk), and lipids. Prolactin is involved in the biosynthesis of milk constituents by binding to the cell membrane and inducing the transcription cascade to make the necessary enzymes for milk production. Lactogenesis does not occur, however; until after parturition because high estrogen and progesterone during pregnancy down regulate prolactin receptors in the breasts. After parturition, the estrogen and progesterone levels fall precipitously. Thus, the inhibitory effects on the breast are removed. As long as suckling is maintained, prolactin level stays elevated after the pregnancy with each episode of feeding producing peak prolactin levels. If the mother does not nurse her baby, prolactin levels fall to non-pregnant levels after 1 to 2 weeks.

(Auriemma et al., 2019)

#### **2.4.5 Reference Range:**

The reference ranges for prolactin in females is as follows:

Adult female: 3-27 ng/ml

Pregnant female: 20-400 ng/mL  
The reference range for prolactin in adult males is 3-13 ng/mL. (Pagana et al., 2019)

#### **2.5.5 Interpretation:**

Hyperprolactinemia is associated primarily with prolactin-secreting pituitary tumors (prolactinoma). Conditions associated with prolactin deficiency include anterior pituitary dysfunction secondary to the following: Postpartum pituitary necrosis (Sheehan syndrome), Pituitary tumor, extrapituitary tumor, Treatment of pituitary/extra pituitary tumor, Parasellar disease, Head injury, Infection (tuberculosis, histoplasmosis) and Infiltrative disease (sarcoidosis, hemochromatosis) (Pagana et al., 2019).

## **2.5 Dyslipidemia and polycystic ovary syndrome.**

Dyslipidemia is common in PCOS characterized by higher triglycerides and lower high density lipoprotein cholesterol. The dyslipidemia occurs independent of body mass index (BMI) however there is a synergistic deleterious effect of obesity and insulin resistance in PCOS analogous to that seen in T2DM. Dyslipidemia in PCOS has multifactorial causation. Insulin resistance plays a pivotal role by stimulation of lipolysis and altered expression of lipoprotein lipase and hepatic lipase. (QilL, 2019).

### **2.5.1 cholesterol:**

Cholesterol is a lipophilic molecule that is essential for human life. It has many roles that contribute to normally functioning cells. For example, cholesterol is an important component of the cell membrane. It contributes to the structural makeup of the membrane as well as modulates its fluidity. Cholesterol functions as a precursor molecule in the synthesis of vitamin D, steroid hormones (e.g., cortisol and aldosterone and adrenal androgens), and sex hormones (e.g., testosterone, estrogens, and progesterone). Cholesterol is also a constituent of bile salt, which is used in digestion to facilitate absorption of fat-soluble vitamins A, D, E, and K. (Di Ciaula et al., 2017).

#### **2.5.1.1 Sources of cholesterol:**

Cholesterol can be introduced into the blood through the digestion of dietary fat via chylomicrons. However, since cholesterol has an important role in cellular function, it can also be directly synthesized by each cell in the body. The synthesis of cholesterol begins from Acetyl-CoA and follows a series of complex reactions that will not be covered in this article. A primary location for this process is the liver. (Asha, 2018).

There are several types of lipoproteins that travel through the blood, and they each have different purposes. There are high-density lipoproteins (HDL), intermediate-density lipoproteins (IDL), low-density lipoproteins (LDL), and very low-density lipoproteins (VLDL). Notably, LDL particles are thought to act as a major transporter of cholesterol, at least two-thirds of circulating cholesterol resides in LDL, to the peripheral tissues. Conversely, HDL molecules are thought to do the opposite. They take excess cholesterol and return it to the liver for excretion. Clinically, these two lipoproteins are significant since high LDL, and low HDL increase a patient's risk of atherosclerotic vascular diseases (Sacksetal., 2017). (Karneyetal., 2017)

#### **2.5.1.2 Functions of cholesterol:**

Within the cell, cholesterol has several vital functions. Some of the primary uses for cholesterol are related to the cell membrane. It is required for the normal structure of the membrane; it contributes to its fluidity. This fluidity can influence the ability of some small molecules to diffuse through the membrane which ultimately changes the internal environment of the cell. Also within the membrane, cholesterol plays a role in intracellular transportation. Beyond its place in the cell membrane, cholesterol has several other biological functions. Of note, cholesterol is known to be an important precursor molecule for the



synthesis of vitamin D, cortisol, aldosterone, progesterone, estrogen, testosterone, bile salts, among others. (Dotson et al., 2017)

### **2.5.1.3 Causes of high cholesterol:**

High cholesterol is a significant risk factor for coronary heart disease and a cause of heart attacks. A build-up of cholesterol is part of the process that narrows arteries, called atherosclerosis. In atherosclerosis, plaques formed cause restriction

of blood flow. Reducing the intake of fat in the diet helps to manage cholesterol levels. In particular, it is helpful to limit foods that contain cholesterol. This is present in animal foods, meat, and cheese. Saturated fat. This occurs in some meats, dairy products, chocolate, baked goods, deep-fried, and processed foods. Trans fats: This occurs in some fried and processed foods. Excess weight or obesity can also lead to higher blood LDL levels. Genetic factors can contribute to high cholesterol. People with the inherited condition familial hypercholesterolemia have very high LDL levels.

Other conditions that can lead to high cholesterol levels, include: diabetes, liver or kidney disease, polycystic ovary syndrome, pregnancy and other conditions that increase levels of female hormones, underactive thyroid gland, drugs that increase LDL cholesterol and decrease HDL (Hammersley et al., 2017).

### **2.5.1.4 Diagnosis of cholesterol:**

A blood test to check cholesterol levels -called a lipid panel or lipid profile- typically reports: Total cholesterol, LDL cholesterol and HDL cholesterol. For the most accurate measurements, don't eat or drink anything (other than water) for nine to 12 hours before the blood sample is taken. (Bethesda, Md., 2019).

According to the 2018 guidelines on the management of blood cholesterol published in the Journal of the American College of Cardiology (JACC), these are the acceptable, borderline, and high measurements for adults.

(American Academy of Pediatrics).

All values are in mg/dL (milligrams per deciliter) and are based on fasting measurement. Total cholesterol HDL cholesterol LDL cholesterol Triglycerides  
Good Less than 200 (but the lower the better) Ideal is 60 or higher; 40 or higher for men and 50 or higher for women is acceptable Less than 100; below 70 if coronary artery disease is present Less than 149; ideal is <100  
Borderline to Moderately elevated 200–239 n/a 130–159  
150–199 High 240 or higher n/a 160 or higher; 190 considered very high  
200 or higher; 500 considered very high Low n/a less than 40 n/a

.(pagana et al., 2019)

## **2.6 triglyceride:**

Triglycerides are a type of fat (lipid) found in your blood. When you eat, your body converts any calories it doesn't need to use right away into triglycerides. The triglycerides are stored in your fat cells. Later, hormones release triglycerides for energy between meals. If you regularly eat more calories than you burn, particularly from high-carbohydrate foods, you may have high triglycerides (hypertriglyceridemia). (Bonow et al., 2019).

### **2.6.2 Normal level of triglyceride:**

Normal: Less than 150 milligrams per deciliter (mg/dL), or less than 1.7 millimoles per liter (mmol/L)

Borderline high: 150 to 199 mg/dL (1.8 to 2.2 mmol/L)

High: 200 to 499 mg/dL (2.3 to 5.6 mmol/L)

Very high: 500 mg/dL or above (5.7 mmol/L or above.

(Kumar p et al., 2018.)

High triglycerides may contribute to hardening of the arteries or thickening of the artery walls (arteriosclerosis) which increases the risk of stroke, heart attack and heart disease. Extremely high triglycerides can also cause acute inflammation of the pancreas (pancreatitis). High triglycerides are often a sign of other conditions that increase the risk of heart disease and stroke, including obesity and metabolic syndrome a cluster of conditions that includes too much fat around the waist, high blood pressure, high triglycerides, high blood sugar and abnormal cholesterol levels. (Mary et al., 2019).

High triglycerides can also be a sign of: Type 2 diabetes or prediabetes

Metabolic syndrome a condition when high blood pressure, obesity and high blood sugar occur together, increasing your risk of heart disease Low levels of thyroid hormones (hypothyroidism) Certain rare genetic conditions that affect how your body converts fat to energy Sometimes high triglycerides are a side effect of taking certain medications, such as: Diuretics ,Estrogen and progestin, Retinoids, Steroids, Betablockers ,Some immunosuppressants and Some HIV medications. (Kumar et al., 2018).

