

CHAPTER ONE

INTRODUCTION

The practice of feeding live stock with sub therapeutic levels of antibiotics has been in use for over fifty years antibiotic usage is possibly the most important factor that promotes the emergence, selection and dissemination of antibiotic resistant microorganisms in both veterinary and human medicine (Castanon, 2007) at slaughtering, resistant strains from the gut may contaminate poultry carcasses and as a result poultry meats are often associated with multi resistant microorganisms the use of antibiotic growth promoters has been banned in many countries, especially in European union (Castanon, 2007) as a result new commercial additives of plant origin, considered to be natural products that consumers would accept, have been proposed to livestock producers Herbs spices and various plant extracts have been received and increased attention as possible antibiotic growth promoter replacement in this view, aromatic plants and essential oils extracted from these plants become interesting due to their antibacterial (Soliman *et al.*, 2002) antioxidant (Dragland *et al.*, 2003); Botsoglou *et al.* 2004) antibacter (Dorman and Deans, 2002) antifungal (Jantan *et al.*, 2003) activities and as hypo cholesterolemic (Craig, 1999) and stimulate effect of an animal digestive enzymes and improve utilization of digestive products through enhanced liver functions (Hernández *et al.*, 2004); lee *et al.*, 2003); (Cifitic *et al.*, 2005).

Essential oils derived mainly from spices and herbs and their purified compounds have been shown to have antimicrobial actions in vitro Cowan, (1999); Ultee *et al.*, (2002); Faleiro *et al.*, (2003). Examples of such natural antimicrobial compounds are carvacrol, thymol, limonene and cineole that are present in the essential oil fractions of oregano, laurel, sage and myrtle (Riebau *et al.*, 1997); Ultee *et al.*, (2002).

Vitamins are defined as group of complex organic compounds present in small amounts in natural food stuffs that the diet causes deficiency disease. Because of the possibility of specific biosynthesis pathways in some species, some vitamins such as carnitine, could be considered only as essential metabolites in these species and dietary sources are not needed.

Super San Soluble is a supplement of amino acid and vitamins for ruminants, pig, rabbits and birds. Vitamins to increase growth. development and production and reduce frequency and severity of many infectious diseases. The protein hydrolysate supplements the diet in certain conditions of a very high protein intake, a requirement which be accomplished with ordinary food.

The objective of this study was to evaluate the effect of super san soluble on the performance of broiler chicks.

CHAPTER TWO

LITERATURE REVIEW

2.1 Feed Additives

The term “additive” is applied by the feed compounder, in a broad sense, to all products other than those commonly called feed stuffs that may be added to the ration with the object of obtaining some special effects. Feed is a major component, affecting net return from the poultry business, because 80% of the expenditure in terms of cash is spent on feed purchase (Javed *et al.*, 2009).

Feed additives, non-nutritive, are sometimes included in the feed mixture in very small quantities and with careful weighing, handling and mixing, to insure that dietary nutrients are ingested, digested, protected from destruction, absorbed and transported to the cells of the body. Other feed additives have been used to alter the metabolism of the chicken in an effort to produce better growth or more desirable finished products (Leeson and Summers, 2001).

The most common types of feed additives used are: Antibiotics and arsenicals which are used to help protect feeds from microbial destruction to prevent production of toxic products by the intestinal microflora, Anticoccidials which are used in broiler feeds and in the diet of rearing replacement pullets, Antifungal to prevent the growth of harmful molds and fungi in feed or in the digestive tract of the chicken; Antioxidant, flavoring agents, pellet binder and carotenoids (Parks *et al.*, 2000) and (Sreenivasaiah, 2006), feed additives like: probiotics, prebiotics, essential oils, enzymes and vitamins.

2.2 Antibiotics

However, the use of antibiotics as feed additives is risky due to, not only cross-resistance, but also to multiple resistance in pathogens (Bach

Knudsen, (2001) and Schwarz *et al.*, (2001). Therefore, antibiotics have been discredited by consumer associations as well as by scientists, e.g. the use of most antibiotic growth promoters has been banned by the European Union (EU). Consequently, the animal feed industry is under increasing consumer pressure to reduce the use of antibiotics as a feed additive and find substitutes for antibiotics in the diet Hertrampf, (2001); Humphrey *et al.*, (2002). Many scientists have searched for alternatives to antibiotics Langhout, (2000); Mellor, (2000); Wenk, (2000) and Kamel, (2001).

The growth promoter effect of antibiotics was discovered in the 1940s, when it was observed that animals fed dried mycelia of *Streptomyces aureofaciens* containing chlortetracycline residues improved their growth. The mechanism of action of antibiotics as growth promoters is related to interactions with intestinal microbial population (Dibner and Richards, 2005; Niewold, 2007). It is commonly known that the sub-therapeutic use of antibiotic growth promoters (AGP) in poultry production may result in the development of antibiotic-resistant pathogenic bacteria, which may be hazardous to human health. In search of effective alternatives to AGP, a special attention is given to their effect on gut microbial community which contributes to the intestine function. Until now, the interest has been focused mainly on fermentable functional feed ingredients, like fructans, or mannanoligosaccharides that exhibit beneficial effect on gut microflora, integrity of intestinal mucosa, enzymes activity and performance parameters in broiler chickens (Kim *et al.*, 2011); (Bogusławska-Tryk *et al.*, 2012);(Nabizadeh, 2012). An insoluble, non-fermentable fiber fraction, including cellulose and lignin, is conventionally considered as a diet diluent which can influence energy balance of broilers (Svihus and Hetland, 2001; Krás *et al.*, 2013), whereas little attention is given to the effect of cellulose or lignin on the gastrointestinal microflora population. However, studies show that cellulose, as an effective feed ingredient, may influence the number of

gut bacteria, especially beneficial *Bifidobacterium* and *Lactobacillus* as well as potential pathogens and its effect depends on the level of cellulose supplementation and bird age (Cao *et al.*, 2003; Shakouri *et al.*, 2006; Saki *et al.*, 2010). It is generally accepted that phenolic fragments of purified lignin exhibit the antimicrobial properties (Baurhoo *et al.*, 2008).

2.3 Prebiotics

Prebiotics defined as non-digestible food ingredient (Gibson and Roberfroid, 1995). The use of prebiotics in broiler's diets does not have a long history . Several authors have observed the positive effects of prebiotics fractions included in the broiler's diet (Rebole *et al.*, 2010) reported diet supplemented with inuline had higher body weight gain and increased growth performance, dressing percentage, breast and thigh muscle weight (Park and Park, 2011), prebiotics improved digestion in clouding enhancing mineral absorption (Coxam VCNOV, 2007).

Gibson and Roberfroid (1995) defined prebiotics as the food ingredients that provide beneficial effect to the host by selectively stimulating the growth and/or metabolism of a limited group of bacteria in the intestinal tract, acting closely to probiotics because it would constitute the “food” of probiotic bacteria and also blocking adherence sites, immobilizing and reducing the fixation capacity of pathogenic bacteria in the intestinal mucous (Silva, 2000; Andreatti Filho and Silva, 2005). This association favors the intestinal microbiota by the action of prebiotics that are able to link themselves to the fimbriae of pathogenic bacteria, conducting them along the fecal bolus, stimulating the growth and accelerating the metabolism of a limited number of non-pathogenic microorganisms. The action of prebiotics is added to this mechanism, making easy the nutrition of cells (enterocytes) that recover the digestive tract and provide balance and intestinal health to birds (Gibson and Roberfroid, 1995).

2.4 Probiotics

Probiotics are defined as feed additives that contain live microorganism and promote beneficial microbiota Fuller, (1989); Huang *et al.*, (2004), probiotics improve immunity and live weight gain and feed conversion rate of broiler Jin *et al.*, (2000); Zulkifli *et al.*, (2000) and Huang *et al.*, (2004) and improve broiler growth performance and prevent poultry pathogens and diseases Tortucro, (1978); Owings *et al.* (1990); Jin *et al.* (1997); Zulkifli *et al.* (2000); Kalavathy *et al.*; (2003); Kabir *et al.* (2004); Gil De Los Santos, *et al.* (2005); Timmerman *et al.*(2005); Mountzouris *et al.* (2007) and Awad *et al.* (2009).

