بسم الله الرحمن الرحيم

صدق الله العظيم سورة الاسراء الاية 85

Dedication

I dedicate this work to

My Parents.....

WHO DID EVERY THINGS FOR ME

My brothers and sisters....

WHO WERE ALWAYS THERE ON MY SIDE

My friends and my colleagues.....

To all who has ever taught me anything

ACKNOWLEDGEMENTS

Firstly, thanks to Almighty ALLAH for giving me patience, health to complete this work.

Secondly, I would like to express my gratitude and appreciation to my supervisor Dr. Mogahid Elhassan, for his invaluable help, encouragement and guidance through all the study.

Thirdly, I would like to Acknowledge Prof. Awad Elseed Mustafa for his care, support, advice and encouragement.

Thirdly, my thanks and gratitude is extended Dr. Somaia Elhag, Mrs. Zakia Yousif, Mr. Baha Eldeen Hassen, Hanan Ahmed Abdella and Mr. Salah Eldein Elzaki Gomaa for their unlimited support and efforts.

My thanks are extended to the staff members of Wad Medani Teaching hospitals, Gynaecology and Obstetrics Teaching Hospital, Institute of Nuclear Medicine, Moleculour Biology and Oncology, Orthopaedic surgery hospital, Dr. Ahmed Abdella Hamadein Hospital for Dermal Diseases, Gezira Hospital for Renal Diseases and Surgery, National Centre of Paediatrics Surgery, Tahily Speciality Hospital, Alyaa Speciality Hospital and Police Hospital for their collaboration.

Lastly, I would like to declare my deep thanks to my colleagues in the research laboratory at Sudan University of Science and Technology and Medical Laboratory at Gezira University, especially; Mrs. Maimona Ahmed, Suhair Rehan, Fadwa Nor Elhadi, Samar Ahmed Elagab, Sara Abo Edrees and Somia Othman for their technical support.

ABSTRACT

Methicillin resistant Staphylococcus aureus (MRSA) is an increasing problem worldwide while in Sudan we still lacking the preliminary data for this pathogen. The objectives of the present study were to provide evidences about the occurrence of MRSA among Sudanese patients as well as to characterize and amplify different genes responsible for this phenomenon. Three hundred and fifty five patients suffering from different clinical diseases (n=355) were included in this study during the period from October 2010 to May 2011. The distribution of the enrolled patients was as follows: general surgery at Wad Medani Teaching Hospital 129/355(36.3%), Gynaecology and Obstetrics Teaching Hospital 107/355 (30.1%), Institute of Nuclear Medicine, Moleculour Biology and Oncology 48/355 (13.5%), Orthopaedic Surgery Hospital 8/355 (2.3%), Dr. Ahmed Abdella Hamadein Hospital for Dermal Diseases 23/355 (6.5%), Gezira Hospital for Renal Diseases and Surgery, National Centre of Paediatrics Surgery 7/355 (2%), Tahily Speciality Hospital 7/355 (2%), Alyaa Speciality Hospital 8/355 and Wad Medani Police Hospital 10/355 (2.8%). All clinical (2.3%),samples were cultured on blood agar and MacConkey agar. Different biochemical tests, Gram's stain and amplification of arcC gene were used to identify the causative pathogens. The results confirmed the existence of Staphylococcus aureus in seventy two (72);(20.3%) of the enrolled patients among which thirty three (33); (45.8%) methicillin resistant Staphylococcus aureus were identified when using modified Kirby Bauer method. The distribution of MRSA among enrolled patients at the hospitals and medical centers were as follows: general surgery at Wad Medani Teaching Hospital

19/72 (26.4%), Gynaecology and Obstetrics Teaching Hospital 5/72 (7%), Orthopaedic Surgery Hospital 1/72 (1.4%), Dr. Ahmed Abdella Hamadein Hospital for Dermal Diseases 2/72 (2.8%) and Institute of Nuclear Medicine, Moleculour Biology and Oncology 6/72 (8.3%). While other medical centers included in the study were appeared free from MRSA.

Confirmation of the results of methicillin disk diffusion Kirby Bauer method was conducted by amplifying *mecA* gene. Furthermore all MRSA isolates were tested against other empirical antibiotics, the results were as follows: 14/33 (42.40%) were resistant to co-trimoxazole, 15/33 (45.50%) were resistant to cephalexin, 22 /33 (66.70%) were resist to tetracycline, 20 /33 (60.60%) were resistant to cefotaxime, 6/33 (18.20%) were resistant to ciprofloxacin and erythromycin, 12 /33 (36.40%) were resistant to pefloxacin and ofloxacin, 33/33 (100%) were resistant to cloxacillin, 7/33 (21.20%) were resistant to clindamycin, 3 /33 (9.10%) were resistant to gentamycin, 30/33 (90.90%) were resistant to ceftriaxone, and 9/33 (27.30%) were resistant to cefuroxime.