# *Chapter Three*

## *Material and Methodology*

### **3. Material and Methodology**

#### **3.1 study design:**

Case- control (Experimental) hospital base study.

#### **3.2 study area:**

This study carried in in al-Khartoum state during the period from May 2019 to November 2019 in DRALSIR Abo- AlHassan Infertility Center.

#### **3.3 study population and sample size:**

The study include volunteer woman whom have medical report of polycystic ovary syndrome. 50 sample were obtained from patient and 50 from healthy non polycystic over syndrome (control group).

#### **3.4 inclusion and exclusion criteria:**

Samples were collect from woman in reproductive age that inclusion of ultrasonography evidence of poly cystic ovary syndrome as diagnostic marker but exclude pregnant woman, infectious disease, metabolic disease.

#### **3.5 collection sample:**

One hundred(100) Sample were collected by using sterile dry plastic syringe and tourniquet was used to make veins more prominent puncture site was cleaned with 70% ethanol and blood sample (5ml) was collect in plane container from each volunteer.

All blood sample were allowed to collect to clot at room temperature. Then they were centrifuged at 4000rpm to obtain the serum and stored in 20c until analysis.

#### **3.6 Ethical approval:**

The study was approved from clinical chemistry department and medical laboratory science in Sudan University for science and technology.

Prior to the beginning of the study, subject were informed about the protocol of the study and were asked to sign Consent form was taken regarding acceptance, the donor known that this specimen was collected for research purpose.

### **3.7 Methodology:**

The laboratory tests were performed at Khartoum University laboratory, by using enzyme linked immunosorbent assay technique designed for detecting and quantifying hormone which an antigen must be immobilized to a solid surface then complexed with an antibody that is linked to an enzyme.

#### **3.7.1 Prolactin:(immune enzymatic assay)**

##### **3.7.1.1 Principle:**

The essential reagent required for an immune enzymatic assay include high affinity and specific antibodies (enzyme labelled and immobilized) with different and distinct epitopes recognition, in excess, and native antigen. In this procedure, the immobilization takes place during the assay at the surface of a micro plate well through the interaction of streptavidin coated on the well and exogenously added biotinylated monoclonal anti-PRL antibody.

Upon mixing monoclonal biotinylated antibody, the enzyme labeled antibody and serum containing the native antigen, reaction results between the native antigen and the antibodies, without competition or steric hindrance to form a soluble sandwich complex. |

##### **3.7.1.2 Procedure of Measurement (appendix)**

#### **3.7.2 Cholesterol:**

(Enzymatic method)

### 3.7.2.1 Principle of cholesterol oxidase method

serum cholesterol is measured by cholesterol ester +H<sub>2</sub>O **cholesterol oxidase**  
=cholesterol+ fatty acid

cholesterol+O<sub>2</sub> **cholesterol oxidase**= 4-cholesterol-3-one +H<sub>2</sub>O<sub>2</sub>

H<sub>2</sub>O<sub>2</sub>+4 aminophenazone**peroxidase**

= quinone mine +H<sub>2</sub>O.

### 3.7.2.2 Procedure of measurement(appendix)

### 3.7.3 Triglyceride

#### 3.7.3.1Principle:

**Triglyceride** -----lipase =**glycerol +fatty acid**

**Glycerol + ATP** -----glucokinase = **Glycerol 3 phosphate + ADP**

**Glycerol 3 phosphate** +-----Glycerol phosphate oxidase =**Dihydroxyacetone  
-phosphate +H<sub>2</sub>O<sub>2</sub>**

**H<sub>2</sub>O<sub>2</sub> + 4-aminoantipyrin** -----peroxidase =**Quinn amine + HCL +H<sub>2</sub>O.**

#### 3.7.3.2 Procedure of measurement (appendix)

### 3.8 quality control:

The precision and accuracy of all method used in this study were checked by commercially prepared pathologically control sample before the application of tests measurement.

### 3.9 statisticalanalysis

Data was analyzed to obtain mean,standard deviation,and correlation by using statistical package for social science (SPSS)computer programmed version 22,

# *Chapter Four*

## *Results*



#### 4.Results:

The result of biochemical parameter prolactin, cholesterol and triglyceride of poly cystic ovary syndrome women are given in tables and figures.

**Table (4-1):** show frequency of infertile women 17(34%) and unfertile were 33(66%), frequency of therapy uptake woman were 16(32%) and 34(68%) for not therapy uptake.

Also, frequency of cosmetic use were 30(60%) and 20(40%) not use cosmetic.

The frequency of women have history of disease 19(38%) and have not history of PCOS 31(62%).

Frequency analysis of regular menstrual cycle women 28(56%) and irregular 22(44%).

**Table (4-2):** show mean of age  $30\pm 6.59$  and the mean of year's duration was  $2.281\pm 0.78$ .

**Table (4-3):** show significant increase in prolactin, cholesterol and triglyceride with p.value (0.000),(0.009),(0.001) respectively.

**Table (4-3):** show mean of prolactin infertile women ( $34.35\pm 25.5$ ) and unfertile ( $64.592\pm 6.53$ ) with (p.value=0.124) and mean of cholesterol of infertile women  $203.0\pm 36.81$  and unfertile ( $178.8\pm 36.21$ ) with p.value (0.031)

Triglyceride mean with infertility women ( $192.8\pm 36.81$ ) and unfertile  $184.0\pm 34.58$  with p.value (0.430).

**Table (4-4):** mean of the prolactin level take therapy  $44.13\pm 27.79$  and not take therapy  $41.64\pm 26.38$  with p.value (0.761) and cholesterol mean of woman take therapy ( $186.1\pm 38.89$ ) and  $187.5\pm 37.94$  with p.value(0.908)

Triglyceride mean of women take therapy  $200.2 \pm 39.47$  and women not take therapy  $180.8 \pm 34.59$  with p.value( 0.084)

**Table (4-5):** prolactin mean of women use cosmetic  $40.85 \pm 25.21$  and not use  $44.81 \pm 29.01$  with p.( 0.611 ) and cholesterol mean  $185.9 \pm 42.54$  and  $188.9 \pm 30.42$  for cholesterol in women not use cosmetic with p.value( 0.786 )

Triglyceride mean  $193.4 \pm 37.12$  in cosmetic women and  $177.5 \pm 35$  in not use with p.value( 0.137).

**Table (4-6):** mean of prolactin have history of disease  $42.38 \pm$  and  $42.38 \pm 27$  for women have not history with p.value(0.986).

Cholesterol mean  $191.37 \pm 36.11$  and  $184.4 \pm 39.22$  with p.value(0.532)

Triglyceride  $181.2 \pm 32.59$  and  $190.6 \pm 39.49$  with p.value (0.387)

**Table(4-7):** mean of prolactin in regular menstrual cycle  $37.60 \pm 24.72$  and irregular  $48.58 \pm 28.15$  with p.value (0.149)

Mean of triglyceride in regular  $180.6 \pm 38.69$  and irregular  $195.2 \pm 35.98$  with p.value(0.180)

Mean of triglyceride in regular  $185.3 \pm 39.30$  and irregular  $189.13 \pm 34.5$  with p.value( 0.721).

Table(4-1):show the demographic data of the patient of polycystic ovary syndrome.

<b>Variables</b>	<b>Frequency</b>	<b>Percentage (%)</b>
<b>Infertility</b>		
Yes	17	34.0
No	33	66.0
<b>Therapy</b>		
Yes	16	32.0
No	34	68.0
<b>Cosmetic</b>		
Yes	30	60.0
No	20	40.0
<b>History</b>		
Yes	19	38.0
No	31	62.0
<b>Menstrual cycle</b>		
Regular	28	56.0
Irregular	22	44.0
<b>Total</b>	<b>50</b>	<b>100.0</b>

Table(4-2):Mean concentration of patient age and duration of diseases.

Variables	Minimum	Maximum	Mean $\pm$ SD
Age (Years)	19.0	45.0	30.0 $\pm$ 6.59
Duration (Years)	1.00	10.0	2.28 $\pm$ 1.78

Table (4-3): show the Mean  $\pm$ SD of prolactin, cholesterol and triglyceride in patient with control group.

Parameters	Case (Mean $\pm$ SD)	Control (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	42.43 $\pm$ 26.57	25.39 $\pm$ 19.46	0.000
Cholesterol (mg/dl)	187.0 $\pm$ 37.85	152.4 $\pm$ 83.24	0.009
Triglyceride (mg/dl)	187.0 $\pm$ 36.96	149.4 $\pm$ 92.59	0.001

\*Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant.