Probiotic, based on Fuller (1989) definition, “are live microbial feed supplement that beneficially affects the host animal by improving its intestinal microbial balance”. Probiotic efficiency depends on several factors, such as microbial species composition (e.g., single or multi strain) and viability, application procedure, dosing level, frequency of application, age, type of diet, sanitation and environmental stressors factors. However, beneficial effects of probiotic on broilers including: performance (Mountzouris *et al.*, 2007; Kralik *et al.*, 2004), modification of intestinal microflora (Teo and Tan, 2007; Mountzouris *et al.*, 2009), nutrient digestibility (Apata, 2008) and immunomodulation and gut mucosal immunity (Farnell *et al.*, 2006; Teo and Tan, 2007) have been reported. These positive effects by application of probiotics could be related to increase population of beneficial microflora and removal of pathogenic bacteria by means of competitive exclusion and antagonism (Fuller, 1989); adapting bacterial metabolism (Jin *et al.*, 1997); improving feed intake digestion and absorption (Nahanshon *et al.*, 1993) and stimulating the immune system (Havenaar and Spanhaak, 1994). The enhancement of the immune system may be in relation to increase production of antibodies production of antibodies particularly immunoglobulin G (IgG) and

immunoglobulin A (IgA) classes and also increase local antibodies at mucosal surface such as gut wall (usually IgA) (Koenen *et al.* , 2004).

2.5 Essential Oils

Definition of essential oils and general introduction: An essential oil is a mixture of fragrant, volatile compounds, named after the aromatic characteristics of plant materials from which they can be isolated (Oyen and Dung, 1999). The term 'essential' was adapted from the theory of 'propose' by Paracelsus who believed that this quintessence was the effective element in a medical preparation (Oyen and Dung, 1999). Because the term 'essential oil' is a poorly defined concept from medieval pharmacy, the term 'volatile oil' has been proposed (Hay and Waterman 1993). However, the name of 'essential oil' will be used preferentially in this review.

Essential oils are very complex mixtures of compounds and their chemical compositions and concentrations of individual compounds are variable. For example, the concentrations of two predominant components of thyme essential oils, I.e. thymol and carvacrol have been reported to range from as low as 3% to as high as 60% of total essential oils (Lawrence and Reynolds, 1984). Cinnamaldehyde, a main principle of cinnamon essential oil, amounts to approximately 60 to 75% of the total oil (Duke, 1986). Because of the large variation in composition, the biological effects (Schilcher, 1985); Janssen *et al.*, 1987; Deans and Waterman, 1993). If any, of essential oils may differ. This diversity of essential oils us to select four pure principles, i.e. thymol cinnamaldehyde, beta-ionone and carvacrol, for evaluating their possible role as alternatives to antibiotics in poultry production. The chemical properties and biological activities of the four compounds are summarized.

Essential oils are volatile compounds, extracted from plants by steam distillation, which have a variety of beneficial properties, such as

flavoring, stimulation of enzyme secretion, and antioxidant or antimicrobial activities. Related to the digestibility of nutrients, thymol, cinnamaldehyde, or a commercial preparation of essential oils (EO) components (CRINA Poultry, DSM Nutritional Products Ltd., Basel, Switzerland) were demonstrated to increase amylase activity in the intestinal digesta of female broilers (Lee *et al.*, 2003). With a plant extract, consisting of capsaicin, cinnamaldehyde, and carvacrol, Jamroz *et al.* (2005) observed an increase in the lipase activity in pancreas and in the intestinal wall, which improved feed conversion in broilers. Dietary piperine, which is the pungent principle of black pepper, was found to favorably stimulate the digestive enzymes of pancreas and thus to enhance the digestive capacity and to significantly reduce the gastrointestinal feed transit time (Srinivasan, 2007). Likewise, plant extracts from a mixture of oregano, cinnamon, and pepper or from sage, thyme, and rosemary improved apparent whole-tract and ideal digestibility of the nutrients in broilers (Hernández *et al.*, 2004).

For commercial poultry production, the antimicrobial effects of EO are also of particular interest. Respective in vitro broad spectrum activities have been reported for oregano and monolaurin EO (Preuss *et al.*, 2005), for rosemary extracts (Santoyo *et al.*, 2005), or for EO from chrysanthemum (Shunying *et al.*, 2005). Furthermore, oregano and thyme oils were found to be active against poultry and pig derived strains of *Escherichia coli* and *Salmonella* (Peñalver *et al.*, 2005). Specific blends of EO components were demonstrated to control colonization and proliferation of *Clostridium perfringens* in the gut of broilers, which is expected to protect against necrotic enteritis (Mitsch *et al.*, 2004).

A mixture of capsaicin, cinnamaldehyde, and carvacrol reduced intestinal *E. coli*, *C. perfringens*, and fungi and increased beneficial *Lactobacillus spp.* in broilers (Jamroz *et al.*, 2005). The cumulative effects of

EO on digestibility of nutrients and on modulation of the gut microflora eventually result in an improvement of broiler performance. Essential oils from different herbs in Turkey were found to improve weight gain, feed conversion and carcass yield of broilers Alcicek *et al.*, (2003). A plant extract, consisting of 3 EO, improved feed conversion and enhanced breast muscle yield in broilers (Jamroz *et al.*, 2005). When testing dietary inclusion of various culinary herbs or their EO in female broiler chicks, generally dietary thyme oil or yarrow herb inclusion had the most positive effects on chick performance, whereas oregano herb and yarrow oil were the poorest supplements (Cross *et al.*, 2007). Besides EO, organic acids are widely used in monogastric animals to improve performance via a modulation of the gut microflora. Blends of various organic acids induced a shift in the intestinal microbiota toward more homogenous and distinct populations and increased *Lactobacillus* colonization of the chick ileum (Nava *et al.*, 2009). Furthermore, citric acid was demonstrated to achieve similar performance results in broilers as the antibiotic growth promoter avilamycin (Chowdhury *et al.*, 2009). Recently, benzoic acid has been identified as an efficient feed additive to improve growth performance, nutrient digestibility, and nitrogen balance as well as to reduce gram-negative bacteria in the gastrointestinal tract of piglets (Kluge *et al.*, 2006). In poultry, benzoic acid was found to have rather negative effects, given that a significant growth depression and deterioration of feed conversion were observed, when supplementing this compound to broilers (Jozefiak *et al.*, 2007). Coliform bacteria in the ceca, however, were decreased, indicating that the considerable antimicrobial activity of benzoic acid could beneficially influence gut health in poultry as well. Because an additive effect between organic acids and EO was suggested due to the fact that digestive enzymes work more efficiently under acidic conditions.

2-6 The Enzymes

Supplementation of mixtures broiler with enzymes is applied in order to the efficiency of the production of poultry meat. This is especially interesting if enzymes which enable utilization of feeds of poorer nutritive value are used. Numerous authors have established that by application of enzymes in the production performances can be improved by even 10% (Cowieson *et al.*, 2000), Cmiljanic *et al.*, 2001). The preparation should provide all requirement nutrients by the broiler chicks. Most of the main feed ingredients used in broiler diets such as cereal grain and oil seed cakes are deficient in essential amino acids quantity and quality wise. The imported supper-concentrate was provided to suffice for essential amino acids and other micro-nutrients which consequently increase the total cost of poultry feed. Researcher now seeking to find means to improve the utilization of the essential amino acids and other nutrients of the plant based diet in order to achieve high production with least cost to maintain and sustain production (Mukhtar *et al.*, 2010a; Mukhtar *et al.*, 2010b). However, use of more than 20% RB in broiler diets reduces the performance and mineral status of the birds while increasing the mineral excretion (Piyaratne *et al.* 2011). Presence of several anti-nutrients such as phytate (Puminn, 2003); fibre (Gallinger *et al.* 2004); lipases (Sharif, 2009) and anti-proteolytic sub-stances (Mujahid *et al.* 2005); Ersin Samli *et al.* 2006) have reported as the reasons for poor in vivo nutritive value of RB. Selle *et al.* (2007) reported that, rice bran contains 17.8g kg⁻¹ of total phosphorus (P) and 14.2g kg⁻¹ phytate-P. Typically, poultry diets contain from 2.5 to 4.0g kg⁻¹ phytate-P (Ravindran, 1995). Phytate-bound P is poorly utilized by monogastric animals, due to either insufficient quantity or a lack of intestinal phytase secretion (Rafacz-livingstan *et al.* 2005); Mohammed *et al.* 2010). Phytic acid acts as a strong chelator, forming protein-mineral-phytic acid complexes and reduced protein and mineral bioavailability (Akande *et al.* 2010). On the other hand, excretion of undigested P creates environmental problems like eutrophication

of water bodies (Selle *et al.* 2007). Consequently increased utilization efficiency of phytate bound P by poultry benefits both industry and environment. Even though the microbial phytase found to be effective in improving phytate degradation, supplementation of diets with phytase increased the litter moisture content (Puminn, 2003). Heat liability of phytase and cost are among the drawbacks of the use of phytase. Therefore alternative methods for increasing phytate degradation need to be developed. Several authors have reported positive effects of citric acid (CA) on phytate degradation (Boling *et al.* 2000; Liem *et al.* 2008; Connelly, 2011), digestibility of protein (Atapattu and Nelligaswatta, 2005); Ao *et al.* 2009) and growth performances (Chowdhury *et al.* 2009; Islam *et al.* 2012) in poultry. Atapattu and Nelligaswatta (2005) showed that CA have positive effects on growth performance and feed intake only when diets are low in available phosphorus (aP). Several other authors (Boling *et al.* 2000); Boling-Frankenbach *et al.*, 2001); (Snow *et al.* 2004) have reported that improved phytate-P utilization efficiency in broilers when the diets were deficient in non-phytate P (NPP).