All MRSA isolates were examined against vancomycin antibiotic using modified Kirby Bauer disk diffusion method. The results obtained excluded the existent of VRSA among all MRSA isolates. More confirmation was adopted by amplifying *Van A*, *Van B* genes.

The study concluded that MRSA still consider as a great in medical field. Also it confirmed the sensitivity of molecular method in the diagnosis of MRSA as well as VRSA among infected patients. Thus, it can substitute the long conventional methods.

مستخلص الأطروحة

تعتبر المكورات العنقودية الذهبية المقاومة للميثيسيلين مشكلة متفاقمة في جميع أنحاء العالم بينما في السودان لا نزال نفتقر إلى البيانات الأولية لهذا الكائن الممرض.

هدفت هذه الدراسة إلى تقديم أدلة عن تواجد المكورات العنقودية الذهبية المقاومة للمثيسيلين بين المرضى السودانيين وكذلك معرفة خصائصه وتضخيم الجينات المختلفة المسؤولة عن هذه الظاهرة.

ثلاثمائة وخمسة وخمسون مريضا يعانون من أمراض سريرية مختلفة (عدد = 355) تم تضمينها في هذه الدراسة خلال الفترة من أكتوبر 2010 إلى مايو 2011. وتوزيع المرضى المسجلين على النحو التالي: الجراحة العامة بمستشفى ود مدنى التعليمي 129 /355 (36.3٪)، المستشفى التعليمي لأمراض النساء والتوليد 355/107 (30.1) ومعهد الطب النووي والأحياء الجزيئية والأورام 355/48 (13.5٪)، مستشفى جراحة العظام 355/8 (2.3٪)، ومستشفى الدكتور أحمد عبد الله حمدين للأمراض الجلدية 355/23 (6.5%)، مستشفى الجزيرة لأمراض وجراحة الكلى ، المركز القومي لجراحة الأطفال 355/7 (2%)، المستشفى التأهيلي التخصصي 355/7 (2%)، مستشفى علياء التخصصي 355/8 (2.3٪)، ومستشفى الشرطة 355/10 (2.8٪). تم زراعة جميع العينات السريرية على آجار الدم وآجار الماكونكي. وعمل الاختبارات البيوكيميائية المختلفة، كما استخدمت صبغة جرام وتم الكشف عن الجين arcc بالتضخيم والبلمرة لتحديد مسببات المرض. أكدت النتائج وجود المكورات العنقودية الذهبية في 72 (72، 20.3٪) من المرضى المسجلين، ثم تم الكشف عن وجود المكورات العنقودية الذهبية المقاومة للميثيسيلين باستخدام طريقة الإنتشار القرصى باور كيربي (لمعرفة تحسس الجراثيم للمضادات الحيوية). عددها ثلاثة والثلاثين (33، 45.8٪). بينما كان توزيع المكورات العنقودية الذهبية المقاومة للميثيسيلين بين المرضى المسجلين في المستشفيات والمراكز الطبية على النحو التالي: جراحة العامة بمستشفى ود مدنى التعليمي 72/19 (26.4٪)، المستشفى التعليمي لأمراض النساء والتوليد 5 / 72 (7٪)، مستشفى جراحة العظام 72/1 (1.4٪)، ومستشفى الدكتور أحمد عبد الله حمدين للأمراض الجلدية 72/2 (2.8٪)، ومعهد الطب النووي الأحياء الجزيئية والأورام 72/6 (8.3٪) . والمراكز الخالية من المكورات العنقودية الذهبية المقاومة للميثيسيلين كانت: المركز القومي لجراحة الأطفال، مستشفى علياء التخصصي، مستشفى الشرطة، مستشفى التأهيلي التخصصي ومستشفى الجزيرة لأمراض وجراحة الكلي.