Table (4-4): show the Mean  $\pm$ SD of prolactin, cholesterol and triglyceride in Infertile and fertile.

Parameters	Yes (Mean $\pm$ SD)	No (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	34.35 $\pm$ 25.50	46.59 $\pm$ 26.53	0.124
Cholesterol (mg/dl)	203.0 $\pm$ 36.81	178.8 $\pm$ 36.21	0.031
Triglyceride (mg/dl)	192.8 $\pm$ 41.71	184.0 $\pm$ 34.58	0.430

Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant.

Table(4-5): show the Mean  $\pm$ SD of prolactin, cholesterol and triglyceride in patient use therapy and non-use.

Parameters	Yes (Mean $\pm$ SD)	No (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	44.13 $\pm$ 27.79	41.64 $\pm$ 26.38	0.761
Cholesterol (mg/dl)	186.1 $\pm$ 38.89	187.5 $\pm$ 37.94	0.908
Triglyceride (mg/dl)	200.2 $\pm$ 39.47	180.8 $\pm$ 34.59	0.084

Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant

Table (4-6) show the Mean  $\pm$ SD of prolactin , cholesterol and triglyceride in patient use Cosmetic and not use.

Parameters	Yes (Mean $\pm$ SD)	No (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	40.85 $\pm$ 25.21	44.81 $\pm$ 29.01	0.611
Cholesterol (mg/dl)	185.8 $\pm$ 42.54	188.9 $\pm$ 30.42	0.786
Triglyceride (mg/dl)	193.4 $\pm$ 37.12	177.5 $\pm$ 35.50	0.137

Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant

Table (4-7): show the Mean  $\pm$ SD of prolactin, cholesterol and triglyceride in patients have history of poly cystic ovary syndrome and not.

Parameters	Yes (Mean $\pm$ SD)	No (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	42.52 $\pm$ 26.08	42.38 $\pm$ 27.31	0.986
Cholesterol (mg/dl)	191.37 $\pm$ 36.11	184.4 $\pm$ 39.22	0.532
Triglyceride (mg/dl)	181.2 $\pm$ 32.59	190.6 $\pm$ 39.49	0.387

Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant

Table (4-8): show the Mean  $\pm$ SD of prolactin, cholesterol and triglyceride in regular menstrual cycle and irregular menstrual cycle patient.

Parameters	R (Mean $\pm$ SD)	IR (Mean $\pm$ SD)	<i>P-value</i>
Prolactin (ng/ml)	37.60 $\pm$ 24.72	48.58 $\pm$ 28.15	0.149
Cholesterol (mg/dl)	180.6 $\pm$ 38.69	195.2 $\pm$ 35.98	0.180
Triglyceride (mg/dl)	185.3 $\pm$ 39.30	189.13 $\pm$ 34.55	0.721

Result given in mean  $\pm$ SD, *p-value* <0.05 consider significant

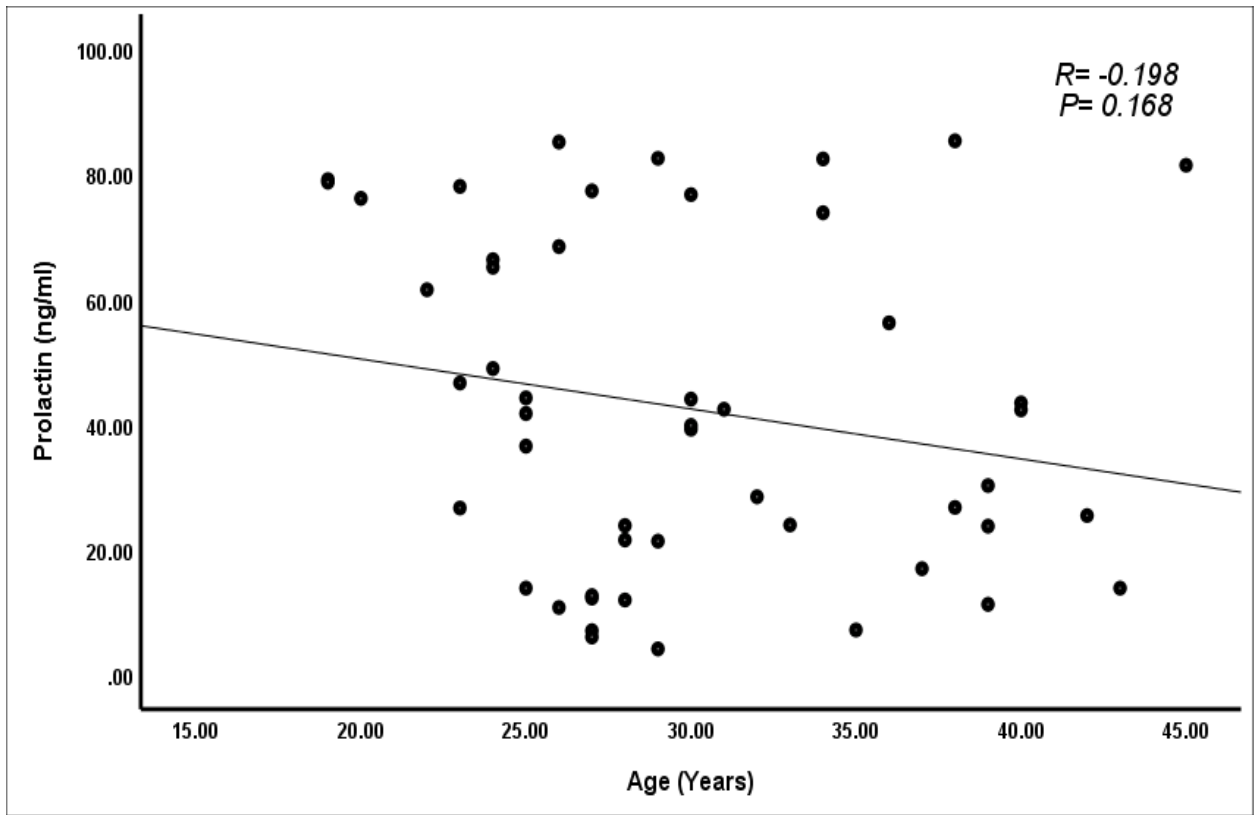


Figure (4-1):scatter plot of pearson correlation between prolactein and age of PCOS woman .

r= -0.198,p.value= 0.168.



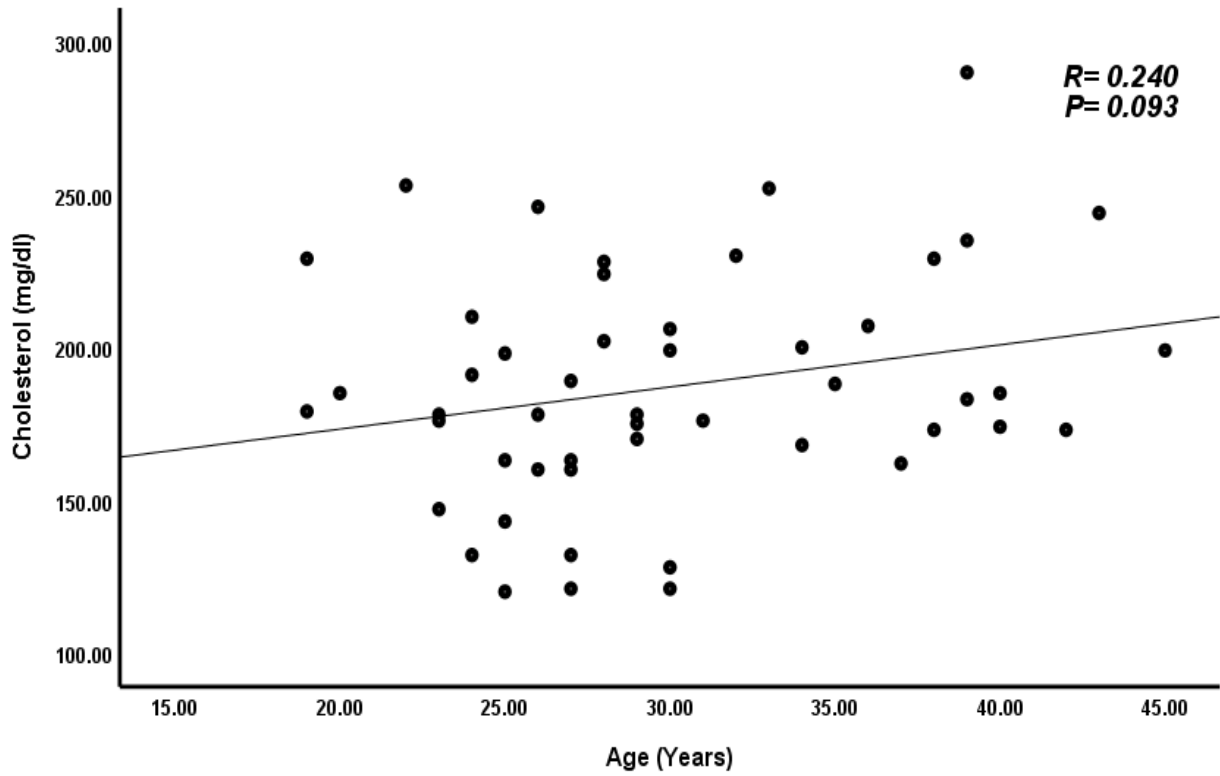


Figure (4-2):scatter plot of pearson correlation between cholesterol and age of PCOS woman .