2.7 The Vitamins

Vitamin and minerals are vital nutrients that are involved in both metabolic and physiological processes which are critical for human and animal health and animal feed production. Vitamins are defined as group of complex organic compounds present in small amounts in natural foodstuffs that are essential to normal metabolism and lack of vitamins in the diet causes deficiency disease: Because of the possibility of specific biosynthesis pathways in some species, some “vitamins”, such as carnitine, could be considered only as essential metabolites in these species and dietary sources are not needed. Vitamins are an essential component of a well-balanced diet and their major function is the metabolism and utilization of nutrients. Through research into the biological mechanisms of vitamin action, it has

now been established that substantially higher intake of some vitamins may significantly influence the immune process in chickens (Siddique, 2004). In poultry, biotin is an essential coenzyme in carbohydrate, fat and protein metabolism.

Poultry sub-sector is an important avenue to reduce the gap between demand and supply of animal protein. Animal protein has higher biological value in comparison with plant protein. The broiler industry demands a fast growing chick and good quality feed with high level of energy, protein, vitamin and essential minerals to support maximum growth within a short period of time. For the sustainability of broiler industry, the production cost should be as minimum as possible. On the other hand, deficiency diseases may cause mortality, reduce body growth and disease resistance etc.(Singsen, 1947). Chickens are more susceptible to vitamin deficiency because gut flora can synthesize very little vitamins but complete absence of dietary vitamins in intensively kept chickens undergoes many stresses (Ward, 1996). Vitamin A, riboflavin and B12 are usually low in poultry diets. Inclusion of multivitamin and amino acids in the formulated diet has become indispensable practice because blending of feed ingredients do may not ensure all essential vitamins at the right amounts needed for chicken. Some important vitamins like vitamin A, D3, E, folic acid, pantothenic acid, pyridoxine and riboflavin etc should be checked carefully in the diet. As the ultimate goal of broiler is to produce more meat within a short period of time, additional multivitamin and enzyme supplement with commercial feed may increase meat production more rapidly. Enzyme and multivitamin protect deficiency diseases and stimulate growth rate. Beside this, amino acids supplement along with multivitamin reduces mortality, keep birds healthy, increase feed intake, improve digestion and feed conversion rate. products contain non-starch polysaccharides (NPS) such as cellulose, xylose, arabinose, galactonic acid which are not easily digested by poultry. Most of

the feed ingredients contain some anti-nutritional factors and indigestible part, which hinders feed utilization and bird's performance.

High ambient temperature reduces feed intake, live weight gain, feed efficiency (Donkoh, 1989), and thus negatively influences the performance of broilers. Hurwitz *et al.* (1980) suggested that decrease in growth rate was due partly to the decrease in feed intake. Animals stressed under improper environmental conditions or subjected to an artificial stress via adrenocorticotropin hormone (ACTH) and epinephrine injections are found to have reduced α -tocopherol, retinol and ascorbic acid concentrations in plasma and blood cells (McDowell, 1989), whereas lipid peroxidation levels were found to be high in plasma and tissues due to increased production of free radicals (Naziroğlu *et al.*, 2000). Moreover, heat stress impairs absorption of vitamins A, E and C, and thus, increases the requirement of these vitamins (Naziroğlu *et al.*, 2000); (Sahin *et al.*, (1999); (Klasing, 1998). On the other hand, stress causes reduction in plasma and tissue concentrations of minerals such as Fe, Zn and Cu which are related to immune system (Beisel, 1982).

The increase in environmental temperature in summer has been the major problem to poultry producer in Sudan, which puts an additional effort on body heat regulatory mechanism, and reduces feed consumption which affects the productive performance of chickens. Vitamin and minerals are vital nutrients that are involved in both metabolic and physiological processes which are critical for human and animal health and animal feed production. Puthongsiriporn *et al.*,(2001) reported that under environmental stress, feed intake, egg production decreased in birds fed with vitamin E and C. Ziaei *et al.*, (2013) found that the inclusion of vitamin E and selenium had not a significant effect on feed intake, they decreased it. Mohiti Asli *et al.*,(2007) found no significant effect on egg weigh and feed conversion ratio due to inclusion of vitamin E. However, Ziaei *et al.*, (2013) studied the effects of

different vitamin E and organic selenium levels on performance and immune response of laying hens, they recorded significant effects on production performance and immune improvement. Increasing the level of vitamin D₃ in the chicks diet increased body weight gain linearly (Edwards *et al.*, 2002; Kasim and Edwards, 2000). Broiler breeder diet influences subsequent egg production Peebles *et al.*, (2000a), embryogenesis and hatchability of broiler eggs Peebles *et al.*, (2000b). Chicks, embryo tissues contain a high proportion of polyunsaturated fatty acids Diplock, (1994). Also, tissues of newly hatched chicks express natural antioxidants (Vit E, glutathione GSH, Carotinoids and ascorbic acid) and antioxidant enzymes cofactors (Se, Zn, Mn, and Fe) Surai, (1999). The level of natural antioxidant in tissues declines progressively after hatching, therefore enhancement of Glutathione peroxidase (GSH-Px) activity as a result of selenium (Se) supplementation is an effective mean of increasing chicks viability post hatch Surai, (2000). Vit E is The main antioxidant of biological membranes Niki, (1996) and due to its location inside the membrane at the water lipid interface, Vit E is able to scavenge free radicals effectively. Trace minerals are essential in poultry diets as they participate in the biochemical processes required for normal growth and development. Selenium is an integral component of glutathione peroxidase (GSH-Px.), which together with Vit E form a part of cellular defense against reactive oxygen species produced during stress Surai, *et al.*, (2001). Vitamin E and selenium appear to participate in the same biochemical relationships and food. Selenium is essential for the proper function of the antioxidant enzyme glutathione peroxidase, which protects the cell by destroying free radicals Rotruck, *et al.*, (1973). Selenium and vitamin E as antioxidants interact with each other to form an efficient antioxidant defense mechanism and when both are supplemented to birds, they play an important role in maintaining bird health, productivity and reproductive characteristics Surai, (2000).

CHAPTER THREE

MATERIALS AND METHODS

3.1 Experimental Site:

The experiment was carried out at the Department of Animal Production, College of Agricultural Studies, Sudan University of Science and Technology (SUST), from 27th September 2014 and ending on 1st November 2014 in an ambient temperature ranging between (27 - 41) °C (Appendix).

3.2 Experimental birds:

A total of sixty three, seven days old unsexed broiler chicks strain Aber acre, purchased from local commercial hatchery (Mico). The chicks were selected from the poultry farm after one week of adaptation period. Moreover, The chicks were randomly divided into three experimental groups as follows: A, B and C. each treatment group was subdivided into three replicates of 7 birds per each. The birds in three groups were reared under the optimum environmental conditions and feeds were provided ad-labitum Feed provided freely, chicks were vaccinated against Marek's disease on hatchery. On farm they were vaccinated against Gumboro D78 disease (IBD) at 7, 13, 21 and 28 days of age and New Castle disease in drinking water at 7, 21 and 28 days (Colon 30). On the other hand, a soluble multi vitamin compounds to guard against stresses. Based on a local vaccination program, chicks in all groups were vaccinated against Gumboro D78 disease (IBD) at 10 days old. During the experiment, birds were weighed weekly and feed intake per pen was recorded at the same time. The measured performance parameters includes: final body weight (g), body weight gain (g), feed intake (g), feed conversion ratio, and mortality rate (%).

3.3 Housing:

An open system poultry house was used. The house was constructed on concrete floor with local materials. The roof was made of metal sheets. The sides were permanently covered. The pens, 1×1 m each, inside the house were prepared using wire mesh partitioning. The pens were cleaned commencement of the experiment. A layer of wood shavings (about 3 cm thick) was laid on the pen floor as a litter material.

Each pen was supplied with 2.5 gallon drinker and 5 kg feeder which were cleaned and disinfected before starting the feeding trial. The feeder's height was adjusted according to progressive growth of the chicks. Over head light were provided 24 hours, in a form of natural light during the day and artificial light during the night 1-2 bulb lamps (100 watt) to each pen.

3.4 Experimental diet:

The chicks were fed a commercial broiler pre-starter (Galdus Koudijs, 100 grams per chick during the first week) for week. Then a basal diet fed to the chicks in all groups which was formulated to meet the NRC, (1994) recommendations for broiler chickens. Super San Soluble, added in the water which is composite of a group of vitamins. Group A as control (tap water), group B and C drinking water was supplemented with Super San Soluble (day interval)1 ml/10L and 2 ml/10L for each pen respectively.