نتائج طريقة الإنتشار القرصي باور كيربي (لمعرفة تحسس الجراثيم للمضادات الحيوية) للميثيسيلين تم تأكيدها بتضخيم الجين mecA. ثم تم اختبار جميع العزلات (المكورات العنقودية الذهبية المقاومة للميثيسيلين) لمعرفة مقاومتها للمضادات الحيوية المستخدمة الأخرى، وقد كانت النتائج على النحو التالي: 33/14 (42.40) كانت مقاومة للكوتريموكسازول، و33/15 (60.60)) مقاومة للسيفاليكسين، و 33/26 (60.60)) مقاومة للسيفوتاكسيم، 33/26 (60.60)) كانت مقاومة للسيبروفلوكساسين والاريثرومايسين، 33/12 (100 ٪) كانت مقاومة للكلوكساسيلين، و 33/33 (100 ٪) كانت مقاومة للكلينداميسين، و 33/33 (100 ٪) كانت مقاومة للجنتاميسين، و 33/33 (20 ٪) كانت مقاومة للطبنداميسين، و 33/33 (20 ٪) كانت مقاومة للجنتاميسين، و 33/33 (20 ٪) كانت مقاومة للسيفترياكسون، و 33/33 (20 ٪) كانت مقاومة للسيفروكسيم.

تم اختبار جميع العزلات للمضادات الحيوية المكورات العنقودية الذهبية المقاومة للميثيسيلين ضد الفانكوميسين باستخدام طريقة الإنتشار القرصي باور كيربي (لمعرفة تحسس الجراثيم للمضادات الحيوية). النتائج التي تم الحصول عليها دلت على خلو جميع عزلات المكورات العنقودية الذهبية المقاومة للميثيسيلين من وجود المكورات العنقودية الذهبية المقاومة للفانكومايسين. و للتأكيد تم الإعتماد على طريقة التضخيم والبلمرة للجينات Van A, Van B.

الدراسة أكدت ان المكورات العنقودية الذهبية المقاومة للميثيسيلين لاتزال تحد كبير في الحقل الطبي. كما أكدت الدراسة ايضا حساسية الطريقة الجزيئية في الكشف عن المكورات العنقودية الذهبية المقاومة للميثيسيلين وايضا المكورات العنقودية الذهبية المقاومة للفانكومايسين على حد سواء بين المرضى المصابين. عليه، يمكن استبدالها عن الطريقة التقليدية الطويلة.

List of Contents

Contents	No. of
	page
الآية القرآنية	i
Dedication	ii
Acknowledgment	iii
Abstract (in English)	iv
Abstract (in Arabic)	vi
List of contents	viii
List of Tables	XV
List of Figures	xvi
List of Abbreviations	xviii
CHAPTER ONE: INTRODUCTION AND OBJECTIVES	1
1.1. Introduction	1
1.2. Rationale	4
1.3. Objectives	5
1.3.1. General Objective	5
1.3.2. Specific Objectives	5
CHAPTER TWO: LITERATURE REVIEW	6

2.1. Definition	6
2.2. Historical Background	6
2.3 Taxonomy	7
2.4 Morphology and Cultural Characters	7
2.5 Biochemical Characters	8
2.6 Pathogenicity	8
2.6.1 Virulence factors	9
2.6.2 Structural components	9
2.6.2.1 Capsule	9
2.6.2.2 Peptidoglycan and Cytoplasmic Membrane	10
2.6.2.3 Teichoic Acids	10
2.6.2.4 Protein A	10
2.6.2.5 Leukocidin	10
2.6.3 Enzymes Production	11
2.6.3.1 Coagulase	11
2.6.3.2 Catalase	11
2.6.3.3 Hyaluronidase	12
2.6.3.4 lipases	12
2.6.3.5 nuclease	12

2.6.3.6 Staphylokinase	12
2.6.3.7 penicillinase	13
2.6.4. Toxins production	13
2.6.4.1 Alpha Toxin	13
2.6.4.2 Beta toxin	13
2.6.4.3 Delta toxin	14
2.6.4.4 Gama toxin and leukocidin	14
2.6.4.5 Panton-Valentine leukocidin (PVL)	14
2.6.4.6 Exfoliative toxins	15
2.6.4.7Superantigens: enterotoxins and toxic shock syndrome toxin	15
2.6.5. Clinical Infections	16
2.6.5.1 Skin Disease	16
2.6.5.1.1 Wound Infections and Abscesses	16
2.6.5.1.2 Impetigo	17
2.6.5.1.3 Folliculitis	17
2.6.5.1.4 Carbuncles	17
2.6.5.1.5 Cellulitis	17
2.6.5.2 Bacteremia	17
2.6.5.3 Endocarditis	18