r= 0.240,p.value= 0.093

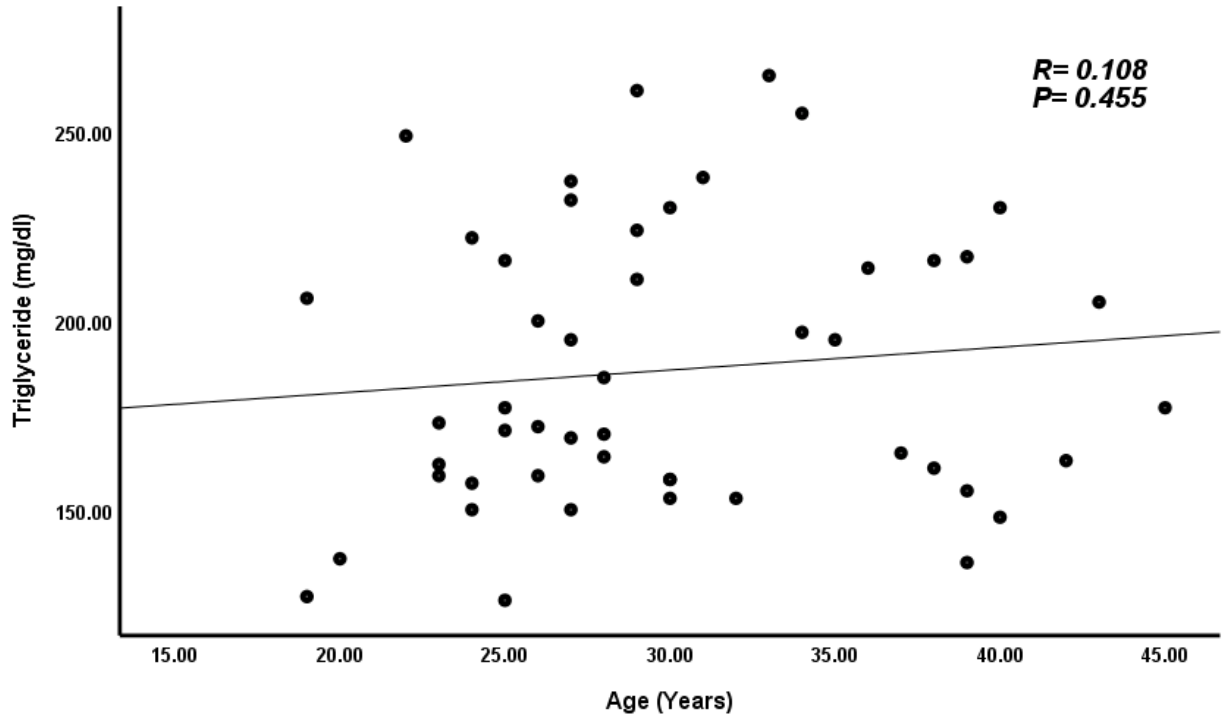


Figure (4-3):scatter plot of pearson correlation between triglyceride and age of PCOS woman .

r= 0.108,p.value= 0.455.

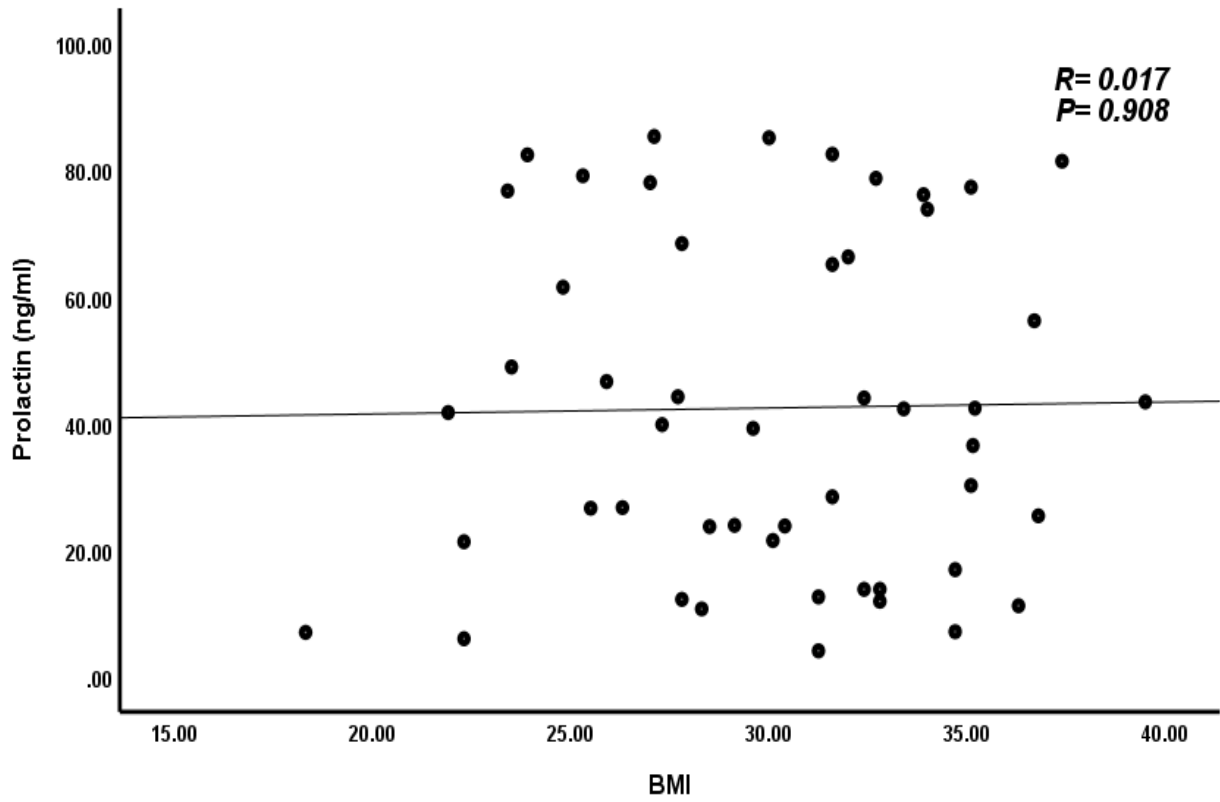


Figure (4-4):scatter plot of pearson correlation between prolactein and BMI of PCOS woman .

r= 0.017,p.value= 0.908 .