Table (1): Composition of the experimental control diet used

Ingredients	Dura	Ground nut Cake	Sesame cake	Meth.	Lysine	Oyster shell	Salt	Conc.*	Dical Phos.
%	64.142	14	15	0.159	0.344	0.487	0.25	5	0.618

*Conc.: Broiler concentrates: Crude protein 22.82%, Crude fiber 4.13%, lysine 1.497%, Methionine 0.63%, Calcium 1.15%, Phosphorus 0.76%, and ME 3105.212.

Table (2): Calculated analysis of experimental diet

ME	CP%	Lysine%	Methionine	Ca%	Phos.%	Fibre%
3105.21 Lodhi <i>et al.</i> , (1976)	22.82	1.5	0.63	1.15	0.76	-

Table (3): Chemical analysis of experimental diet

DM	Moisture	CP	Ash	EE	NNF
6.39	93.67	21.9	10.6	3.4	57.71

ME= metabolizable energy, CP= crude protein, EE= ether extract, Ca= calcium
Phos.= phosphorus, NNF= nitrogen free extract, DM= dry matter

Table (4): Chemical composition of the experimental diet

Broiler starter		3100	22	1.2	0.5	1	0.8	3.5
	%	ME	CP	Lysine	Meth.	Ca	P	Fiber
Dura	64.142	2205.202	8.485987	0.141112	0.07697	0.032071	0.19884	1.539408
Lysine	0.344	0	0	0.33712	0	0	0	0
G.N. Cake	14	383.46	6.1012	0.224	0.063	0.0868	0.0784	1.3608
Sesame Cake	15	416.55	6.2355	0.195	0.18	0.3015	0.1395	1.227
Meth.	0.159	0	0	0	0.15741	0	0	0
Oster Shell	0.487	0	0	0	0	0.179898	0.000292	0
Salt	0.25	0	0	0	0	0	0	0
Dical. Phos.	0.618	0	0	0	0	0.14832	0.11124	0
Conc.	5	100	2	0.6	0.15	0.4	0.23	0
	100	3105.212	22.82269	1.497232	0.62738	1.148589	0.758272	4.127208

ME= metabolizable energy, CP= crude protein, Meth.= methionine, Ca= calcium
Dical. Phos.= Di Calcium phosphorus, Conc.= concentration, G. N. Cake = Ground nut
cake

3.4.1 Supersan Soluble:

It is a supplement of amino acids and vitamins for ruminants, pigs, rabbits and birds. Vitamins to increase growth, development and production, and reduce frequency and severity of many.

3.4.2 Composition:

Vitamin A 600.000 I.U; vitamin D3 50.000 I.U.; Vitamin E 100 mg; Vitamin B1 100 mg; vitamin B2 100 mg; vitamin B6 200 mg; vitaminB12 1,5 mg; Vitamin K 200 mg; Calcium pantothenate 500 mg; Nicotinic acid 1 g; Glutamic acid 1.4 g; Aspartic acid 960 mg; Arginine 270 mg; Cystine 760; Lysine 500 mg; Histidine 220 mg; Glycine 810 mg; Serine 480 mg; Alanine 700 mg; Tyrosine 340 mg; Threonine 320 mg; phenylalanine 340 mg; Valine 270 mg; Methionine 250 mg; Leucine 760 mg; Isoleucine 250 mg; Tryptophan 100mg; Excipient to 100mL.

3.4.3 Indication:

Cases of stress, convalescence, loss of appetite, decreased organic defenses against infection, illness, after vaccination and transport, male sexual frigidity, artificial feeding in suckling animals and hepatic failure . As a growth stimulant for underdeveloped animals.

3.4.4 Dosage and administration route for Birds and rabbits:

Oral route, 0.07 – 0.13 ml of Supersan soluble per each litre of drinking water.

3.5 Parameters:

Birds of each replicate were group weighed at weekly intervals and feed intake was recorded at the time of weighing. Feed Conversion Ratio (FCR) and body weight gain were calculated weekly and mortality was recorded daily.

3.6 Carcass preparation:

At end of the experiment one bird was selected from each group and was weighed individually. Then they were slaughtered. After bleeding, the

slaughtered chicks were scaled in hot water, feather plucked manually then washed. The head was removed close to the skull. Feet and shanks were removed at the hock joint, and eviscerated, gizzard, abdominal fat, heart, gizzard and liver were expressed as a percentage of live weight.

3.7 Calculation:

The cold and hot carcass weights were expressed as a percentage of live weight. The commercial cuts were expressed as a percentage of cold carcasses. Non carcass components such as head, heart, gizzard and liver were expressed as a percentage of live weight.

3.8 Statistical Analysis:

The data obtained were statistically analyzed with the standard procedures of analysis of variance (ANOVA) using SPSS Inc. (1999) program.

CHAPTER FOUR

RESULTS AND DISCUSSION

4.1 Results:

The effects of Supersan administration via in drinking water on growth and carcass performance of broiler chicks were summarized in Table 2 and 3, respectively. Results obtained showed no significant ($P>0.05$) effects of treated drinking water on feed intake, feed conversion ratio and mortality rate of chicks compared to control group. Although there was a numerical increase in feed consumption for chicks group supplemented with 2 mg/10 L drinking water compared to other tested groups. While chicks supplemented with 1 mg/10 L drinking water showed significantly ($P>0.05$) low live body weight, feed intake, body weight gain and feed conversion ratio.

Results also showed no significant ($P>0.05$) difference for non carcass components due to Supersan administration compared to control group.

Table(5): effect of Supersan administration in drinking water for broiler chicks performance

Treatment	Feed intake (g)	Body weight (g)	Weight gain (g)	FCR
Control	2294.00	1898.00	1529.60	1.50
B	2144.10	1440.00	1218.60	1.76
C	2313.90	1670.40	1379.10	1.70
SE	154.70	133.39	94.59	0.178

Table(6): effect of Supersan administration in drinking water on non carcass components of the experimental broiler chicks

Treatment	Hot weight	Abdominal fat	Liver	Heart	Gizzard	Head
Control	1658.33	0.17	0.64	0.22	0.5	0.57
B	1232.33	0.20	0.49	0.17	0.17	0.54
C	1556.33	0.25	0.53	0.18	0.18	0.60

4.2 Discussion

The experiment was carried out in a semi-closed house, cleaned and disinfected. Feed intake is more or less similar; the highest feed intake was recorded in treated group C (2313.9 g) and the lowest value in treated group B. Similar results were recorded for feed conversion ratio. This may be due to that the environmental condition of the experiment could be considered as responsible factor for inconsistency in results, so the birds did not consumed more water, although the benefit of treatment is not enough, or the dose recommended by the manufacture company did not enough under our condition, and from this view the chicks performance did not affected by treatment. However, chicks on group C (2 gm/10 L) performed better than those of group B (1 gm/ 10 L). This result were similar to the findings of Puron *et al.*, (1994) who examined 200 ppm dietary vitamin C supplementation and found no effect on performance and survivability when the average environmental temperature was 26 °C, Mohiti Asli *et al.*, (2007) who found no significant effect on egg weigh and FCR due the inclusion of vitamin E, also Puthongsiriporn *et al.*, (2001) recorded decreased feed intake for birds fed with vitamin E and C under environmental stress, Preston *et al.*, (2000) who found that enzyme inclusion did not improve performance; Richter *et al.*, (1994) who reported that live weight did not improved by enzyme supplementation and Ziaei *et al.*, (2013) when fed laying hens with vitamin E and organic selenium. On the other hand, the results of this study contradict with Edwards *et al.*,(2002); Kasim and Edwards (2000), who found increased body weight gain for chicks fed increasing levels of vitamin D₃;Christmas *et al.*, (1995); Meng *et al.*, (2005); Saleh *et al.*, (2005) and Wang *et al.*, (2005) who reported improved feed utilization by exogenous enzyme, also Paul *et al.*, (2010) who observed that multivitamins and enzyme supplementation significantly ($P < 0.05$) increased body weight of carcass traits. Supplementation also did not influence birds weight or weight gain ,

which was explained by a reduction in feed intake. This may be due to the antimicrobial compounds concentration, composition of diet and environment of the experiment. There are a few studies on this subject.

CONCLUSION AND RECOMMENDATIONS

Conclusion:

Supersan administration via drinking water did not affect growth performance and carcass traits. The dose recommended by Manufactory Company did not enough under our condition.

Recommendations:

- Practical implications

- The result of the present study showed that the commercial (Supersan soluble) can be used in drinking water in open system at summer season as heat stress control; otherwise it is better to increase the level of inclusion of Supersan soluble in drinking water.