2.6.5.4 Pneumonia	18
2.6.5.5 Osteomyelitis	18
2.6.5.6 Urinary Tract Infections	19
2.6.5.7 Food poisoning	19
2.6.5.8 Enterocolitis	19
2.6.5.9 Toxic Shock Syndrome	20
2.6.5.10 Scalded skin syndrome	20
2.6.6. Community and Hospital Associated Infections	21
2.7. Epidemiology	22
2.8. Treatment and Control	23
2.8.1 MRSA	25
2.0.2 LIDGA	
2.8.2 VRSA	27
CHAPTER THREE: MATERIALS AND METHODS	30
CHAPTER THREE: MATERIALS AND METHODS	30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design	30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study	30 30 30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study 3.1.2. Study Area	30 30 30 30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study 3.1.2. Study Area 3.1.3. Sample Size and Study Population	30 30 30 30 30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study 3.1.2. Study Area 3.1.3. Sample Size and Study Population 3.1.4. Ethical Clearance	30 30 30 30 30 30
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study 3.1.2. Study Area 3.1.3. Sample Size and Study Population 3.1.4. Ethical Clearance 3.2 Methodology	30 30 30 30 30 30 31
CHAPTER THREE: MATERIALS AND METHODS 3.1. Study Design 3.1.1. Type of the study 3.1.2. Study Area 3.1.3. Sample Size and Study Population 3.1.4. Ethical Clearance 3.2 Methodology 3.2.1Data Collection	30 30 30 30 30 30 31 31

3.2.3.1 Dry heat	32
3.2.3.2 Moist heat	32
3.2.3.3 Control of sterilization	33
3.2.4 Phenotypic Characterization of the Isolates	33
3.2.4.1 Culture Media	33
3.2.4.2 Colonial Characteristics	33
3.2.4.3 Morphological Characteristics	34
3.2.4.4 Biochemical Tests for the Identification of Bacterial	34
Isolates	
3.2.4.4.1 Catalase Test	34
3.2.4.4.2 Coagulase Test	34
3.2.4.4.3 DNase Test	35
3.2.4.5 Mannitol Salt Agar	35
3.2.4.6 Susceptibility of Isolated Bacteria to Different Antibiotics	35
2.2.4.6.1 Mueller Hinton Agar	35
3.2.4.6.2 Turbidity standard equivalent to McFarland 0.5	36
3.2.4.6.3 Preparation of Inoculum and Inoculation	36
3.2.4.6.4 Application of Sensitivity Discs	36
3.2.4.6.5 Interpretation of Zone Sizes	37
3.2.5 Preservation of Isolated Organism	38
3.2.6 Molecular Analysis by Using PCR Method	38
3.2.6.1 DNA Extraction	40
3.2.6.2 Measurement the Concentration of DNA by DNA Reader	40
3.2.6.3 Oligonucleotide Primers	40
3.2.6.4 Master Mix	40

3.2.6.5 PCR Amplification	40
3.2.6.5.1 <i>arcC</i> Gene	41
3.2.6.5.2 <i>mecA</i> Gene	41
3.2.6.5.3 <i>Van</i> A1 and B1Genes	41
3.2.6.5.4 <i>Van</i> A2 and B2 Genes	42
3.2.6.5.5 Detection of PCR Products	44
3.2.6.5.6 Visualization of RCR Product	44
3.2.6.5.7 Interpretation the Results of PCR	45
3.2.7 Statistical Analysis	45
CHAPTER FOUR: RESULTS	46
4.1 Epidemiological Findings	46
4.1.1 Samples Distribution	46
4.1.2. Demographic Data	47
4.1.2.1 Gender	47
4.1.2.2 Age Groups	49
4.1.2.2 Other Demographic Data	49
4.1.2.3 Type of Skin and Soft Tissue Infections (SSIs)	50
4.2 Phenotypic Characteristics	50

4.2.1 Gram's Colony Morphology and Biochemical Characters	50
4.2.1 Bacteriological Findings	51
4.2.3 Drug Susceptibility Testing (DST)	51
4.2.3.1 Surgical Prophylaxis	53
4.3 Polymerase Chain Reaction (PCR)	53
4.3.1 Extraction of DNA	54
4.3.2 Detection of <i>arcC</i> gene	54
4.3.3 Detection of <i>mecA</i> gene	54
4.3.4 Detection of Van genes	55
4.3.5 Molecular Method versus Conventional Method	56
CHAPTER FIVE: DISCUSSION, CONCLUSION AND RECOMMENDATIONS	57
5.1 Discussion	57
5.2 Conclusion	62
5.3 Recommendations	63
REFERENCES	64
Appendix I (Colored Plates)	70
Appendix II (Preparation of Media)	73

Appendix III (Reagents and Stains)	76
Appendix IV (PCR Reagents)	80
Appendix V (The questionnaire)	82