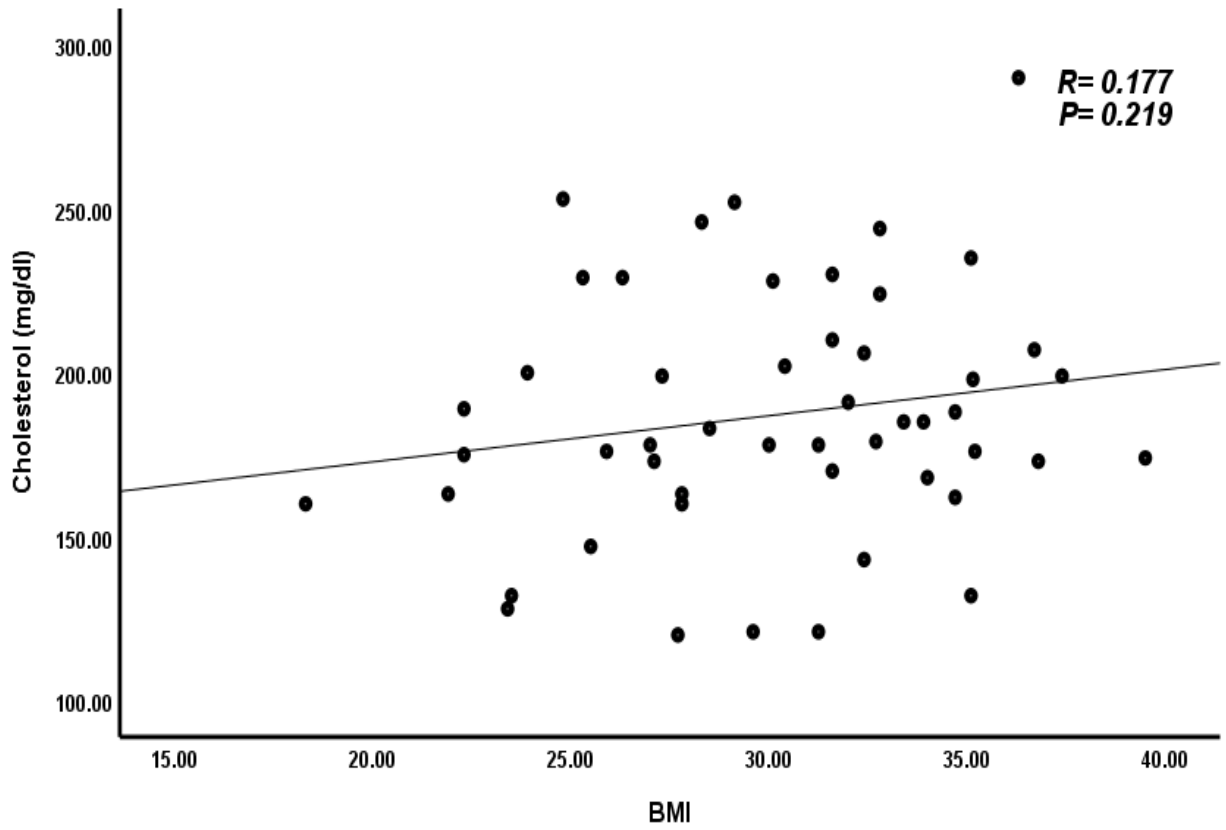


Figure (4-5):scatter plot of pearson correlation between cholesterol and BMI of PCOS woman .

r= 0.177,p.value= 0.219

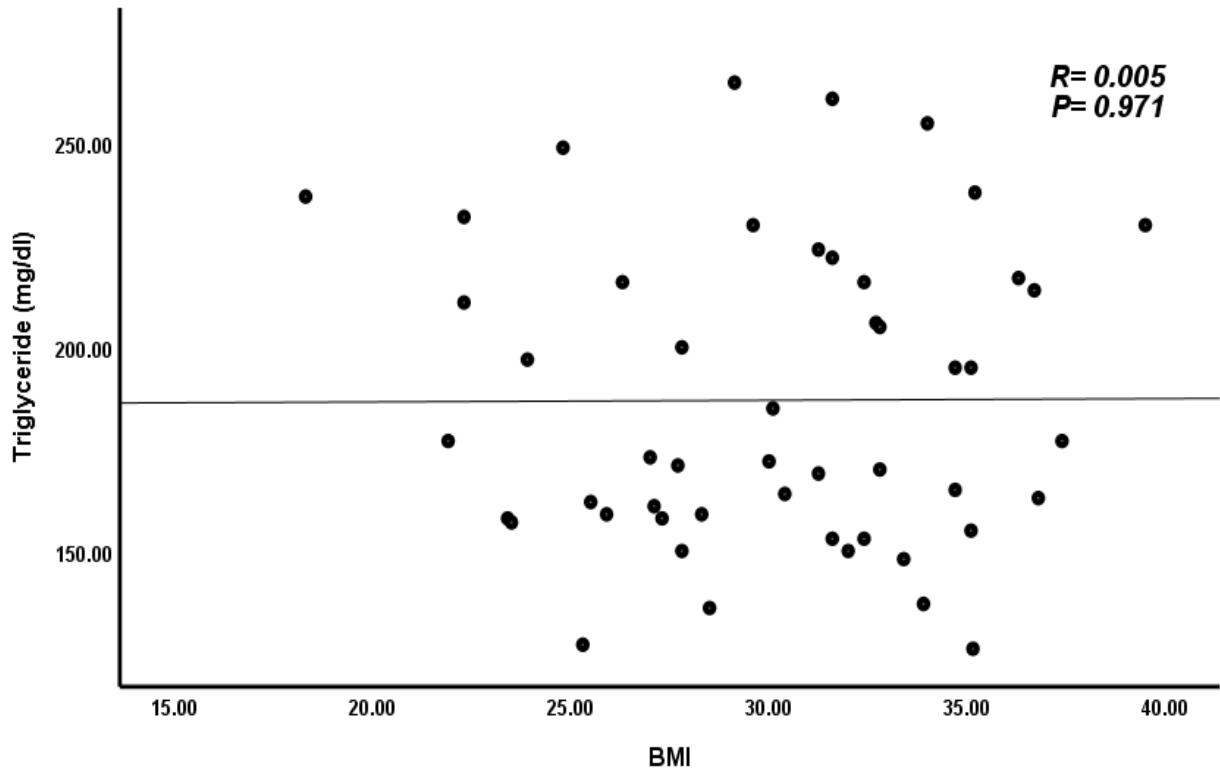


Figure (4-6):scatter plot of pearson correlation between triglyceride and BMI of PCOS woman

$r = 0.005, p. value = 0.971.$

# *Chapter Five*

*Discussion, Conclusion*

## 5.1 Discussion

Poly cystic ovary syndrome is a common disorder affecting women of reproductive age and the most common cause of underlying ovulatory problems. (Azzizet al., 2016).

In this study mean age of the females in the reproductive period was  $30.0 \pm 6.59$  years. The result 50 females with PCOS, the level of prolactin increase significantly with p.value (0.000) compared with control group According to infertility, therapy and cosmetic use, regular of menstrual cycle and history of disease the study showed that prolactin increase insignificant with infertility, therapy, cosmetic and history with (p.value=0.124) (p.value=0.761) (p.value=0.611) (p.value=0.986)

Patients with PCOS can have mildly elevated prolactin; the exact mechanism of hyperprolactinemia in PCOS is unknown. One theory is that constant high levels of estrogen experienced in PCOS would stimulate prolactin production. It is important to rule out other causes of hyperprolactinemia before making the diagnosis of PCOS (Ji Hyun, 2014)

The present study confirms the presence of a more atherogenic lipid profile in woman with PCOS. Results of studies by (Roa et al., 2009) are generally consistent with the results of my study, cholesterol increase significant compared with infertility with (p.value=0.031) and insignificant with therapy (p.value=0.908), cosmetic (p.value=0.786) and menstrual cycle (p.value=0.180).

Triglyceride increased insignificantly with infertility (p.value=0.430), therapy (p.value=0.084) cosmetic (p.value=0.137) and menstrual cycle (p.value=0.721).

that in all, the levels of serum lipids (total cholesterol and triglycerides) in patients with polycystic ovary were higher than healthy persons. However, there

was a weak increase in cholesterol indicates the presence of primary alteration in lipid metabolism in patient with PCOS. The significant increase in triglyceride may be due to increase accumulation of triacylglycerol could as result of increased lipogenesis, decrease clearance or reduced fatty acid oxidation.

Addition, lipid metabolism in women with PCOS may also be affected by ovarian and/or adrenal secretion of sex steroids and obesity. Androgens affect lipids not only directly, but also by affecting obesity, catecholamines, and insulin. (patel, et al 2018).

Hyperandrogenism has been associated with increased hepatic lipase (HL) activity. HL hydrolyses phospholipids on the surface of HDL mediating the conversion of HDL-2 to the smaller denser HDL-3. This being a better substrate for the liver, increases the clearance of HDL. Androgens, through interaction with the androgen receptor, also decrease the catabolic removal of LDL by attenuating estrogen receptor mediated induction of LDL receptor activity.(Gobal et al., 2016)

The present study reveal negative correlation between prolactin level with age ( $r = -0.198, p.value = 0.168$ .) and weakly increase correlation with cholesterol and triglyceride ( $r = 0.240, p.value = 0.093$ ) and positive correlation between triglyceride and age ( $r = 0.108, p.value = 0.455$ ).