- Suggestion for future study

- Further research is needed to get better understand about the effect of commercial (Supersan soluble) in improving the performance, carcass characteristics and immune of broiler chicks.

References

- Akande**, K.E.; Doma, U.D.; Agu, H.O. and Adamu, H.M. (2010). Major anti nutrients found in plant protein sources: their effect on nutrition. *Pakistan J. Nutr.*, 9 (8): 827-832.
- Alcicek**, A.; Bozkurt, M. and Cabuk, M. (2003). The effect of an essential oil combination derived from selected herbs growing wild in Turkey on broiler performance. *S. Afr. J. Anim. Sci.*, 33: 89-94.
- Andreatti Filho**, R.L. and Silva, E.N. (2005). Probióticos correlatos na produção avícola. In: Palermo Neto J, Spinosa HS, Górnaiak SL. *Farmacologia aplicada à avicultura*. São Paulo: Roca; Cap.15, pp:225-248.
- Ao**, T.; Cantor, A.H.; Pescatore, A.J.; Ford, M.J.; Pierce, J.L. and Dawson, K.A. (2009). Effect of enzyme supplementation and acidification of diets on nutrient digestibility and growth performance of broiler chicks. *Poult. Sci.*, 88 (1): 111-117.
- Apata**, D.F. (2008). Growth performance, nutrient digestibility and response of broiler chicks fed diets supplemented with a cultuer of *Lacotbacillus bulgaricus*. *J. Sci. food Agri.*, 88: 1253 – 1258.
- Atapattu** N.S.B.M. and Nelligaswatta C.J. (2005). Effects of citric acid on the performance and the utilization of phosphorous and crude protein in broiler chickens fed on rice by products based diets. *Int. J. Poult. Sci.* 4(12), 990-993.
- Awad** , W.A.; Ghareeb, K; Abdel – Raheem, S. and Bohm,. (2009). Effects of dietary unclusion of probiotic and symbiotic on growth performance, organ weights and weights and intestinal histomorphology of broiler chickens – *Poult. Sc.*, 88: 49 – 56.

- Bach Knudsen, K.E.** (2001). Development of antibiotic resistance and options to replace antimicrobials in animal diets. *Proc. Nutr. Soc.*, 60: 291-299.
- Baurhoo, B.; Ruiz-Feria, C.A. and Zhao, X.** (2008). Purified lignin: nutritional and health impacts on farm animals – a review. *Anim. Feed Sci. Tech.*, 144: 175-184.
- Beisel, W.R.** (1982). Single nutrients and immunity. *Am. J. Clin. Nutr.*, 35: 442-451.
- Bogusławska-Tryk, M.; Piotrowska, A. and Burlikowska, K.** (2012). Dietary fructans and their potential beneficial influence on health and performance parameters in broiler chickens. *J. Centr. Eur. Agr.*, 13: 272-291.
- Boling, S.D.; Webel, D.M.; Marvormichelis, I.; Parsons, C.M. and Baker, D.H.** (2000). The effects of citric acid on phytate phosphorus utilization in young chicks and pigs. *J. Anim. Sci.*, 78, 682-689.
- Boling-Frankenbach, S.D., Snow, J.L.; Parsons, C.M. and Baker, D.H.** (2001). The effects of citric acid on the calcium and phosphorus requirements of chicks fed corn-soybean meal diets. *Poult. Sci.*, 80, 783-78.
- Botsoglou, N.A.; Christaki, E.; Florou, P.; Giannenas, I.; Papageorgiou, G. and Spais, A.B.** (2004). The effect of mixture of herbal essential oils or alpha-tocopheryl acetate on performance parameters and oxidation of body lipids in broiler. *South Africa Journal Animal Science*, 34: 52-61.
- Cao, B.H., Zhang, X.P., Guo, Y.M.; Karasawa, Y. and Kumao, T.** (2003). Effects of dietary cellulose levels on growth, nitrogen utilization, retention time of diets in digestive tract and caecal microflora of chickens. *Asian-Australian J. Anim. Sci.*, 16: 863-866. *Sci*, 15: 15-20.

- Castanon, J.I.R.** (2007). History of the use of antibiotic as growth promoters in European poultry feed. *Poult. Sci.*, 86: 2466-2471.
- Chowdhury, R.;** Islam, K.M.; Khan, M.J.; Karim, M.R.; Haque, M.N.; Khatun, M. and Pesti, G.M. (2009). Effect of citric acid, avilamycin, and their combination on the performance, tibia ash, and immune status of broilers. *Poult. Sci.*, 88: 1616-1622.
- Christmas, R.B.;** Hars, R.H. and Sloan, D.R. (1995). The absence of vitamins and trace mineral and broiler performance. *Journal of applied Poultry Research*, 4: 407- 410.
- Ciftic, M.;** Guler, T.; Dalkilic, B. and Ertas, on (2005). The effect anise oil (*Pimpinella Aanisum 1.*) on broiler performance. *Inter. J. of poult. Sci.*, 4 (11): 851-855.
- Cmiljanic, R.;** Sretenovic, L.J.; Trenkovski, S. and Marinkov, G. (2001). Systems of poultry nutrition and their effect on production traits and quality of product. *Biotechnology in Animal Husbandry*, 17, 5-6, 179-185.
- Connelly, P.** (2011). Nutritional advantages and disadvantages of dietary phytates. *J. Australian Trad. Med. Soci.*, 17(1): 16-20.
- Cowan, M.M.** (1999). Plant products as antimicrobial agents. *Clin. Microb. Rev.*, 12: 564-582.
- Cowieson, A.J.;** Acamovic, T. and Bedford, M.R. (2000): Enzyme supplementation of diets containing *Camelina sativa* meal for poultry. *British Poultry Science*, 41: 689-690.
- Coxam V CNOV**(2007). Current data with inulin type fructans and calcium targeting bone health in adults. *J. Nutr.*, 137 (11 suppl) 25275-25335- PIMD 17951497.
- Craig, W. J.** (1999). Health promoting of common herbs AM,J, *Clin , Nutr.*, 70 Suppl:491- 499.

- Cross, D.E.; McDevitt, R.M.; Hillman, K. and Acamovic, T. (2007).** The effect of herbs and their associated essential oils on performance, dietary, digestibility and gut microflora in chickens from 7 to 28 days of age. *British Poultry Science*, 496-506.
- Deans, S.G. and Waterman, P.G. (1993).** Biological activity of volatile oils. Page 97-111. in *Volatile oil crops*, Hay, R.K.M. and Waterman, P.G, Eds . Longman Scientific and Technical, Essex.
- Dibner, J.J. and Richards, J.D. (2005).** Antibiotic growth promoters in agriculture: History and mode of action. *Poult. Sci.*, 84: 634–643.
- Diplock, A.T. (1994).** Antioxidants and disease prevention. *Mol. Asp. Med.*, 15:295 –376.
- Donkoh, A. (1989).** Ambient temperature: a factor affecting performance and physiological response of broiler chickens. *Int. J. Biometerol.*, 33:259-265.
- Dorman, H.J.D. and Deans, S.G. (2004).** Antimicrobial agent from plant volatile oils. *J. Appl. Microbial.*, 88: 308-316.
- Dragland, S.; Senooh , ; Wake, k. and Blomhoh, R. (2003).** Several culinary and medicinal herbs are important sources of dietary antioxidants. *J. Nutr.*, 133: 1286-1290.
- Duke , J.A (1986).** CRC handbook of medicinal herbs. CRC press, Florida.
- Edwards, H.M.; Shirley, R.B.; Escoe, W.B. and Pesti, G.M. (2002).** Quantitative evaluation of 1- α -hydroxycholecalciferol as a cholecalciferol substitute for broilers. *Poult. Sci.*, 81:664–669.
- Ersin samli, H.;Nizamettin, S.; Hasan, A. and Aylin, A. (2006).** Using rise bran in laying hen diets. *J. Cen. Euro. Agric.*, 7 (1): 135-140.
- Faleiro, M.L.; Miguel, M.G.; Ladeiro, F.; Venancio, F.; Taveres, R.; Brito, J.C.; Figueiredo, A.C.; Barroso, J.G. and Pedro, L.G. (2003).**

Antimicrobial activity of essential oils isolated from Portuguese endemic species of *Thymus*. *Lett. Appl. Microbiol.*, 36: 35-40.