List of Tables

Tables	No. of page
Table 1. Distribution of the patients among the medical centers	31
Table 2. Interpretation of inhibition zone diameter	37
Table 3. Sequences of the used primers in the present study	43
Table 4. Preparation of reaction mixture of arcC, Van A and B gene	43
Table 5. PCR amplification programs of (arcC, mecA, VanA &B1 and Van A&B 2)	44
Table 6. Distribution of samples according to different Hospitals and medical centers	47
Table 7. Distribution of MRSA among study subjects in different Hospitals, Sections and medical centers according to the gender	48
Table 8. Other demographic data restricted to the study subjects	49
Table 9. Frequencies of MRSA versus MSSA among isolated Staphylococcus aureus	51
Table 10. Results of DST for MRSA Isolates against Different Antibiotics	52
Table 11. Results of DST of MRSA against Second and Third Generation of Cephalosporins	53

Table	12.	Result	Staph.	aureus	and	MRSA	by	conventional	and
molec	ular	method							

56

List of Figures

Figures	No. of
	page
Figure 1. Summary of virulence factors of Staphylococcus	15
aureus	
Figure 2: a plasmid-like element called the Staphylococcus	27
cassette chromosome carry <i>mecA</i> .	
Figure 3. Distribution of samples according to gender	48
Figure 4. Distributions of enrolled patients according to Age	49
group	
Figure 5. Distributions of SSIs among the enrolled patients	50
Figure 6. Pure DNA obtained by QIAGEN DNA Mini Kit as	54
detected by agarose gel electrophoresis	
Figure 7. The ampilicon of <i>arcC</i> gene on 1.5% agarose gel:	54
lane 1 and 10: DNA marker (100 bp); lane 2: negative control;	
lane 3: positive control (Staph. aureus from the reference	
bacteria at research Lab.), lanes (4 to 9) positive arcC gene of	
MRSA (456 bp)	
Figure 8. The result of mecA gene (310 bp) by PCR: lanes (1	55
to 6), lane 0: DNA marker (100 bp), lane 3: positive control	
(MRSA from the reference bacteria at research Lab.), lane 6:	
negative control (methicillin susceptible Staph. aureus) and	

lanes (1, 2, 4, and 5) positive mecA gene of MRSA	
Figure 9. The result of VanA gene (732 bp) by PCR: lanes (1	55
to 6), lane 0: DNA marker (100 bp), lane 3: positive control	
(VRSA from the reference bacteria at research Lab.), lane 6:	
negative control (vancomycin susceptible Staph. aureus) and	
lanes (1, 2, 4, and 5) Negative VanA gene	

List of Abbreviations

Carbamate kinase
Bio Medical Center
Base pairs
Community-acquired methicillin resistant
Staphylococcus aureus
The Centers for Disease Control and Prevention
Coagulase-negative staphylococci
Diabetes Mellitus
Deoxyribonucleic acid
Deoxyribonuclease
Deoxynucleotide triphosphate
Double-stranded
Drug Susceptibility Testing
Ethylene diaminotetracetic acid
Exfoliative toxins
fatty acid modifying enzyme
gram
Hospital acquired methicillin resistant <i>Staphylococcus</i>

	aureus
ICU	Intensive care unit
IgG	Immunoglobulin G
IV	Intravascular
Mb	Mega base pair
McF	McFarland
MIC	Minimum inhibitory concentration
min.	minute
Ml	milliliter
Mm	millimeter
MRSA	Methicillin resistant Staphylococcus aureus
MSA	Mannitol salt agar
MSSA	Methicillin susceptible Staphylococcus aureus
PBP2	Penicillin binding protein 2
PCR	Polymerase chain reaction
pH	Power of Hydrogen
PRSA	Penicillin-resistant Staphylococcus aureus
PTSgs	Pyrogenic toxin super antigens

Psi	Pounds per square inch
PVL	Panton Valentine Leukocidin
Staph. aureus	Staphylococcus aureus
SCCmec	Staphylococcal Cassette Chromosome mec
spp.	Species
Staph	Staphylococcus
SSSS	staphylococcal scalded-skin syndrome
SSTI	Skin and soft tissue infection
TSS	Toxic shock syndrome
U	unit
USA	United States of America
UTI	Urinary tract infection
UV	Ultraviolet
VISA	Vancomycin intermediate <i>Staphylococcus aureus</i>
VRE	Vancomycin-resistant enterococci
VRSA	Vancomycin-resistant Staph. aureus
v/v	volume per volume