The present study show no correlation between prolactin and BMI ( $r = 0.017, p.value = 0.908$ ) but cholesterol positive correlation ( $r = 0.177, p.value = 0.219$ ) and no correlation between triglyceride and BMI ( $r = 0.005, p.value = 0.971$ ).



## **5.2 Conclusion:**

From the result and finding of the study, it is concluded that the serum level of prolactin is high in PCOS females patient, also increased in triglyceride and no observed change in cholesterol level .

## **5.3 Recommendation:**

1. Prolactin hormone must be measurement with other androgenic hormone to exclude misdiagnosis with other disease.
2. Woman with PCOS showed be screened for lipid profile to help in decrease risk of atherosclerosis.
3. Woman must be modified dietary and life .

## References:

A.A. Shaman, H.B. Mukhtar, H.O. Mirghani.(2017). Risk factors associated with metabolic syndrome and cardiovascular disease among women with polycystic ovary syndrome Electron Physician, volume 9, pp. 5697-5704 Cross

Asha Kumari (2018) cholesterol synthesis ,sweet Bio chemisstry.

Auriemma RS, Pirchio R, De Alcubierre D, Pivonello R, Colao A.(2019). Dopamine Agonists. Neuroendocrinology. Volume 109 issue (1)pp34-41.

B. Mlinar, J. Marc, M. Jensterle, E.V. Bokal, A. Jerin, M.Pfeifer (2011) Expression of 11beta-hydroxysteroid dehydrogenase type 1 in visceral and subcutaneous adipose tissues of patients with polycystic ovary syndrome is associated with adiposity , Steroid Biochemistry and Molecular Biology, volume 123, pp. 127-132

Bernard V, Young J, Binart N. Prolactin (2019)a pleiotropic factor in health and disease. Nature Reviews Endocrinology. Volume 15 issue(6):356-365.

Bonow RO, et al.,(2019) Risk markers and the primary prevention of cardiovascular disease. Cardiovascular Medicine. 11th ed.

D. Macut, J. Bjekic-Macut, A.Savic-Radojevic (2013) Dyslipidemia and oxidative stress in PCOS, Frontiers of Hormone Research, volume 40, pp. 51-63.

D.E. Broughton, K.H. Moley(2017), Obesity and female infertility. Fertility and Sterility, volume107 pp.840-847.

Debbie Bridges, MD. (2012), infertility and Reproduction.

Dewailly D, Lujan ME, Carmina E, et al(2014). Definition and significance of polycystic ovarian morphology. Human Reproduction.volume20 issue(3):334–352 .

Di Ciaula A, Garruti G, Lunardi Baccetto R, Molina-Molina E, Bonfrate L, Wang DQ, Portincasa P (2017) Bile Acid Physiology. *Annals of Hepatology*. volume16(Suppl. 1: s3-105.):s4-s14.

Dotson RJ, Smith CR, Bueche K, Angles G, Pias SC. (2017) Influence of Cholesterol on the Oxygen Permeability of Membranes: Biophysical volume 112 issue (11):2336-2347.

E. Seyam, G.S. Al, A.G.A. Abd, M. Mohamed, A.M. Youseff, E.M. Ibrahim, et al (2017). Evaluation of prolonged use of statins on the clinical and biochemical abnormalities and ovulation dysfunction in single young women with polycystic ovary syndrome *Gynecological Endocrinology*. volume 34 issue(7)pp.1-8.

F.R. Tehrani, H. Rashidi, M.B. Khomami, M. Tohidi, F. Azizi(2014) The prevalence of metabolic disorders in various phenotypes of polycystic ovary syndrome. *Biology Endocrinology*, volume 12 , p.89 .

FahimehRamezani ,Tehrani ,samiraBehboudi -gandevani(2015)poly cystic ovary syndrome ,contemporary Gynecologic practice ,chapter 3454.

G.S. Gobl, Ott,L. Bozkurt, M.feichtinger, V.Rehmann, A.cserjan, et al,(2016).The association between glucose metabolism and ectopic lipid content in different clinical classification of PCOS. *Plose One*, 11,Article e 16057.

Ghaffarзад, R. Amani, S.M. Mehrzad, M. Darabi, B. Cheraghian, (2014) Correlation of serum lipoprotein ratios with insulin resistance in infertile women with polycystic ovarian syndrome. *Fertility and Sterility*, volume10 , pp. 29-35.

Hammersley, D., & Signy, M. (2017) Ezetimibe. *Therapeutic Advances in Chronic Diseases*, volume 8 issue (1):p 4–11 .

J. Zhang, J. Hu, C. Zhang, Y. Jiao, X. Kong, W. Wang, (2018) Analyses of risk factors for polycystic ovary syndrome complicated with non alcoholic fatty liver disease. *Experimental and Therapeutic Medicine*, volume 15,pp.4259-4264.

J.X. Pan, Y.J. Tan, F.F. Wang, N.N. Hou, Y.Q. Xiang, J.Y. Zhang, et al(2018). Aberrant expression and DNA methylation of lipid metabolism genes in PCOS., *clinical Epigenetics*. volume10,p.6.

Jameson JL ,et al .(2017)Hyperandrogenism, hirsutism, and poly cystic ovary syndrome ,*Endocrinology* 7<sup>th</sup> edition.

Ji Hyun Chun,PA-c,MPAS,BC-ADM, (2014), Syndrome .*Clinician Reviews* volum 24 issue(2):p26-27 .

Jin Y, Fan M. (2019) Treatment of gynecomastia with prednisone. *International Medical Research*.volume 47 issue (5)p2288-2295.

Joanna Goldberg ,Timjewell (2016) prolactin level test ,*Health care*.

John C. Marshall .MD et al , (2012)ALL Women with PCOS should be treated for insulin resistance. *Fertility and sterility*. volum79 , issue(1) pp18-22.

Jones MR, et al ,(2016)genetic determinants of poly cystic ovary synderome ,*fertility and sterility* volum 106 issue (25).

Karney A, Brągoszewska H, Soluch L, OłtarzewskiM.(2017) [Risk factors for atherosclerosis in obese children aged 6-12 years. *Developmental Period Medicine*. volume 21 issue (3)p259-265.

KovanciE,buster JE. (2015) polycysticovary syndrome. *Clinical gynecology* .second edition.

Kumar MS. (2019) Peptides and Peptidomimetics as Potential Antiobesity Agents, Front Nutrition .volume(6 )issue 11.

Kumar P, et al. (2017) Lipid and metabolic disorders. Clinical Medicine 9<sup>th</sup>

Li H, Huang Y, Li Y, Zheng B, Cui J, Liu M.(2019) Endocrine Manifestations in POEMS syndromes BMC Endocrine Disorders .volume;19 issue (1)p:33.

Lobo RA, et al. (2017) Polycystic ovary syndrome., Comprehensive Gynecology. 7th ed.

M Gibson-Helm et al (2016) Delayed diagnosis and lack of information associated with dissatisfaction in woman with polycystic ovary syndrome. Clinical Endocrinology and metabolism.

M. Spalkowska, S. Mrozinska, A. Galuszka-Bednarczyk, K. Gosztyla, A. Przywara, J. Guzik, et al.(2018) The PCOS patients differ in lipid profile According to their phenotypes. Experimental and Clinical Endocrinology and Diabetes, volume126 pp.437-444.

Mary EllenT ,Sweeney, MD et al (2019) Hypertriglyceridemia, Endocrinology

Matalliotakis M, Koliarakis I, Matalliotaki C, Trivli A, HatzidakiE.(2019) Clinical manifestations, evaluation and management of hyperprolactinemia in adolescent and young girls: a brief review. Acta Biomed .volume 90 issue (1);p149-159.

Milica Popovic Gideon sartorius ,mirjam Christ crain (2019),polycystic ovary syndrome : is there apathophysiological role for interleukin -1?.Seminars in immunopathology volum 41,447-459 .

P.M. Spritzer, S.B. Lecke, F. Satler, D.M. Morsch (2015) Adipose tissue dysfunction, adipokines, and low-grade chronic inflammation in polycystic

ovary syndrome Society for Reproduction, volume 149 issue (5), pp. R219-R227.

Pagana KD, Pagana TJ, Pagana TN. Mosby's (2019) Diagnostic & Laboratory Test Reference. 14th ed.

paganaKD,paganaTJ,paganaTN(2019) Diagnostic and laboratory Test Reference. 14th

QiLiu .et al, (2019) Dyslipidemia involvement in development of poly cystic ovary syndrome. obstetrics and Gynecology, volume 58. issue (4) p 447-453.