- Farnell, M.B.**; donoghue, A.M.; De Los; Santos, F.S.; Bloer, P.J.; Hargis, B.M.; Tellez, G. and Donoghue, D.J. (2006). Upregulation of oxidative burst and degranulation in chicken heterophils stimulated with probiotic bacteria. *Poultry Sci.*, 85: 1900- 1906 with probiotic in man and animal J. Fuller, R. (1989). Probiotic in man and animal. *J. Appl. Bacteriol.*, 66: 365 – 378.
- Fuller, R.** (1989). Probiotics in man and animals. *Journal of Applied Bacteriology*, 66: 365-378.
- Gallinger, C.I.**; Suarez, D.M. and Irazusta, A. (2004). Effects of rice bran inclusion on performance and bone mineralization in broiler chicks. *J. Appl. Poult. Res.*, 13: 183-190.
- Gibson, G.R.** and Roberfroid, M.B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *Journal of Nutrition*, 125: 1401-1412.
- Gil De Los Santos, J.R.**; Storch, O.B. and Gil – turns, C. (2005). *Bacillus Cereus* Var. *Toyoi* and *Saccharomyces boulardii* increased feed efficiency in broiler infected with *Salmonella Enteritidis*. *Br. Poult. Sci.*, 46: 494 – 497.
- Havenaar, R.** and Spanhaak, S. (1994). Probiotics from an immunological point of View. *Curr. Opin Biotech.*, 5: 320–325.
- Hay, R.K.M.** and Waterman, P.G. (1993). Volatile oil crops: their biology, biochemistry and production. Longman Scientific and Technical, Essex.
- Hernández, F.**; Madrid, J.; Garcia, V.; Orengo, J.; and Megias, M.D. (2004). Influence of two plant extracts on broilers performance, digestibility, and digestive organ size. *Poult. Sci.*, 83: 169–174.

- Hertrampf, J.W.** (2001). Alternative antibacterial performance promoters. *Poult. Int.*, 40: 50-52.
- Huang, M.K.; Choi, Y.J.; Houde, R.; Lec, J.W. and Zho, X.** (2004). effects of lacto bacilli and acidophitic, fungus on the production performance and immune responses in broiler. *Poult. Sc.*, 83: 788- 795.
- Humphrey, B.D.; Huang, N. and Klasing, K.C.** (2002). Rice expressing lactoferrin and lysozyme has antibiotic-like properties when fed to chicks. *J. Nutr.*, 132: 1214-1218.
- Hurwitz, S.; Weiselbrg, M.; Eisner, U.; Bartov, I.; Riesenfeld, G.; Sharvit, M.; Niv, A. and Bornstens, S.** (1980). The energy requirements and performance of growing chickens and turkeys, as affected by environmental temperature. *Poultry Sci.*, 59: 2290-2299.
- Islam, K.M.S.; Schaeublin, H.; Wenk, C.; Wanner, M. and Liesegang, A.** (2012). Effect of dietary citric acid on the performance and mineral metabolism of broiler. *J. Anim. Physiol. Anim. Nutr.*, 96(5): 808-817.
- Jamroz, D.; Wiliczekiewicz, A.; Wertelecki, T.; Orda, J. and Skorupinska, J.** (2005). Use of active substances of plant origin in chicken diets based on maize and locally grown cereals. *Br. Poult. Sci.*, 46: 485-493.
- Jantan, I.B.; Yassin, M.S.M.; Chin, C.B.; Chen, L.L. and Sim, N.L.** (2003). Anti Lungal activity of essential oils of nine Zingiberaceae species. *Journal British Poultry Science*, 41(5): 392-397.
- Janssen, A.M.; Scheffer, J.J.C. and Svendsen, A.B.** (1987). Antimicrobial activities of essential oils. *Pharmaceutical Weekblad (Sci.)*, 9: 193-197.
- Javed ,M. ; Durrani, F.R.; Hafeez , A.; Khan, R.U. and Ahmed, I.** (2009). Effect of aquesous extract of plant mixture on carcass quality of

broiler chicks. *Arpan Journal of Agriculture and biological Science*, 4 (1): 37- 40.

Jin, L.; HO, Y.W.; Abdullah, N. and Jajaludin, S. (2000). Digestive and bacteria enzyme activities in broiler diets supplemented with lacto bacillus culture. *Polt. Sc.*,

Jin, L.Z.; Ho, Y.M.; Abdullah, S. and Jalaludin, S. (1997) . Probiotic in poultry: modes of action. *World's Poultry Science Journal*, 53: 351-368.

Jozefiak, D.; Kaczmarek, S.; Bochenek, M. and Rutkowski, A. (2007). A note on effect of benzoic acid supplementation on the performance and microbiota population of broiler chickens. *J. Anim. Feed Sci.*, 16: 252-256.

Kabir, M.L.; Rahman, M.M.; Rahman, B.M., and Ahmed, U.S. (2004). The dynamics of probiotics on growth performance and immune response in broilers. *Int. J. Poult. Sc.*, 3: 361 -364.

Kalavathy, R.; Abdullah, N.; Jalaludin, S. and HO, Y.W. (2003). Effects lacto bacillus cultures on growth performance abdominal fat deposition, serum lipids and weight of organs of broiler chickens. *Br. Poult. Sc.*, 44: 139-144.

Kamel, C. (2001). Tracing modes of action and the roles of plant extracts in non-Numinants in: Gams wor thy pc, and Wiseman. J. (editor) recent advance in animal nutrition, Nottingham University Press. 133-150.

Kasim, A.B. and Edwards, H.M. (2000). Evaluation of cholecalciferol sources using broiler chick bioassays. *Poult. Sci.*, 79:1617–1622.

Kim, G.B.; Seo, Y.M.; Kim, C.H. and Paik, I.K. (2011). Effect of dietary prebiotics supplementation on the performance, intestinal microflora, and immune response of broilers. *Poult. Sci.*, 90: 75-82.

- Klasing**, K.C. (1998). *Comparative Avian Nutrition*. University Press, Cambridge, 277- 299.
- Kluge**, H.; Broz, J. and Eder, K. (2006). Effect of benzoic acid on growth performance, nutrient digestibility, nitrogen balance, gastrointestinal microflora and parameters of microbial metabolism in piglets. *J. Anim. Physiol. Anim. Nutr. (Berl.)*, 90: 316-324.
- Koenen**, M.E.; Kramer, J.; Van Der Hulst, R.; Heres, L.; Jeurissen, S.H. and Boersma, W.J.A. (2004). Immunomodulation by probiotic lactobacilli in layer- and meat-type chickens. *Br. Poult. Sci.*, 45: 355- 366.
- Kralik**, G.; Milakovic, Z. and Ivankovic, S. (2004). Effect of probiotic supplementation on the performance and intestinal microflora of broilers. *Acta. Agri. Kapo.*, 8: 23- 31.
- Krás**, R.V.; Kessler, A.M.; Ribeiro, A.M.L.; Henn, J.D.; Il dos Santos; Halfen, D.P. and Bockor, L. (2013) . Effect of dietary fiber and genetic strain on the performance and energy balance of broiler chickens. *Braz. J. Poult.*,
- Langhout**, P. (2000). New additives for broiler chickens. *World Poultry-Elsevier*, 16 (3): 22-27.
- Lawrence**, B.M. and Reynolds, R.J. (1984). Progress in essential oils. *Perfumer and Flavorist*, 9: 23- 31.
- Lee**, K.W.; Everts, H.; Kappert, H.J.; Frehner, M.; Losa, R. and Beynen, A.C. (2003). Effects of dietary essential oil components on growth performance, digestive enzymes and lipid metabolism in female broiler chickens. *Br. Poult. Sci.*, 44: 450–457.
- Leeson**, S. and Summers, J. (2001). *Nutrition of the chicken* (4th edition) Published by University Books. P.O. Box. 1326, Guelph, Ontario, Canada.

- Liem, A.;** Pesti, G.M. and Edwards, H.M. (2008). The effects of several organic acids on phytate phosphorus hydrolysis in broiler chicks. *Poult. Sci.*, 87: 689-693.
- Lodhi, G.N.;** Singh, D. and Khopani, J.S. (1976). Metabolizable energy values for poultry. *J. Agric. Sci.*, 86: 293-303.
- McDowell, L.R.** (1989). *Vitamins in Animal Nutrition comparative Aspects to Human Nutrition . Vitamin A and E.* Academic Press, London, 10 (52) :93-131.
- Mellor, S.** (2000). Nutraceuticals—alternatives to antibiotics. *World Poultry-Elsevier*, 16(2): 30-33.
- Meng, X.;** Slominski, B.A.; Nyachoti, C.M.; Campbell, L.D. and Guenter, W. (2005). Degradation of cell wall polysaccharides by combinations of carbohydrates enzymes and their effect on nutrient utilization and broiler chicken performance. *Poultry Science*, 84 (1): 37-47.
- Mitsch, P.;** Zitterl-Eglseer, K.; Kohler, B.; Gabler, C.; Losa, R. and Zimpernik, I. (2004). The effect of two different blends of essential oil components on the proliferation of *Clostridium perfringens* in the intestines of broiler chickens. *Poult. Sci.*, 83: 669–675.
- Mohammed, K.A.;**Toson, M.A.; Hassanien, H.H.M.; Soliman, M.A.H. and Sanaa, H.N. (2010). Effects of phytase supplementation of performance and egg quality of laying hens fed diets containing rice bran. *Egyptian Poult. Sci.*, 30: 649-659.
- Mohiti Asli, M.;** Hosseini, S.A.; Lotfollahian, H. and Shariatmadari, F. (2007). Effect of probiotics, yeast, vitamin E and vitamin C supplements on performance and immune response of laying hen during high environmental temperature. *Inter. J. Poult. Sci.*, 6 (12): 895-900.