R. Azziz, E. Carmina, Z. Chen, A. Dunaif, J.S. Laven, R.S. Legro, et al.(2016) Polycystic ovary syndrome. Nature Reviews Disease Primers, volume 2, p. 16057.

R. Patel, G. Shah,(2018) High-fat diet exposure from pre-pubertal age induces polycystic ovary syndrome (PCOS) in rats, Reproduction, volume 155 , pp. 141-151.

R.wang,(2017), The Rotterdam criteria for poly cystic ovary syndrome .Hum Reprod .volume32 issue (2) p 261-264.

Raut S, Deshpande S, BalasinorNH.(2019) Unveiling the Role of Prolactin and Receptor in Male Reproduction. Hormone and. Metabolic. Research volume 51 issue (4): p215-219.

Roa BM, Arata-Bellabarba G, Valeri L, et al. (2009) [Relationship between the triglyceride/high-density lipoprotein-cholesterol ratio, insulin resistance index and cardiometabolic risk factors in women with polycystic ovary syndrome. Endocrinology Y Nutrition. volume 56 issue (2) p59–65.

Robert Ferry jr, MD.(2012) prolactinoma. Medicine Net

S.L. Manner, H. Leonhardt, J. Kullberg, E. Jennische, N.A. Ode, G.R. Holm, et al (2011) Adipose tissue has aberrant morphology and function in PCOS: Endocrinology, volume 152, p. 332.

Sacks FM, Lichtenstein AH, Wu JHY, Appel LJ, Creager MA, Kris-Etherton et al (2017) Dietary Fats and Cardiovascular Disease. Circulation. volume 136 issue (3)p23.

Selma Feldman, Helena Teede, Alexia s pena (2019) curtailing PCOS,pediatric research volume 87.

Štelcl M, Vrublovský P, Machač Š (2018). Prolactin and alteration of fertility. CeskaGynekol.

V. Pergialiotis, E. Trakakis, C. Chrelias, N. Papantoniou, E. Hatziagelaki,(2018)

The impact of mild hypercholesterolemia on glycemic and hormonal profiles, Hormone Molecular Biology Clinical Investigation , volume 10.

V.de leo , M.C. musacchio, (2016), genetic ,hormonal and metabolic aspect of pcos .Reproductive Biology and Endocrinology ,article number 38 .

Vilar L, Vilar CF, Lyra R, Freitas MDC,(2019) Pit falls in the Diagnostic Evaluation of Hyperprolactinemia. Neuroendocrinology.volume;109issue(1):p7-19.

# *Appendices*



# Appendixes

COD 1180 1 x 20 mL	COD 1180 1 x 200 µL	COD 1180 1 x 20 µL	COD 1120 (x1)
STORE AT 2-8°C			
Reagent for measurement of cholesterol concentration (500 for 0-400 mg/dl in the clinical laboratory)			

**PRINCIPLE OF THE METHOD**  
Free and esterified cholesterol in the serum, triglyceride, by means of the cuprous reaction described below, is oxidized to cholestanol but can be measured by spectrophotometry.

$$\text{Cholesterol} + \text{H}_2\text{O} \xrightarrow{\text{HCl solution}} \text{Cholestanol} + \text{Fatty acid}$$

$$\text{Cholestanol} + \text{S}_2\text{O}_8^{2-} + \text{H}_2\text{O} \xrightarrow{\text{HCl solution}} \text{Cholestanone} + \text{H}_2\text{SO}_4$$

$$2 \text{H}_2\text{O} + \text{S} \xrightarrow{\text{Antimonydioxide + Phosphoric}} \text{Glycerol} + 4 \text{H}_2\text{SO}_4$$

**CONTENTS**

	COD 1180	COD 1180	COD 1180	COD 1120
A. Reagent	1.120 mL	1 x 200 µL	1 x 20 µL	1 x 1
B. Standard	1.120 mL	1 x 10 µL	1.0 µL	1 x 10 µL

**COMPOSITION**  
A. Reagent: Phos. 20 mmol/L, sodium citrate 0.8 mmol/L, phos. 20 mmol/L, cholesterol oxime = 0.3 mg/L, potassium iodate = 0.1 mg/L, potassium = 0.8 mg/L, 2-mercaptoethanol 0.5 mmol/L, pH 7.2  
B. Cholesterol Standard: Cholesterol 200 mg/dL (5.18 mmol/L) - reference primary standard

**STORAGE**  
Store at 2-8°C.  
Reagent and Standard are stable until the expiry date shown on the label when stored tightly closed and if not contaminated are protected during their use.  
Indications of deterioration:  
- Reagent: Presence of particulate material, turbidity, absorption of the blank over 2.00 at 500 nm (1 cm cuvette)  
- Standard: Presence of particulate material, turbidity

**REAGENT PREPARATION**  
Reagent and Standard are provided ready to use.

**ADDITIONAL EQUIPMENT**  
- Thermobath water bath at 37°C  
- Analytic, spectrophotometer or photometer able to read at 500 ± 20 nm

**SAMPLES**  
Serum or plasma collected by standard procedures.  
Cholesterol is stable for 7 days at 2-8°C. Heparin, EDTA, oxalate and fluoride may be used as anticoagulants.

**PROCEDURE**

- Bring the Reagent to room temperature.
- Pipette six labelled test tubes (Table 1)

	Blank	Standard	Sample
Cholesterol Standard (B)	---	10 µL	---
Sample	---	---	10 µL
Reagent (A)	1.0 mL	1.0 mL	1.0 mL

- Mix thoroughly and incubate the tubes for 10 minutes at room temperature (16-20°C) or in a thermostat at 37°C.
- Measure the absorbance (A) of the Standard and Sample at 500 nm against the blank. The colour is stable for at least 15 minutes.

**CALCULATIONS**  
The cholesterol concentration in the sample is calculated using the following general formula:

$$\frac{A_{\text{sample}}}{A_{\text{blank}}} = \frac{C_{\text{sample}}}{C_{\text{blank}}}$$

If the Cholesterol Standard provided has been used to calibrate (Table 2):

$\frac{A_{\text{sample}}}{A_{\text{blank}}}$	$\frac{1.00 \pm \text{mg/dL cholesterol}}{1.0 (10 \pm \text{mg/dL cholesterol})}$
--	---

**REFERENCE VALUES**  
The following reference cut-off points have been established by the WHO National Cholesterol Education Program and have also been adopted in many other countries for the evaluation of coronary artery disease risk:

Up to 200 mg/dL = 5.2 mmol/L 200-239 mg/dL = 5.2-6.2 mmol/L 240 mg/dL = 6.2 mmol/L	Borderline to High Risk
--	-------------------------------

**CHOLESTEROL**

**CHOLESTEROL**  
CHOLESTEROL, CHOLESTEROL, CHOLESTEROL

**QUALITY CONTROL**  
It is recommended to use the Biosystems Control Department (cods 10002, 10003 and 10004) and if used 10001, 10010 and 10015 to verify the performance of the measurement procedure. Each laboratory should establish its own internal Quality Control scheme and procedures for corrective action if possible. It is always better to measure within the reference interval.

**METROLOGICAL CHARACTERISTICS**

- Correlation: 0.99 (2.2 mg/dL, 0.57 mmol/L)
- Linearity: 0-400 mg/dL = 0-10 mmol/L. For higher values, dilute samples 10 with distilled water and repeat measurement.
- Reproducibility: within-run

Mean Cholesterol	CV	n
50 mg/dL = 1.36 mmol/L	1.7%	20
200 mg/dL = 5.18 mmol/L	0.9%	20

- Reproducibility: day-to-day

Mean Cholesterol	CV	n
100 mg/dL = 2.6 mmol/L	1.9%	20
200 mg/dL = 5.18 mmol/L	1.0%	20

- Precision: Results obtained with this reagent are not more variable than those obtained when compared with reference reagents (Table 2). Details of the comparison experiments are available on request.
- Interference: Specificity: Interference: 10 µL of each of the following: Bile acids (1.0 mg/dL) and triglycerides (1.0 g/dL) has no effect on results. Other drugs and substances may interfere.

These metrological characteristics have been obtained using an analyzer. Results may vary if a different instrument or a different procedure are used.