- Mountzouris, K. C.; Tsistsikos, P.; Kalamara, E.; Nitsch, S.; Schatzmayr, G. and fegeros, K. (2007).** Evaluation of the efficacy of a probiotics containing *Lactobacillus*, *Bidofidobaslerium* , *Enterococcus* and *Pediococcus* strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. *Poult. Sc.*, 86: 309 – 317.
- Mountzouris, K. C.; Balaskas, C.I.; Xanthakos, A. Tzivinikou, and Fegeros, K. (2009).** Effects a multispecies probiotic on biomarkers of competitive exclusion effecacy in broilers challenged with *Salmonella Enteritidis*. *Br. Poult. Sci.*, 50: 467 – 478.
- Mujahid, A.; Ikram, U.H.; Musaddiq, A. and Abrar, H.G. (2005).** Effect of various processing techniques and different levels of antioxidant on stability of rice bran during storage. *J. Sci. Food Agric.*, 85: 847-852.
- Mukhtar, M.A.; Makkawi, A. and Tigani, M. (2010a).** Effect of amino acids supplementation to Marginally Deficient local Broiler Chick diets. *Journal of Science and Technology*, 11 (2): 80-82.
- Mukhtar, M.A.; Mohammed, K.A and Musa, M. H. (2010b).** Replacement Value of Lysine and Methionine for Super Concentrate in Broiler Chick's Yield and Quality. *Journal of Science and Technology*, 11 (2): 27-29.
- Nabizadeh, A. (2012).** The effect of inulin on broiler chicken intestinal microflora, gut morphology, and performance. *J. Anim. Feed Sci.*, 21: 725-734.
- Nahanshon, S.N.; Nakaue, H.S. and Mirosh, L.W. (1993).** Effect of direct fed microbials on nutrient retention and parameter of Single White Leghorn pullets. *Poult. Sci.*, 72: 87.
- Nava, G.M.; Attene-Ramos, M.S.; Gaskins, H.R. and Richards, J.D. (2009).** Molecular analysis of microbial community structure in the

chicken ileum following organic acid supplementation. *Vet. Microbiol.*, 137: 345-353.

Naziroğlu, M.; Sahin, k.; Simsek, H.; Aydilek, N. and Ertas, O.N. (2000). The effect of food withdrawal and darkening on lipid peroxidation of laying hens in high ambient temperatures *Dtsch. Tierarztl. Wschr.*, 107: 199-202.

Niewold, T.A. (2007). The nonantibiotic anti-inflammatory effect of antimicrobial growth promoters, the real mode of action? A hypothesis. *Poult. Sci.*, 86: 605–609.

Niki, E. (1996). E∞-Tocopherol, In: cadenas E. and packer L. eds. handbook of antioxidant.; 3–25. mercel dekker.

NRC, National Research Council. (1994). Nutrient Requirement of Poultry; 9th ed., National Academy Press, Washington, D.C., USA.

Owings, W.J.D.l.; Reyenolda, R.J.; Hasia, K. and Ferket, P.R. (1990). Influence of dietary supplementation with *sterptococcus faccium* M-74 on broiler body weight, feed conversion, carcass characteristics and intestinal microbial colonization. *Poult. Sc.*, 69: 1257-1264.

Oyen, L.P.A. and Dung, N.X. (1999). Essentail-oil plants. Oyen , L.P.A. and Dung, N.X. (Eds). Backhuys Publishers, Leiden.

Parks , C.W. F.; Grimers, J.I.; Freket, P.R. and Fairchild, A.S. (2000). The case for manna oligosaccharides in poultry diets – An alternate top growth promoter antibiotics – In: Bio technology in the feed industry Ed Lyons and Jacques. Pul Altech. Inc Ky.

Park, S.O. and Park, B.S. (2011). Effect of dietary micro encapsulated inulin carcass characteristics and growth performance in broiler chickens. *Journal of Animal and Veterinary Advances*, 10:1342-1412.

- Patterson, J.A.** and Burkolder, K.M. (2003). Application of prebiotics and probiotics in poultry production. *Poultry Sci.*, 82: 627- 631.
- Paul, R.C.;** Ahmad, N.; Moinuddin, M.A. and Hasan, N. (2010). Effects of administration of multivitamins and enzymes for broilers either singly or in combination on body weight and Haematobiochemical parameters. *J. Bangladesh Agril. Univ.*, 8 (1): 39-44.
- Peebles, E.D.;** Zumwalt, C.D.; Doyle, S.M.; Gerard, P.D.; Latour, M.A, Boyle, C.R. and Smith, T.W. (2000a). Effects of breeder age and dietary fat sources and level on broiler hatching egg characteristics. *Poult. sci.*, 79: 698-704.
- Peebles, E.D.;** Zumwalt, C.D.; Doyle, S.M.; Gerard, P.D.; Latour, M.A, Boyle, C.R. and Smith, T.W. (2000b). Effects of dietary fat type and level on broiler breeder performance. *Poult. Sci.*, 79: 629-639.
- Peñalver, P.;** Huerta, B.; Borge, C.; Astorga, R.; Romero, R. and Perea, A. (2005). Antimicrobial activity of five essential oils against origin strains of the Enterobacteriaceae family. *APMIS*, 113: 1–6.
- Piyaratne, M.K.D.K.;** Atapattu, N.S.B.M.; Mendis, A.P.S. and Amarasinghe, A.G.C. (2011). Effects of balancing rice bran based diets for up to four amino acids on growth performance of broilers. *Trop. Agric. Res. Ext.*, 12 (2): 57-61.
- Preston, C.M.;** Mckracken, K.J. and McAllister, A. (2000). Effect of the diet form and enzyme supplementation on growth, efficiency and energy utilization of wheat-based diets for broilers. *British Poultry Science*, 41: 324-31.
- Preuss, H.G.;** Echard, B.; Enig, M.; Brook, I. and Elliott, T.B. (2005). Minimum inhibitory concentrations of herbal essential oils and

monolaurin for gram-positive and gram-negative bacteria. *Mol. Cell. Biochem.*, 272: 29–34.

Puminn, O. (2003). Broiler performance and mineral utilization of enzyme supplemented defatted rice bran diets during heat stress. PhD. Thesis. Univ., Tennessee, Knoxville. 388.

Puron, D.; Santamaria, P. and Segura, J.C. (1994). Effect of Sodium Bicarbonate, Acetylsalicylic and Ascorbic acid on broiler performance in a tropical environment. *Journal of Applied Poultry Research*, 3: 141-145.

Puthongsiriporn, U.; Scheideler, S.E.; Shell, J.L. and Beck, M.M. (2001). Effects of Vitamin E and C Supplementation on Performance, In Vitro Lymphocyte Proliferation, and Antioxidant Status of Laying Hens during Heat Stress¹ *Poult Sci*, 80: 1190-1200.

Rafacz-livingstan, K.A.; Marteninez-amezcua, C.; Parsons, C.M.; Baker, D.H. and Snow, J. (2005). Citric acid improves phytate phosphorous utilization in cross breed and commercial broiler chicks. *Poult. Sci.*, 84: 1370-1375.

Ravindran, V. (1995). Phytases in poultry nutrition. An overview. pp. 135-139 in *Proc. Australian Poult. Sci. Symp.*, Sydney, Australia.

Rebole, A.; Ortiz, I.T.; Rodriguez M.L.; Alzueta, G.; Trevino, J. and Ve Lasco, S. (2010). Effect of inulin and enzyme complex, individually or in combination, on growth performance, intestinal microflora, and cecal fermentation characteristics and jejunal histomorphology in broiler chickens fed a wheat – and barely – based diet . Departamento de Production Animal , Facultad de Veterinaria , Universidad complutense de Madrid . 28040, Spain. Accepted Jan. 2008 Poultry Science Association Inc.

- Richter, G.; Cyriaci, G. and Stoken, B. (1994).** Effect of enzyme supplementation in barley, rye or triticale based broiler diets. *Journal of Animal Nutrition*, 47 (1): 11-22.
- Riebau, F.J.M.; Berger, M.B.; Yegen, O. and Cakır, C. (1997).** Seasonal variations in the chemical compositions of essential oils of selected aromatic plants growing wild in Turkey. *J. Agric. Food Chem.*, 45: 4821-4825.
- Rotruck, J.T., Pope, A.L.; Ganther, H.E.; Swanson, A.B.; Hafeman, D.G. and Hoekstra, W.G. (1973).** Selenium: biochemical roles a component of glutathione peroxidase. *J. Poult. Sci.*; 179:588-590.
- Saki, A.A.; Hemati Matin, H.R.; Tabatabai, M.M.; Zamani, P. and Naseri Harsini, R. (2010).** Microflora population, intestinal condition and performance of broilers in response to various rates of pectin and cellulose in the diet. *Arch Geflügelk*, 74: 183-188.
- Saleh, F.; Tahir, M.; Ohtsuka, A. and Hayashi, K. (2005).** A mixture of pure cellulase, hemicellulase and pectinase improves broiler performance. *British Poultry Science*, 46: 602-6.
- Santoyo, S.; Cavero, S.; Jaime, L.; Ibanez, E.; Senorans, F.J. and Reglero, G. (2005).** Chemical composition and antimicrobial activity of *Rosmarinus officinalis*L. essential oil obtained via supercritical fluid extraction. *J. Food Prot.*, 68: 790–795.
- Schilcher, H. (1985)** Effects and side –effects of essential oils pages 217 – 231 in : *Proceedings of the 15th international symposium on essential oils*. Noordwijkerhout, The Netherlands.
- Schwarz, S.; Kehrenberg, C. and Walsh, T.R. (2001).** Use of antimicrobial agents in veterinary medicine and food animal production. *Int. J. Antimicrob. Agents*, 17: 431-437.

- Selle, P.H.;** Ravindran, V.; Caldwell, R.A. and Bryden, W.L. (2007). Phytate and phytase: consequences for protein utilisation. *Nutr. Res. Rev.*, 13 (2): 255-278.
- Shakouri, M.D.;** Kermanshahi, H. and Mohsenzadeh, M. (2006). Effect of different non starch polysaccharides in semi purified diets on performance and intestinal microflora of young broiler chickens. *Int. J. Poult. Sci.*, 5: 557-561.
- Sharif, M.K.** (2009). Rice industrial by products management for oil extraction and value added products. PhD. Thesis. Agriculture Univ., Faisalabad.
- Sahin, K.;** Ertas, O.N. and Guler, T. (1999). Sicaklik stresi altindaki yumurta tavuklarinda farkli yemleme yontemlerinin vitamin A, vitamin E ile bazi kan parametreleri uzerine etkileri. In: Serbest Radikaller ve Antioksidanlar Arastrima. Dernegi II, Ulusal Kongresi. P:24-58.
- Shunying, Z.;** Yang, Y.; Huaidong, Y.; Yue, Y. and Guolin, Z. (2005). Chemical composition and antimicrobial activity of the essential oils of *Chrysanthemum indicum*. *J. Ethnopharmacol.*, 96: 151–158.
- Siddique, M.** (2004). Role of vitamins in immune response in poultry. Department of Veterinary Microbiology. University of Agriculture, Faisalabad. Pakissan. Com., pp: 1-3.
- Silva, E.N.** (2000). Proboticos e preboticos na alimentacao de aves. In: Conferência Apinco de ciência e tecnologia avícolas. Campinas, Cao Paulo. Brasil. v. 2, p. 241-251.
- Singsen, E.P.** (1947). Nutritional factors influencing growth and efficiency of feed utilization in growing chick. *Poultry Science*, 26:555-557.

- Snow, J.L.;** Baker, D.H. and Parsons, C.M. (2004). Phytase, citric acid and 1 α -hydroxycholecalciferol improve phytate phosphorus utilization in chicks fed a corn soybean meal diet. *Poult. Sci.*, 83: 1187-1192.
- Soliman, K.M. and Badea, R.I.** (2002). Effect of oil extracted from some medical plants on different mycotoxigenic fungi food chem. 40:1669-75.
- Sreenivasaiah, P.V.** (2006). Scientific poultry production. Third edition. International Book Distributing CO. (publishing division). Chaman Studio building, 2nd floor, Charbagh, Lucknow 226004U.P. India.
- Srinivasan, K.** (2007). Black pepper and its pungent principle-piperine: A review of diverse physiological effects. *Crit. Rev. Food Sci. Nutr.*, 47: 735–748.
- Surai, P.F.** (1999). Vitamin E in avian reproduction. *Poult. Avian Biol. Rev.*, 10:1-60.
- Surai, P.F.** (2000). Organic selenium: benefits to animals and humans, a biochemist's view In: *Biotechnology in the Feed Industry. Proc. of the 16th annual Symposium* (ed. Lyons, T.P. and Jacques, K. A.). Nottingham University Press, Nottingham, UK, 205-260.
- Surai, P.F.;** Fujihara, N.; Speake, P.K.; Pbrillard, J.; Wishart, G.J. and Spark, N.H.C. (2001). Polyunsaturated Fatty Acids, Lipid Peroxidation and Antioxidant Protection in Avian Semen. *Asian – Australian J. Anim. Sci.*, 14 (7):1024-1050.
- Svihus, B. and Hetland, H.** (2001). Ileal starch digestibility in growing broiler chickens fed on a wheat-based diet is improved by mash feeding, dilution with cellulose or whole wheat inclusion. *Br. Poult. Sci.*, 42: 633-637.

- Teo, A.Y.** and Tan, H.M. (2007). Evaluation of the performance and intestinal gut microflora of the performance and intestinal gut microflora of broilers fed on corn – soy diets supplemented with *Bacillus subtilis* PB6(CloSTAT). *J. Appl. Poult. Res.*, 16: 296-303.
- Timmerman, H.M.A .;** Veldman, E.; Van Den Eken, F.; Rombouts, M and Beyrun, A.C. (2005). Mortality and growth performance of broiling given drinking water supplemented with chicken – specific prebiotics. *Poult. Sc.* 85: 1383-1388.
- Tortucro, E.** (1978). Influence of the implantation of *lactobacillus acidophilus* in chicks on the growth, feed conversion, Mal absorption of fats syndrome and intestinal floro. *Poult. Sc.*, 52: 197- 203 .
- Ultee, A.;** Bennik, H.J. and Moezelaar, R. (2002). The phenolic hydroxyl group of carvacrol is essential for action against the food-borne pathogen, *Bacillus cereus*. *Appl. Environ. Microbiol.*, 3: 1561-1568.
- Wang, Z.R.;** Qiao, S.Y.; Lu, W.Q. and Li, D.F. (2005). Effect of enzyme supplementation on performance, nutrient digestibility of broilers fed wheat-based diets. *Poultry Science*, 84: 875-81.
- Ward, N.E.** (1996). Commercial vitamin supplementation for poultry. *Poultry Adviser*, New Jerrey, USA, 29(3): 29-50.
- Wenk, C.** (2000). Recent advances in animal feed additives such as metabolic modifiers, antimicrobial agents, probiotics, enzymes and highly available minerals. Review. *Asian-Aus. J. Anim. Sci.*, 13: 86-95.
- Ziaei, N.;** Kor, N.M. and Pour, E.E. (2013). The effects of different levels of vitamin-E and organic selenium on performance and immune

response of laying hens. African Journal of Biotechnology, 12 (24): 3884-3890.

Zulkifli, I; Abdullah, N.; Azrin, N.M. and HO, Y.W. (2000). Growth performance and immune response of two commercial broiler strains fed diets containing lacto bacillus culture and oxytetracycline under heat stress conditions. British Poultry Science, 41: 593- 597.

Abbreviations:

IgG = immunoglobulin G

IgA = immunoglobulin A

EO = essential oils

CA= citric acid

ACTH= adrenocorticotropin hormone

GSH= glutathione

GSH-Px= Glutathione peroxidase

Se= selenium

NPS= non-starch polysaccharides

IBD= Infectious Bursal Disease

CRINA =

DSM= is a global Life Sciences and Materials Sciences company active in health, nutrition and materials.

Bifidobacterium = A genus of Gram-positive, nonmotile, anaerobic bacteria, some of them used as probiotics.

Appendix(1):

COMPOSITION OF SUPERSAN SOLUBLE :

king water.

Vitamin A600,000 IU
Vitamin D3	50,000 IU
Vitamin E	100 mg
Vitamin B1	100 mg
Vitamin B2	100 mg
Vitamin B6	200 mg
Vitamin B12	1.5 mg
Vitamin K	200 mg
Calcium pantothenate	500 mg
Nicotinic acid	1 g
Protein hydrolysate	10 g
Excipient to	100 ml

Appendix (2):

Weekly medium air temperature during the period 7th April to 27th may 2012

Weeks	medium temperature °C
1	33.4
2	32
3	31
4	32.3
5	30.8
6	31.3
7	34.3

Source: thermometer

Appendix (3):



Appendix (4):