**DIAGNOSTIC CHARACTERISTICS**  
Cholesterol is a class of high molecular weight and possesses the sphingosterol character. Delayed absorption is partially absorbed and it is slow excreted by the liver and other tissues. Cholesterol is transported in plasma by lipoproteins. It is oxidized and degraded into bile or after transformation in bile acids.

Increased total cholesterol values are associated with a progressively increasing risk of atherosclerosis and coronary artery disease.  
Clinical diagnosis should not be based on the findings of a single test result, but should include both clinical and laboratory data.

**NOTES**

- This reagent may be used in several automatic analyzers. Instructions for many of them are available on request.
- Calculation with the provided reagent constant may cause a small error, specially in some analyzers. In Polar units, it is recommended to calibrate using a serum based standard (Biochemistry Calibration cods 10011 and 10044).

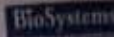
**BIBLIOGRAPHY**

- Allen CC, Pevell LG, Gian CSE, Richmond W and Fu PC. Spectrometric determination of total serum cholesterol. Clin Chem 1976; 22: 470-473.
- Mustafa F, Pevell L, Serradi F, Giamali G and Taki P. The 4-hydroxycholesterol-3-oxocholesterol chromatographic system used in the enzymic determination of serum cholesterol. Clin Chem 1975; 21: 2181-2185.
- National Cholesterol Education Program Expert Panel. Third report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (ATP III). BMJ Publishing, Statistics, National Heart, Lung, and Blood Institute, 2001.
- Young DS. Effects of drugs on clinical laboratory tests, 2nd ed. AACCC Press, 2000.
- Textbook of Clinical Chemistry and Laboratory Diagnostics, 4th ed. Burtis CA, Bruns DE, Tietz NC, Saunders CV, 2002.
- Friedman and Young. Effects of disease on clinical laboratory tests, 4th ed. AACCC Press, 2001.

W1220-02

**Biosystems S.A. Costa Brava, 30. 08030 Barcelona (Spain)**  
Quality System certified according to  
ISO 13485 and ISO 9001 standards

TRIGLYCERIDES

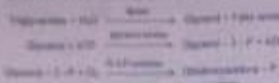


TRIGLYCERIDES  
GLYCEROL PHOSPHATE OXIDASE/PEROXIDASE

CODE 1104	CODE 1105	CODE 1106
1 x 50 ml	4 x 50 ml	1 x 250 ml
01/788 at 20°C		
Requests for measurements of triglyceride concentration may be made up to the stated validity		

PRINCIPLE OF THE METHOD

Triglycerides in the sample, diglycerides, by means of the enzyme reaction described herein, a colorless complex that can be measured by spectrophotometry.



CONTENTS

	CODE 1104	CODE 1105	CODE 1106
1. Reagent	1 x 50 ml	4 x 50 ml	1 x 250 ml
2. Standard	1 x 50 ml	1 x 50 ml	1 x 50 ml

COMPOSITION

- Reagent: Phos. 40 mmol/L, Magnesium chloride 5 mmol/L, 4-aminophenol 8 mmol/L, NaOH 1.00 mmol/L, glycerol kinase 1.15 U/L, glycerol phosphate oxidase 4 U/L, peroxidase 0.2 U/L, 4-aminosalicylic acid 0.75 mmol/L, ATP 0.3 mmol/L, pH 7.5.
- Triglyceride Standard: Glycerol equivalent to 200 mg/dl (2.26 mmol/L) stearin. Assay as primary standard.

STORAGE

Store at 2-8°C.

Reagent and Standard are stable with the expiry date shown on the label when stored tightly closed and protected from light and when the instructions are followed during their use.

Indications of deterioration:

- Reagent: Presence of particulate material, turbidity, discoloration of the fluid over 1:100 at 300 nm (1 cm cuvette).
- Standard: Presence of particulate material, turbidity.

REAGENT PREPARATION

Reagent and Standard are provided ready to use.

ADDITIONAL EQUIPMENT

- Thermometer with bath at 37°C.
- Analyzer, spectrophotometer or photometer able to read at 300 or 20 nm.

SAMPLES

Serum or plasma collected by standard procedures.

Triglycerides in serum or plasma are stable for 8 days at 2-8°C. Heparin, EDTA, oxalate and fluoride may be used as anticoagulants.

PROCEDURE

- Bring the Reagent to room temperature.
- Pipette into labeled test tubes (Note 1).

	Blank	Standard	Sample
Triglyceride Standard (2)	---	10 µl	---
Sample	---	---	20 µl
Reagent (1)	1.0 ml	1.0 ml	1.0 ml

- Mix thoroughly and incubate the tubes for 10 minutes at room temperature (16-20°C) or for 5 minutes at 37°C.
- Measure the absorbance (A) of the Standard and Sample at 300 nm against the Blank. The assay is stable for at least 2 hours.

CALCULATIONS

The triglyceride concentration in the sample is calculated using the following general formula:

$$\frac{A_{\text{Sample}}}{A_{\text{Standard}}} \times C_{\text{Standard}} \times F \times \text{Dilution}$$

If the Triglyceride Standard provided has been used to calibrate (Note 2)

$\frac{A_{\text{Sample}}}{A_{\text{Standard}}}$	$\times 200 \times \text{mg/dl Triglycerides}$
	$\times 2.26 \times \text{mmol/l Triglycerides}$

REFERENCE VALUES

The following within-unit limits have been established by the ISO National Institute of Health and have also been adopted by many other countries for the evaluation of lipid.

10-150 mg/dl = 0.33 mmol/L 100-200 mg/dl = 0.66 mmol/L 200-300 mg/dl = 1.32 mmol/L > 300 mg/dl = > 1.32 mmol/L	Normal Borderline High Very High
---	---

QUALITY CONTROL

It is recommended to use the Biochemistry Control Serum level 1200, 1800, 1800 and 18042 and 71 (code 1800, 1805 and 1806) to verify the performance of the measurement procedure.

Each laboratory should establish its own internal Quality Control system and procedures to determine when it controls do not measure within the acceptable tolerance.

METROLOGICAL CHARACTERISTICS

- Detection limit: 0.2 mg/dl = 2.26 µmol/L.
- Linearity limit: 600 mg/dl = 6.78 mmol/L. For higher values other sample or a well diluted water and repeat measurement.
- Repeatability (within-run):

Mean Concentration	CV	%
100 mg/dl = 1.13 mmol/L	1.7%	26
300 mg/dl = 3.39 mmol/L	0.7%	26

- Reproducibility (day to day):

Mean Concentration	CV	%
100 mg/dl = 1.13 mmol/L	2.2%	26
300 mg/dl = 3.39 mmol/L	1.7%	26

- Trueness: Results obtained with this reagent did not show systematic differences when compared with reference reagents (Note 2). Details of the comparison experiments are available on request.
- Interference: Phenolphthalein (10 g/L) does not interfere. Bilirubin (2.5 mg/dl) may interfere (Other drugs and substances may interfere).

These metrological characteristics have been obtained using an analyzer. Results may vary if a different instrument or a manual procedure are used.

DIAGNOSTIC CHARACTERISTICS

Triglycerides are esters of glycerol and fatty acids coming from the diet or obtained by synthesis mainly in the liver. Triglycerides are transported in plasma by lipoproteins and used by adipose tissue, muscle and liver. Their primary function is to provide energy in the cell.

Elevated serum triglycerides levels can be caused by liver disease, diabetes mellitus, nephrosis, hypothyroidism, acromegaly, familial hyperlipoproteinemia II and V, and others.

Clinical diagnosis should not be made on the basis of a single test result, but should integrate both clinical and laboratory data.

NOTES

- This reagent may be used in several automatic analyzers. Instructions for many of them are available on request.
- Calibration with the provided aqueous standard may cause a matrix related bias, specially in some analyzers. In those cases, it is recommended to calibrate using a serum based standard (Biochemistry Calibrator, code 18011 and 18044).

BIBLIOGRAPHY

- Scott G and David H. Quantitative determination of serum triglycerides by use of enzymes. Clin Chem 1972; 18: 479-482.
- Fossati P and Principe L. Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. Clin Chem 1982; 28: 2017-2020.
- National Cholesterol Education Program Expert Panel. Third report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (ATP III). NIH Publication, Bethesda: National Heart, Lung, and Blood Institute, 2001.
- Young DS. Effects of drugs on clinical laboratory tests. 3th ed. AACC Press, 2000.
- Friedman and Young. Effects of disease on clinical laboratory tests. 4th ed. AACC Press, 2001.